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(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICII. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.

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#### TRANSPORTERS AND ION CHANNELS

This invention relates to nucleic acid and amino acid sequences of transporters and ion channels and to the use of these sequences in the diagnosis, treatment, and prevention of transport, 5 neurological, muscle, immunological and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

#### **BACKGROUND OF THE INVENTION**

Eukaryotic cells are surrounded and subdivided into functionally distinct organelles by hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K<sup>+</sup>, NH<sub>4</sub><sup>+</sup>, P<sub>i</sub>, SO<sub>4</sub><sup>2</sup>, sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic 15 neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational 20 change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with 25 simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the 30 ubiquitous Na<sup>+</sup>/K<sup>+</sup> ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide 35 thyroid-imaging techniques and for specific targeting of radioisotopes to the thyroid gland (Levy,

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O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

One of the largest families of transporters is the major facilitator superfamily (MFS), also called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. 10 Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 15 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulin-regulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, 20 M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H<sup>+</sup>-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H<sup>+</sup>-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K<sub>m</sub> values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na<sup>+</sup>-monocarboxylate cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions in organs including the

kidney, intestine and liver. Organic anion transporters are selective for hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) Am. J. Physiol. 264:C761-C782; Price, N.T. et al. (1998) Biochem. J. 329:321-328; and Martinelle, K. and I. Haggstrom (1993) J. Biotechnol. 30:339-350).

ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC 10 transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "half-15 molecules" form homo- and heterodimers, such as Tapl and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic 20 hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) Meth. Enzymol. 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum,
selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example,
copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development,
by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase
(ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and
are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to
the liver and other target organs, where specific transporters move the ions into cells and cellular
organelles as needed. Imbalances in metal ion metabolism have been associated with a number of
disease states (Danks, D.M. (1986) J. Med. Genet. 23:99-106).

P-type ATPases comprise a class of cation-transporting transmembrane proteins. They are integral membrane proteins which use an aspartyl phosphate intermediate to move cations across a membrane. Features of P-type ATPases include: (i) a cation channel; (ii) a stalk, formed by

extensions of the transmembrane α-helices into the cytoplasm; (iii) an ATP binding domain; (iv) a phosphorylated aspartic acid; (v) an adjacent transduction domain; (vi) a phosphatase domain, which removes the phosphate from the aspartic acid as part of the reaction cycle; and (vii) six or more transmembrane domains. Included in this class are heavy metal-transporting ATPases as well as aminophospholipid transporters.

The transport of phosphatidylserine and phosphatidylethanolamine by aminophospholipid translocase results in the movement of these molecules from one side of a bilayer to another. This transport is conducted by a newly identified subfamily of P-type ATPases which are proposed to be amphipath transporters. Amphipath transporters move molecules having both a hydrophilic and a hydrophobic region. As many as seventeen different genes belong to this P-type ATPases subfamily, being grouped into several distinct classes and subclasses (Halleck, M.S. et al., (1999) Physiol. Genomics 1:139-150; Vulpe, C. Et al., (1993) Nat. Genet. 3:7-13).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant 15 fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process. Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

The lipocalin superfamily constitutes a phylogenetically conserved group of more than forty proteins that function as extracellular ligand-binding proteins which bind and transport small hydrophobic molecules. Members of this family function as carriers of retinoids, odorants, chromophores, pheromones, allergens, and sterols, and in a variety of processes including nutrient transport, cell growth regulation, immune response, and prostaglandin synthesis. A subset of these proteins may be multifunctional, serving as either a biosynthetic enzyme or as a specific enzyme inhibitor. (Tanaka, T. et al. (1997) J. Biol. Chem. 272:15789-15795; and van't Hof, W. et al. (1997) J. Biol. Chem. 272:1837-1841.)

Members of the lipocalin family display unusually low levels of overall sequence conservation. Pairwise sequence identity often falls below 20%. Sequence similarity between 30 family members is limited to conserved cysteines which form disulfide bonds and three motifs which form a juxtaposed cluster that functions as a target cell recognition site. The lipocalins share an eight stranded, anti-parallel beta-sheet which folds back on itself to form a continuously hydrogen-bonded beta-barrel. The pocket formed by the barrel functions as an internal ligand binding site. Seven loops (L1 to L7) form short beta-hairpins, except loop L1 which is a large

omega loop that forms a lid to partially close the internal ligand-binding site (Flower (1996) Biochem. J. 318:1-14).

Lipocalins are important transport molecules. Each lipocalin associates with a particular ligand and delivers that ligand to appropriate target sites within the organism. Retinol-binding 5 protein (RBP), one of the best characterized lipocalins, transports retinol from stores within the liver to target tissues. Apolipoprotein D (apo D), a component of high density lipoproteins (HDLs) and low density lipoproteins (LDLs), functions in the targeted collection and delivery of cholesterol throughout the body. Lipocalins are also involved in cell regulatory processes. Apo D, which is identical to gross-cystic-disease-fluid protein (GCDFP)-24, is a progesterone/pregnenolone-binding protein expressed at high levels in breast cyst fluid. Secretion of apo D in certain human breast cancer cell lines is accompanied by reduced cell proliferation and progression of cells to a more differentiated phenotype. Similarly, apo D and another lipocalin, α<sub>1</sub>-acid glycoprotein (AGP), are involved in nerve cell regeneration. AGP is also involved in anti-inflammatory and immunosuppressive activities. AGP is one of the positive acute-phase proteins (APP); circulating 15 levels of AGP increase in response to stress and inflammatory stimulation. AGP accumulates at sites of inflammation where it inhibits platelet and neutrophil activation and inhibits phagocytosis. The immunomodulatory properties of AGP are due to glycosylation. AGP is 40% carbohydrate, making it unusually acidic and soluble. The glycosylation pattern of AGP changes during acutephase response, and deglycosylated AGP has no immunosuppressive activity (Flower (1994) FEBS 20 Lett. 354:7-11; Flower (1996) supra).

The lipocalin superfamily also includes several animal allergens, including the mouse major urinary protein (mMUP), the rat α-2-microgloobulin (rA2U), the bovine β-lactoglobulin (βlg), the cockroach allergen (Bla g4), bovine dander allergen (Bos d2), and the major horse allergen, designated *Equus caballus* allergen 1 (Equ c1). Equ c1 is a powerful allergen responsible for about 80% of anti-horse IgE antibody response in patients who are chronically exposed to horse allergens. It appears that lipocalins may contain a common structure that is able to induce the IgE response (Gregoire, C. et al., (1996) J. Biol. Chem. 271:32951-32959).

Lipocalins are used as diagnostic and prognostic markers in a variety of disease states. The plasma level of AGP is monitored during pregnancy and in diagnosis and prognosis of conditions including cancer chemotherapy, renal disfunction, myocardial infarction, arthritis, and multiple sclerosis. RBP is used clinically as a marker of tubular reabsorption in the kidney, and apo D is a marker in gross cystic breast disease (Flower (1996) supra). Additionally, the use of lipocalin animal allergens may help in the diagnosis of allergic reactions to horses (Gregoire supra), pigs, cockroaches, mice and rats.

Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions

and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease carrier protein, a protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995) Biochemistry, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189 Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in Man (OMIM) \*275000 Graves Disease).

This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int. Med. 245:637-642).

#### **Ion Channels**

The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ion20 selective pores within the membrane. There are two basic types of ion channels, ion transporters and gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient.

These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion transporters, including Na<sup>+</sup>-K<sup>+</sup> ATPase, Ca<sup>2+</sup>-ATPase, and H<sup>+</sup>-ATPase, are activated by a phosphorylation event. P-class ion transporters, also known as E1-E2 type ATPases, are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na<sup>+</sup> and Ca<sup>2+</sup> are low and cytosolic concentration of K<sup>+</sup> is high. The vacuolar (V) class of ion transporters includes H<sup>+</sup> pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are

responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H<sup>+</sup> pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P<sub>i</sub>).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) Curr. Opin. Cell Biol. 11:517-522). The V-ATPases are composed of two functional domains: the V<sub>1</sub> domain, a peripheral complex responsible for ATP hydrolysis; and the V<sub>0</sub> domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and evolutionarily related to the V-ATPases. The F-ATPase F<sub>0</sub> domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V<sub>0</sub> domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) J. Biol. Chem. 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na<sup>+</sup> down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca<sup>2+</sup> out of the cell with transport of Na<sup>+</sup> into the cell (antiport).

#### **Gated Ion Channels**

Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, and Cl channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g., acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

35 Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of

touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca<sup>2+</sup> and Na<sup>+</sup> (Suzuki, M. et al. (1999) J. Biol. Chem. 274:6330-6335).

The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na<sup>+</sup> and Ca<sup>2+</sup> subfamilies, this domain is repeated four times, while in the K<sup>+</sup> channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K<sup>+</sup> channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

Voltage-gated Na<sup>+</sup> and K<sup>+</sup> channels are necessary for the function of electrically excitable

cells, such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na<sup>+</sup> and K<sup>+</sup> ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na<sup>+</sup> channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na<sup>+</sup> channels, which propagates the depolarization down the length of the cell.

Depolarization also opens voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the

channel's N-terminus, which acts as a plug that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

Voltage-gated Na<sup>+</sup> channels are heterotrimeric complexes composed of a 260 kDa pore-forming  $\alpha$  subunit that associates with two smaller auxiliary subunits,  $\beta 1$  and  $\beta 2$ . The  $\beta 2$  subunit is a integral membrane glycoprotein that contains an extracellular Ig domain, and its association with  $\alpha$  and  $\beta 1$  subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) Cell 83:433-442).

Non voltage-gated Na<sup>+</sup> channels include the members of the amiloride-sensitive Na<sup>+</sup> channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini

located within the cell. The NaC/DEG family includes the epithelial Na\* channel (ENaC) involved in Na\* reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized H\*-gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na\*-permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from C. elegans. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) Curr. Opin. Neurobiol. 8:418-424; Eglen, R.M. et al. (1999) Trends Pharmacol. Sci. 20:337-342).

K\* channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca<sup>2+</sup> and cAMP. In non-excitable tissue, K\* channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K\* channels are responsible for setting the resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na\*-K\* pump and ion channels that provide the redistribution of Na\*, K\*, and Cl. The pump actively transports Na\* out of the cell and K\* into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K\* and Cl\* to flow by passive diffusion. Because of the high negative charge within the cytosol, Cl\* flows out of the cell. The flow of K\* is balanced by an electromotive force pulling K\* into the cell, and a K\* concentration gradient pushing K\* out of the cell. Thus, the resting membrane potential is primarily regulated by K\* flow (Salkoff, L. and T. Jegla (1995) Neuron 15:489-492).

Potassium channel subunits of the <u>Shaker</u>-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The <u>Shaker</u>-like channel family includes the voltage-gated K<sup>+</sup> channels as well as the delayed rectifier type channels such as the human ether-ago-go related gene (HERG) associated with long QT, a cardiac dysrythmia syndrome (Curran, M.E. (1998) Curr. Opin. Biotechnol. 9:565-572; Kaczorowski, G.J. and M.L. Garcia (1999) Curr. Opin. Chem. Biol. 3:448-458).

A second superfamily of K<sup>+</sup> channels is composed of the inward rectifying channels (Kir).

Kir channels have the property of preferentially conducting K<sup>+</sup> currents in the inward direction.

These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K<sup>+</sup> channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) Curr. Opin. Neurobiol. 5:268-277; Curran, supra).

The recently recognized TWIK K<sup>+</sup> channel family includes the mammalian TWIK-1,
TREK-1 and TASK proteins. Members of this family possess an overall structure with four
transmembrane domains and two P domains. These proteins are probably involved in controlling
the resting potential in a large set of cell types (Duprat, F. et al. (1997) EMBO J 16:5464-5471).

The voltage-gated Ca<sup>2+</sup> channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca<sup>2+</sup> channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca<sup>2+</sup> channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α<sub>1</sub> subunit forms the membrane pore and voltage sensor, while the α<sub>2</sub>δ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel. These subunits are encoded by at least six α<sub>1</sub>, one α<sub>2</sub>δ, and four β genes. A fourth subunit, γ, has been identified in skeletal muscle (Walker, D. et al. (1998) J. Biol. Chem. 273:2361-2367; McCleskey, E.W. (1994) Curr. Opin. Neurobiol. 4:304-312).

The high-voltage-activated Ca <sup>2+</sup> channels that have been characterized biochemically include complexes of a pore-forming alpha1 subunit of approximately 190-250 kDa; a

25 transmembrane complex of alpha2 and delta subunits; an intracellular beta subunit; and in some cases a transmembrane gamma subunit. A variety of alpha1 subunits, alpha2delta complexes, beta subunits, and gamma subunits are known. The Cav1 family of alpha1 subunits conduct L-type Ca <sup>2+</sup> currents, which initiate muscle contraction, endocrine secretion, and gene transcription, and are regulated primarily by second messenger-activated protein phosphorylation pathways. The Cav2 family of alpha1 subunits conduct N-type, P/Q-type, and R-type Ca <sup>2+</sup> currents, which initiate rapid synaptic transmission and are regulated primarily by direct interaction with G proteins and SNARE proteins and secondarily by protein phosphorylation. The Cav3 family of alpha1 subunits conduct T-type Ca <sup>2+</sup> currents, which are activated and inactivated more rapidly and at more negative membrane potentials than other Ca <sup>2+</sup> current types. The distinct structures and patterns of regulation of these three families of Ca <sup>2+</sup> channels provide an array of Ca <sup>2+</sup> entry pathways in

response to changes in membrane potential and a range of possibilities for regulation of Ca 2+ entry by second messenger pathways and interacting proteins (Catterall, W.A. (2000) Annu. Rev. Cell Dev. Biol. 16:521-555).

The alpha-2 subunit of the voltage-gated Ca 2+-channel may include one or more Cache domains. An extracellular Cache domain may be fused to an intracellular catalytic domain, such as the histidine kinase, PP2C phosphatase, GGDEF (a predicted diguanylate cyclase), HD-GYP (a predicted phosphodiesterase) or adenylyl cyclase domain, or to a noncatalytic domain, like the methyl-accepting, DNA-binding winged helix-turn-helix, GAF, PAS or HAMP (a domain found in istidine kinases, denylyl cyclases, ethyl-binding proteins and phosphatases). Small molecules are 10 bound via the Cache domain and this signal is converted into diverse outputs depending on the intracellular domains (Anantharaman, V. and Aravind, L.(2000) Trends Biochem. Sci. 25:535-537).

The transient receptor family (Trp) of calcium ion channels are thought to mediate capacitative calcium entry (CCE). CCE is the Ca<sup>2+</sup> influx into cells to resupply Ca<sup>2+</sup> stores depleted by the action of inositol triphosphate (IP3) and other agents in response to numerous hormones and 15 growth factors. Trp and Trp-like were first cloned from Drosophila and have similarity to voltage gated Ca<sup>2+</sup> channels in the S3 through S6 regions. This suggests that Trp and/or related proteins may form mammalian CCE channels (Zhu, X. et al. (1996) Cell 85:661-671; Boulay, G. et al. (1997) J. Biol. Chem. 272:29672-29680). Melastatin is a gene isolated in both the mouse and human, whose expression in melanoma cells is inversely correlated with melanoma aggressiveness in vivo. The human cDNA transcript corresponds to a 1533-amino acid protein having homology to members of the Trp family. It has been proposed that the combined use of malastatin mRNA expression status and tumor thickness might allow for the determination of subgroups of patients at both low and high risk for developing metastatic disease (Duncan, L.M. et al (2001) J. Clin. Oncol. 19:568-576).

Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, Cl<sup>-</sup>enters the cell across a basolateral membrane through an Na<sup>+</sup>, K<sup>+</sup>/Cl<sup>-</sup> cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of Cl from the apical surface, in response to hormonal stimulation, leads to flow of Na and water into the secretory lumen. The cystic fibrosis transmembrane conductance 30 regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear.

35 The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and

devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane domains, as well as two small globular domains known as CBS domains. The CLC subunits probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na+ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as  $\gamma$ -aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four transmembrane domains and probably function as pentamers (Jentsch, supra). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol. 4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K+ channels are gated by internal calcium ions. In nerve cells, an influx of 25 calcium during depolarization opens K<sup>+</sup> channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The  $\alpha$  subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The  $\beta$  subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998) Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best examples of these are the cAMP-gated Na+ channels involved in olfaction and the cGMP-gated

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cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also represent a major pathway for Ca<sup>2+</sup> entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an  $\alpha$  subunit which 5 can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K+ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signalling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the Gβγ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. 15 Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.E. and D.S. Bredt (1998) Cell 93:495-498).

### **Disease Correlation**

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The etiology of numerous human diseases and disorders can be attributed to defects in the transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small 25 molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of 30 sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers. Mutations in muscle sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrythmia disorders such as the long QT 35 syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium

channels (Cooper, E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, supra).

Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 10 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na+ channels have been useful in the treatment of neuropathic pain (Eglen, supra). Calcium-channel protein expression is altered in metastatic melanomas (Enklaar, T. et al. (2000) Genomics 67:179-187.

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious immunosuppressants (Cahalan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

The discovery of new transporters and ion channels, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of transport, neurological, muscle, immunological and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

### SUMMARY OF THE INVENTION

The invention features purified polypeptides, transporters and ion channels, referred to collectively as "TRICH" and individually as "TRICH-1," "TRICH-2," "TRICH-3," "TRICH-4," "TRICH-5," "TRICH-6," "TRICH-7," "TRICH-8," "TRICH-9," "TRICH-10," "TRICH-11," "TRICH-12," "TRICH-13," "TRICH-14," "TRICH-15," "TRICH-16," "TRICH-17," "TRICH-18," "TRICH-19," "TRICH-20," "TRICH-21," "TRICH-22," "TRICH-23," "TRICH-24," "TRICH-25,"

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"TRICH-26," "TRICH-27," "TRICH-28," "TRICH-29," "TRICH-30," "TRICH-31," and "TRICH-32." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-32. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:1-32.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide.

30 In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of

SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous

nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional TRICH,

comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide 5 comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample 10 comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide 20 having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide 30 having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of 35 the polypeptide in the presence of the test compound with the activity of the polypeptide in the

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absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a 5 polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) 15 a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide 20 in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEO ID NO:33-64, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide 25 complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization 30 complex in the treated biological sample is indicative of toxicity of the test compound.

#### BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

35 Table 2 shows the GenBank identification number and annotation of the nearest GenBank

homolog for polypeptides of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

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#### **DESCRIPTION OF THE INVENTION**

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

#### **DEFINITIONS**

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"TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of 15 nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition 20 are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding TRICH. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a 25 functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar 30 side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or 35 synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally

occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

5 Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of TRICH. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')<sub>2</sub>, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an in vitro evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No. 5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH<sub>2</sub>), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules,

e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker. (See, e.g., Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13.)

The term "intramer" refers to an aptamer which is expressed in vivo. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding TRICH or fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-

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dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, 10 Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
20 25	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
	Gln ·	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Пе	Leu, Val
30	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
35	Thr	Ser, Val
	$\operatorname{Trp}$	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the

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absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of TRICH or the polynucleotide encoding TRICH which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a 25 molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:33-64 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:33-64, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEO ID NO:33-64 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:33-64 from related polynucleotide sequences. The precise length of a fragment of SEQ 35 ID NO:33-64 and the region of SEQ ID NO:33-64 to which the fragment corresponds are routinely

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determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-32 is encoded by a fragment of SEQ ID NO:33-64. A fragment of SEQ ID NO:1-32 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-32. For example, a fragment of SEQ ID NO:1-32 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-32. The precise length of a fragment of SEQ ID NO:1-32 and the region of SEQ ID NO:1-32 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default

20 parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e
sequence alignment program. This program is part of the LASERGENE software package, a suite of
molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in
Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992)

CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters

25 are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted"
residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the
"percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2

Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html.

The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool

Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62
Reward for match: 1
Penalty for mismatch: -2

10 Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50:

Expect: 10
Word Size: 11
Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap

penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas

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wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 µg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence at a defined ionic strength and pH. The T<sub>m</sub> is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for 10 calculating T<sub>m</sub> and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, 15 for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., Cot or Rot analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate 30 substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression 35 of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect

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cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide

fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of TRICH.

"Probe" refers to nucleic acid sequences encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule.

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Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences.

10 Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2<sup>nd</sup> ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple

sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence
that is made by an artificial combination of two or more otherwise separated segments of sequence.
This artificial combination is often accomplished by chemical synthesis or, more commonly, by the
artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques
such as those described in Sambrook, supra. The term recombinant includes nucleic acids that have
been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently,
a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter
sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to
transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or

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cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, mugnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is 25 selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by

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infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence 10 having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, or at least 98%, or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the 20 reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single. nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, or at least 98%, or greater sequence identity over a certain defined length of one of the polypeptides.

#### 55 THE INVENTION

The invention is based on the discovery of new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, immunological and cell proliferative disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide 10 sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are transporters and ion channels. For example, SEQ ID NO:1 is 83% identical to human sodium-hydrogen exchanger 6 (GenBank ID g2944233) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.1e-242, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:1 also contains a sodium/hydrogen

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exchanger family domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, and other BLAST analyses provide further corroborative evidence that SEQ ID NO:1 is a sodium-hydrogen exchange transporter. In another example, SEQ ID NO:7 is 85% identical to Rattus norvegicus Na<sup>+</sup>/,K<sup>+</sup>-ATPase alpha subunit (GenBank ID g619915) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:7 also contains an E1-E2 ATPase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-10 based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:7 is a cation-transporting ATPase. In yet another example, SEQ ID NO:13 is 77% identical to a human carrier-like protein (GenBank ID g3694661) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.5e-209, which indicates 15 the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:13 also contains a mitochondrial energy transfer protein domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) The presence of this domain is confirmed by BLIMPS, MOTIFS, and PROFILESCAN analyses, providing further corroborative evidence that 20 SEQ ID NO:13 is a transporter. Further, SEQ ID NO:16 is 41% identical to human novel ATPase (GenBank ID g8979801) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.1e-165, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains an E1-E2 ATPase domain as determined by searching for statistically significant matches in the hidden 25 Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:16 is a cation-transporting ATPase. In a further example, SEQ ID NO:19 is 43% identical to Sinorhizobium sp. As4 ArsA, the catalytic subunit of the arsenic oxyanion-translocating ATPase (GenBank ID g5802945) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 7.7e-125, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:19 also contains an anion-transporting ATPase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from additional BLAST and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:19 is an anion-transporting ATPase. In yet a further

example, SEQ ID NO:21 is 54% identical to a murine putative E1-E2 ATPase (GenBank ID g28577) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.2e-190, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:21 also contains six transmembrane domains as determined using TMAP, a program which delineates transmembrane segments. (See Table 3.) Data from BLIMPS, and MOTIFS, analyses provide further corroborative evidence that SEQ ID NO:21 is an ATPase. In a futher example, SEQ ID NO:24 is 52% identical, from residue A77 to residue L1007, to rat NMDAR-L (GenBank ID g2160125) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.5e-262, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:24 also contains a ligand gated ion channel domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS analysis provide further corroborative evidence that SEQ ID NO:24 is a glutamate receptor. SEQ ID NO:2-6, SEQ ID NO:8-15 12, SEQ ID NO:14-15, SEQ ID NO:17-18, SEQ ID NO:20, SEQ ID NO:22-23, and SEQ ID NO:25-32 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO: 1-32 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide sequences of the invention, and of fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:33-64 or that distinguish between SEQ ID NO:33-64 and related polynucleotide sequences.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank cDNAs or ESTs which contributed to the assembly of the full length polynucleotide sequences. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences

including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence 5 identified as FL\_XXXXXX\_N<sub>1</sub>\_N<sub>2</sub>\_YYYYY\_N<sub>3</sub>\_N<sub>4</sub> represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and  $N_{123}$ , if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons 10 brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as FLXXXXXX\_gAAAAA\_gBBBBB\_1\_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs
GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).
INCY	Full length transcript and exon prediction from mapping of EST
	sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in

Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant
Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide

sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural characteristic of TRICH.

The invention also encompasses polynucleotides which encode TRICH. In a particular 10 embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:33-64, which encodes TRICH. The polynucleotide sequences of SEQ ID NO:33-64, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding TRICH. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID 20 NO:33-64 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:33-64. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of TRICH.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant 25 of a polynucleotide sequence encoding TRICH. A splice variant may have portions which have significant sequence identity to the polynucleotide sequence encoding TRICH, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% 30 polynucleotide sequence identity to the polynucleotide sequence encoding TRICH over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide sequence encoding TRICH. Any one of the splice variants described above can encode an amino acid sequence which contains at least one functional or 35 structural characteristic of TRICH.

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It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode TRICH and its variants are generally capable

of hybridizing to the nucleotide sequence of the naturally occurring TRICH under appropriately
selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding
TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of nonnaturally occurring codons. Codons may be selected to increase the rate at which expression of the
peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with

which particular codons are utilized by the host. Other reasons for substantially altering the
nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid
sequences include the production of RNA transcripts having more desirable properties, such as a
greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding TRICH or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of
hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID
NO:33-64 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and
S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol.
152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in
"Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the

ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence

preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

10 The nucleic acid sequences encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and 25 PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express TRICH.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians; F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with

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fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding TRICH may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular 10 Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring 15 polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active TRICH, the nucleotide sequences encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in 25 polynucleotide sequences encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no 30 additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ.

20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding TRICH and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989)

Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding TRICH can be achieved using a multifunctional <u>E. coli</u> vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding TRICH into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for <u>in vitro</u> transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M.

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Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 10 1995, supra, Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of TRICH. Transcription of sequences encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the oniega leader sequence from TMV (Takamatsu, N. (1987) EMBO 15 J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or

pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to 25 obtain infective virus which expresses TRICH in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of 30 DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of TRICH in cell lines is preferred. For example, sequences encoding TRICH can be transformed

into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., 15 Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing sequences encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

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Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols,

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding TRICH, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by

Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding TRICH may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for

post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding TRICH may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available 10 affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable 15 immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra. ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled TRICH may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, <sup>35</sup>S-methionine.

TRICH of the present invention or fragments thereof may be used to screen for compounds that specifically bind to TRICH. At least one and up to a plurality of test compounds may be screened for specific binding to TRICH. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which TRICH binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case,

the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, Drosophila, or E. coli. Cells expressing TRICH or cell membrane fractions which contain TRICH are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

TRICH of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence 20 of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an in vitro or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 30 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous 35 recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or

developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

## **THERAPEUTICS**

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. In addition, examples of tissues expressing TRICH can be found in Table 6. Therefore, TRICH appears to play a role in transport, neurological, muscle, immunological and cell proliferative disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity of TRICH. In the treatment of disorders associated with decreased TRICH expression or activity, it is desirable to increase the expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug

resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, 5 mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder. paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup 15 disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating 20 diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess. suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, 25 encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular 35 dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy,

lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple 15 sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be

administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such disorders include, but are not limited to, those transport, neurological, muscle, immunological and cell proliferative disorders described above. In one aspect, an antibody which specifically binds TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made

15 by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of TRICH may be produced using methods which are generally known in the
art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of
pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may
also be generated using methods that are well known in the art. Such antibodies may include, but
are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments,
and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which
inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to,

30 Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to
TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will

consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and

Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-20 10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to,  $F(ab')_2$  fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the  $F(ab')_2$  fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies

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reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an 5 association constant, K<sub>n</sub>, which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K<sub>a</sub> determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies for TRICH. The K<sub>n</sub> determined for a preparation of monoclonal antibodies, which are monospecific for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K<sub>a</sub> ranging from about 10<sup>9</sup> to 10<sup>12</sup> L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K<sub>a</sub> ranging from about 10<sup>6</sup> to 10<sup>7</sup> L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, 15 preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense 30 oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence

complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding TRICH may be used for 10 somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase 15 (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 20 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida 25 <u>albicans</u> and <u>Paracoccidioides brasiliensis</u>; and protozoan parasites such as <u>Plasmodium falciparum</u> and <u>Trypanosoma cruzi</u>). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr.

Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)): the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen): the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi. F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized manimalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant")

discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4<sup>+</sup> T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with 20 respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. 25 Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also 30 taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and 35 propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to

those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into 10 the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by

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scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6.

Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2'O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in

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altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a 5 compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding 10 TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a 15 change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa 20 cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable

excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example
TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of

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TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED<sub>50</sub> (the dose therapeutically effective in 50% of the population) or LD<sub>50</sub> (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD<sub>50</sub>/ED<sub>50</sub> ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED<sub>50</sub> with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1  $\mu$ g to 100,000  $\mu$ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

## DIAGNOSTICS

In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect

TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression.

Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of TRICH expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding TRICH may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:33-64 or from genomic sequences including promoters, enhancers, and introns of the TRICH gene.

Means for producing specific hybridization probes for DNAs encoding TRICH include the cloning of polynucleotide sequences encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as <sup>32</sup>P or <sup>35</sup>S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic

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fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive

dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive

supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia 10 congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, 15 diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. The polynucleotide sequences encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding TRICH may be useful in assays

PCT/US01/46963 WO 02/46415

that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a 5 standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from 15 normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent

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conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in highthroughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this

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information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is

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useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance 20 under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are 30 compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) Anal. 5 Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Scilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteonuc profile. In addition, the analysis of transcripts in body fluids is difficult, 15 due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embediment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount 30 of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; 35 Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad.

Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in DNA Microarrays: A Practical Approach, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during 10 chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. 15 (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide

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sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with TRICH.

In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below, including U.S. Ser. No. 60/254,303, U.S. Ser. No. 60/256,190, U.S. Ser. No. 60/257,504, U.S. Ser. No. 60/261,546, U.S. Ser. No. 60/262,832, U.S. Ser. No. 60/264,377, and U.S. Ser. No. 60/266,019, are expressly incorporated by reference herein.

### **EXAMPLES**

### I. Construction of cDNA Libraries

35 Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD

database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles 10 (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected 20 (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid 25 (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Life Technologies.

### II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems 35 or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids

were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

### III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows.

Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae,

Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs,

stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:33-64. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

### IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range

of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

### V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example 20 III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the 25 interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm 30 in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases.

genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

### "Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST

analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs

(HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

### VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:33-64 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:33-64 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously

identified disease genes map within or in proximity to the intervals indicated above.

### VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, supra, ch. 7; Ausubel (1995) supra, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

15

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous

system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

### 10 VIII. Extension of TRICH Encoding Polynucleotides

Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment.

One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg<sup>2+</sup>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech),

ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100  $\mu$ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5  $\mu$ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the

concentration of DNA. A 5  $\mu$ l to 10  $\mu$ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates,

digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and
sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For
shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%)
agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended
clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector

(Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction
site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on
antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in
384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase

(Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following
parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min;
Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was
quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low
DNA recoveries were reamplified using the same conditions as described above. Samples were
diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer
sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI
PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

### IX. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:33-64 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [γ-32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech).

An aliquot containing 107 counts per minute of the labeled probe is used in a typical membrane-

based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

### X. ... Microarrays

The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645;

Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

### **Tissue or Cell Sample Preparation**

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)<sup>+</sup> RNA is purified using the oligo-(dT) cellulose method. Each poly(A)<sup>+</sup> RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/µl oligo-(dT) primer (21mer), 1X

first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)<sup>+</sup> RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)<sup>+</sup> RNAs are synthesized by in vitro

5 transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories. Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

### Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from becterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5  $\mu$ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1  $\mu$ l of the array element DNA, at an average concentration of 100 ng/ $\mu$ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0,2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

### 5 Hybridization

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Hybridization reactions contain 9  $\mu$ l of sample mixture consisting of 0.2  $\mu$ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140  $\mu$ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

### 10 <u>Detection</u>

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Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-

compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

### XI. Complementary Polynucleotides

Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

### XII. Expression of TRICH

Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription.

35 Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or

washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

### XVI. Identification of Molecules Which Interact with TRICH

Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as Gβγ proteins (Reimann, supra) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, supra). TRICH, or biologically active fragments thereof, are labeled with <sup>125</sup>I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA, and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the TRICH interacting proteins (fusion proteins with an activation domain) reconstitute a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH ion channel activity using the assays described in section XVIII.

### XVII. Demonstration of TRICH Activity

Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as B-galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after

transformation under conditions appropriate for the cell line to allow expression and accumulation of TRICH and  $\beta$ -galactosidase.

Transformed cells expressing \( \text{B}\)-galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or \( \text{B}\)-galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel, and the associated conductance.

Alternatively, ion channel activity of TRICH is measured as current flow across a TRICH-, containing Xenopus laevis oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., supra; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate Xenopus oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into mature stage IV oocytes. Injected oocytes are incubated at 18 °C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca<sup>+2</sup> (in the form of CaCl<sub>2</sub>), where appropriate.

Electrode resistance is set at 2-5 MΩ and electrodes are filled with the intracellular solution lacking mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay. In particular, the activity of TRICH-1 is measured as Na<sup>+</sup> conductance and the activity of TRICH-3 is measured as Ca<sup>2+</sup> conductance.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates into Xenopus laevis oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl 2, 1mM MgCl 2, 1mM Na<sub>2</sub>HPO<sub>4</sub>, 5 mM Hepes, 3.8 mM NaOH, 50µg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl 2, 1mM MgCl 2, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with <sup>3</sup>H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na<sup>+</sup>-free medium, measuring the incorporated label, and comparing with controls. TRICH

activity is proportional to the level of internalized labeled substrate. Test substrates include ran-GTP for TRICH-2, glucose for TRICH-5, amino acids for TRICH-6 and TRICH-14, cations for TRICH-7 and TRICH-16, Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> ions for TRICH-15, reduced folate or analogues such as methotrexate for TRICH-17, divalent cations for TRICH-18, anions such as arsenate and antimonite for TRICH-19, and nitrate or oligopeptides for TRICH-20.

ATP-[γ-<sup>32</sup>P], separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered <sup>32</sup>P using a scintillation counter. The reaction mixture contains ATP-[γ-<sup>32</sup>P] and varying amounts of TRICH in a suitable buffer incubated at 37°C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of <sup>32</sup>P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

Lipocalin activity of TRICH is measured by ligand fluorescence enhancement spectrofluorometry (Lin et al. (1997) Molecular Vision 3:17). Examples of ligands include retinol (Sigma, St. Louis MO) and 16-anthryloxy-palmitic acid (16-AP) (Molecular Probes Inc., Eugene OR). Ligand is dissolved in 100% ethanol and its concentration is estimated using known extinction coefficients (retinol: 46,000 A/M/cm at 325 nm; 16-AP: 8,200 A/M/cm at 361 nm). A 700  $\mu$ l aliquot of 1  $\mu$ M TRICH in 10 mM Tris (pH 7.5), 2 mM EDTA, and 500 mM NaCl is placed in a 1 cm path length quartz cuvette and 1  $\mu$ l aliquots of ligand solution are added. Fluorescence is measured 100 seconds after each addition until readings are stable. Change in fluorescence per unit change in ligand concentration is proportional to TRICH activity.

### XVIII. Identification of TRICH Agonists and Antagonists

TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the Ca<sup>2+</sup> indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Cl<sup>-</sup> indicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric plate reading system (Molecular Devices). In a more generic version of this assay, changes in

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membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC<sub>4</sub> (Molecular Probes). DiBAC<sub>4</sub> equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC<sub>4</sub> entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

### Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
· · · · · · · · · · · · · · · · · · ·	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide
. 007.07.12				A A
7484831	1	7484831CD1	33	7484831CB1
2477266	2	2477266CD1	34	2477266CB1
3552033	3	3552033CD1	35	3552033CB1
4778139	4	4778139CD1	36	4778139CB1
4787433	5	4787433CD1	37	4787433CB1
7483598	9	7483598CD1	38	7483598CB1
7484823	7	7484823CD1	39	7484823CB1
143935	8	143935CD1	40	143935CB1
5923789	6	5923789CD1	41	5923789CB1
6046484	10	6046484CD1	42	6046484CB1
7481427	11	7481427CD1	43	7481427CB1
7483595	12	7483595CD1	44	7483595CB1
3788427	13	3788427CD1	45	3788427CB1
6972455	14	6972455CD1	46	6972455CB1
8077668	15	8077668CD1	47	8077668CB1
55120485	16	55120485CD1	48	55120485CB1
3112883	17	3112883CD1	49	3112883CB1
4253888	18	4253888CD1	20	4253888CB1
7479974	19	7479974CD1	51	7479974CB1
7483850	20	7483850CD1	25	7483850CB1
5508353	21	5508353CD1	53	5508353CB1
8543628	22	8543628CD1	54	8543628CB1
7482754	23	7482754CD1	55	7482754CB1
3794818	24	3794818CD1	56	3794818CB1
4717525	25	4717525CD1		4717525CB1
5091793	26	5091793CD1	58	5091793CB1
5945527	27	5945527CD1		5945527CB1
6941124	28	6941124CD1	09	6941124CB1
6972530	29	6972530CD1	61	6972530CB1

Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
	SEQ ID NO:	Polypeptide ID SEQ ID NO:	SEQ ID NO:	Polynucleotide ID
6991750	30	6991750CD1	62	6991750CB1
71726948	31	71726948CD1	63	71726948CB1
7487393	32	7487393CD1	64	7487393CB1

Polypeptide SEQ ID NO:	Incyte Polypeptide ID	GenBank ID NO:	Probability score	GenBank Homolog
٦	7484831CD1	g29 <u>4</u> 4233	5.1e-242	Sodium-hydrogen exchanger 6 [Homo sapiens] (Numata, M. et al. (1998) J. Biol. Chem. 273:6951-6959)
2	2477266CD1	g2102696	1.0e-37	omo sap
			- 24	Yaseen, N.R. and Blobel, G. (1997) Proc. Natl. Acad. Sci. U.S.A. 94:4451-4456
3	3552033CD1	g3243075	0.0	[Homo sapiens] melastatin 1
				<pre>Hunter, J.J. et al. (1998) Genomics 1998 54:116-123</pre>
4	4778139CD1	g8131903	5.1e-107	[Mus musculus] transient receptor potential-related protein
5	4787433CD1	g2337865	2.3e-251	[Homo sapiens] organic cation transporter
9	7483598CD1	g6978016	1.9e-32	[Rattus norvegicus] neuronal glutamine transporter
	•			Varoqui,H. et al. (2000) J. Biol. Chem. 275:4049-4054
7	7484823CD1	g619915	0.0	[Rattus norvegicus] Na,K-ATPase alpha subunit
				Shamraj, O.I., and Lingrel, J.B. (1994) Proc. Natl. Acad. Sci. USA 91:12952-12956
8	143935CD1	g179304	7.8e-116	B12 protein [Homo sapiens] (Wolf, F.W. et al. (1992) J. Biol. Chem. 267:1317-1326)
6	5923789CD1	g1552526	0.0	sodium-calcium exchanger form 3 [Rattus norvegicus] (Nicoll, D.A. et al. (1996) J. Biol. Chem. 271:24914-24921)
10	6046484CD1	g3243075	0.0	melastatin 1 [Homo sapiens] (Hunter, J.J. (1998) Genomics 54:116-123)
11	7481427CD1	g178661	9.8e-93	adenine nucleotide translocator-2 [Homo sapiens] (Ku, D.H. et al. (1990) J. Biol. Chem. 265: 16060-16063)

Table 2 (cont.)

Polypeptide SEQ ID NO:	Incyte Polypeptide ID	GenBank ID NO:	Probability score	GenBank Homolog
12	7483595CD1	g9453726	3.7e-61	bA48209.2 (Novel sulphate transporter family member) [Homo sapiens]
13	3788427CD1	g3694661	5.5e-209	carrier protein-like; similar to 001888 (PID:9266574) [Homo sapiens]
14	6972455CD1	g4155688	1.7e-24	(Helicobactor pylori J99) AMINO ACID ABC TRAUSPOTER, BINDING PROTEIN PRECUSSOR.
15	8077668CD1	95081312	6.60-47	[Partus norvegicus] bumetanide- sensitive Ka-K-2Cl cotransporter Anzai, H., et al. (1999) Roles of vastpressin and hypertonicity in baselateral Ra/K/2Cl cotransporter expression in rat kidney inner medullary collecting duct cells. Jpn. J. Physiol. 49, 201-206
16	55120485CD1	g8979801	1.1e-165	dJ37C10.3 (novel ATPase) [Homo sapiens]
17	3112883CD1	g3115983	4.0e-128	dJ206D15.1 (Reduced Folate Carier protein RFC LIKE) [Homo sapiens]
18	4253888CD1	g3925431	1.6e-29	[Caenorhabditis elegans] (Z82084) contains similarity to Pfam domain: PF01769 (Divalent cation transporter), Score=211.5, E- value=4.2e-60, N=2
19	7479974CD1	g5802945	7.7e-125	[Sinorhizobium sp. As4] ArsA (catalytic subunit of arsenic oxyanion-translocating ATPase)
20	7483850CD1	g11933414	6.3e-11	[Glycine max] nitrate transporter NRT1-5
21	5508353CD1	g6457270	5.2e-190	[Mus musculus] putative E1-E2 ATPase Halleck,M.S. et al., (1999) Physiol. Genomics (Online) 1:139-150 MEDLINE : 20473714
22	8543628CD1	g6967939	3.7e-45	[Campylobacter jejuni] amino-acid ABC transporter integral membrane protein Takamori S. et al., (2000) Nature 407:189-94

Table 2 (cont.)

Polypeptide SEQ ID NO:	Incyte Polypeptide ID	GenBank ID NO:	Probability score	GenBank Homolog
23	7482754CD1	g11640743	1.9e-20	[Homo sapiens] amino acid
				transporter system Al
				Waiig n. et al., (2000) Biochem Riophys Res Commin
				273:1175-9
24	3794818CD1	g2160125	1.5e-262	NMDAR-L [Rattus norvegicus]
				Sucher, N.J. et al. (1995) J.
				Neurosci. 15:6509-6520
25	4717525CD1	g6841066	7.8e-111	calcium-binding transporter [Homo
26	5091793CD1	93880532	1 10-51	try to miltidania
			1	protein (SW.BMR1 BACSI)
				[Caenorhabditis elegans]
				The C. elegans Segmenting Consortium
				(1998) Science 282: 2012-2018
27	5945527CD1	g7543982	1.5e-161	glycerol 3-phosphate permease (Homo
				sapiens]
28	6941124CD1	g476222	6.8e~66	anion exchanger 3 brain isoform
				[Homo sapiens]
				Res. 75:603-614
29	6972530CD1	g10175963	3.5e-16	potassium channel protein [Bacillus
				halodurans]
				Takami, H. et al. (1999)
30	6991750CD1	g6273849	5.5e-11	Cardiac sodium-calcium exchanger
		•		Xue, X.H. et al. (1999) Am. J.
				Physiol. 277:C693-C700
31	71726948CD1	g1628579	1.0e-152	sodium iodide symporter [Homo
	•			sapiens]
				.A. et al. (19
000	200000			Res. Commun. 226:
	/48/393CDI	g//0/622	2.2e-118	organic anion transporter 4 [Homo
				Cha, S.H. et al. (2000) J. Biol.
				Chem. 275:4507-4512

### Table 3

Analytical Methods and Databases	HMMER	3, TMAP	HMMER-PFAM	BLIMPS-PRINTS	BLIMPS-PRINTS	BLIMPS-PRODOM	BLAST-PRODOM	cer, BLAST-PRODOM	BLAST-DOMO 117 124 1120,
Signature Sequences, Domains and Motifs	Signal peptide: M1-A37	Transmembrane domains: P18-A39, R68-I88, R95-I115, K175-H203 L208-L236, D245-A269, A282-Q306, A319-L347, L360-L388, Y428-G456, H482-T502, Q508-L536 N-terminus is non-cytosolic	1	Na+/H+ exchanger isoform signatures PR01088: S44-A63, E64-188, W89-1107, Y108-Q134 S299-D316, A318-M337, G588-D606, P612-O640, V641-D668	ld	Na+ transport exchanger PD01672: V182-M230, A319-V344, F381-F414, F419-S465, F466-T512	<pre>Sodium hydrogen exchanger 6, myeloblast PD177855: Y557-N725, G527-E547</pre>	lēā	Deta Na exchanger:  DM02572   P48764   10-734: D171-R539, R21-L116  DM02572   Q01345   12-703: D179-D563, L22-R117  DM02572   P50482   16-723: E181-F558, G16-S124  DM02572   P26434   14-716: D179-D563, A48-T120
Potential S Glycosyl- ation Sites		H		Z 0.			о ц	2 0	
	52 S66 S147 S198 S260 S546 S689 S694 S712 T59 T154 T177								
Amino Acid Residues									
Incyte Polypep- tide ID	7484831CD1								
EQ LD NO:	1								

Analytical Methods and Databases	ТМАР	BLAST_PRODOM	MOTIFS	HMMER_PFAM	TMAP	BLIMPS_PRINTS	BLAST_PRODOM
Signature Sequences, Domains and Motifs	Transmembrane domains: P197-L213 R255-V275 E498-P520 I839-V863 N terminus cytosolic	TEIN	Leucine zipper pattorn L177-L198 Phospholipase A2 histidine active site	Transient receptor: Y943-M1001, R817-E882, E750-L807, D562-W608	rane domains: G51-175 D397-R425 E786-R806 V822-G842 M852-A872 s cytosolic	Transient receptor potential family signature PR01097: A941-T962, F963-F976, V990-M1003	ROMOSOME TRANSMEMBRANE PD151509: 1829-L1117 PD022180: W328-R438
Potential Glycosyl- ation Sites	N165 N686	÷		N144 N233 N298 N420 N576 N579 N789 N915 N960 N1058 N1074			
Potential Phosphoryl- ation Sites	S47 S79 S148 S180 S192 S315 S493 S615 S639 S697 S1011 T18 T25 T61 T147 T167 T328 T532 T586 T786 T813 T871 T881 T907			S212 S235 S300 S366 S401 S528 S558 S618 S687 S688 S884 S1017 S1059 S1060 S1069 S1076 S1088 S1125 T9 T147 T422 T459 T460 T917 T962 T984 T1031 T1112 T1118 T1132 T1155			
due	1 80 0 1			1172 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8 8			
Incyte Polypep- tide ID	747/700000			3552033CD1			
SA S.	N			m			

Analytical Methods and Databases	BLAST_DOMO	I MOTIFS	N BLAST_PRODOM				HMMER_PFAM		TMAP		t	BLIMPS PRINTS	MOTIFS	HMMER_PFAM	TMAP		BLAST PRODOM	
Signature Sequences, Domains and Motifs	ANK MOTIF REPEAT DM03196 P34586 38-822: 1819-C1009, L583-G619, L702-L807, D114-N144, T9-Q48	Aminoacyl-transfer RNA synthetases class-II signature 2 A1111-L1120	KINASE TRANSFERASE SERINE/THREONINE PROTEIN ATP BINDING ELONGATION FACTOR EEF2 EEF2K CALCTIM/CALMONITINDEDENDENT FIRABPORTS	PD011701: K536-R709			Sugar (and other) transporter: K120-E538		rane domains: G17-G4	L185-1205  F214-1234  S243-1265  L269-F293  1355-8377  N390-D410  T416-P436  L442-Y462	- 1	Na+/H+ exchanger isoform PR010870 I32-V46	Leucine zipper pattern L146-L167	Transmembrane amino acid transporter protein: S56-G412		A250-F278 F328-A348 L358-P378 T391-N419	ACID AMINO PROFEIN TRANSPORTER PERMEASE	TRANSMEMBRANE INTERGENIC REGION PUTATIVE PROLINE PD001875: F39-T209
Potential Glycosyl- ation Sites			N238 N258 N294 N650 N711	1			N31 N57 N65 N68 N108 N345	N352 N546 N558						N100 N331 N436 N441 N457				
Potential Phosphoryl- ation Sites			S42 S104 S135 S194 S203 S214	S239 S277 S319 S339 S352 S354	S491 S491 S735	T371 T619	3119 S337	S550 S560 T135 T356 T521 T534 T535 T569 Y10						S24 S56 S90 S242 S243 S393 T282 T391	ł			
Amino Acid Residues			742				577							462				
Incyte Polypep- tide ID		•	4778139CD1				4787433CD1							7483598CD1				
OH CON	3		4				2							<b>9</b>				

Phosphory -   Glycosy -   Gl		Incyte	Amino	Potential		Signature Sequences,	Analytical
1018   S10 S49 S151	t Po	lypep- de ID	Acid Residues	Phosphor ation Si		Domains and Motifs	ethods and
T873 593 T44	74	84823CD1	1018	S10 S49 S1 S157 S372 S525 S585	N212 N480	(cation transport) ATPase:	HMMER_PFAM
1339 1494 1494     1444 1489     1465 1491     1464 1489     1464 1489     1465 1491				8938 1250 1			
Na+/K+ ATPase C-terminus:   R829-Y1017, E30-S113     Transmembrane domains: H287-L315 E781-T809     I845-P873 V909-1931 A972-R1000 N terminus     non-cytosolic     E1-E2 ATPases phosphoryl BL00154: V329-     G365, T367-V385, K604-C514, D588-I628, V707-G730, G733-N766, G185-L202     E1-E2 ATPases phosphorylation site atpase_e1_e2.prf: L354-E401     P-type cation-transporting atpase superfamily signature PR00119: D211-S225, C371-V385, G582-A553, A604-D614, T710-M729, S734-L746     H-transporting ATPase (proton pump)     signature PR00120: E682-E698, T710-G726, D742-L767     Sodium/potassium-transporting ATPase     signature PR00121: L100-L1519, I782-L803, V849-A869, F911-1931, R945-A969, B718-S225, V581-I629     ATPASE TRANSMEMBRANE TRANSPORT PUMP     MAGNESIUM PD000131: L643-N749, I587-I629     PTYPE TRANSPORTING ATPASE I PD111120: I				T399 T482			
Na+/K+ ATPase C-terminus:   R829-Y1017, E30-S113				635 T755 461 Y889			
Transmembrane domains: H287-L315 E781-T809							HMMER_PFAM
Detacrosolic   Detacrosolic		•				ne domains: H287-L315 E781-T809	MAP
E1-E2 ATPases phosphory1 BL00154: V329-G365, T367-V385, K504-C514, D588-I628, V707-G730, G733-N766, G185-L202 E1-E2 ATPases phosphorylation site atpase e1 e2.prf: L354-E401 P-type cation-transporting atpase superfamily signature PR00119: D211-S225, C371-V385, G582-A593, A604-D614, T710-M729, S734-L746 H+-transporting ATPase (proton pump) signature PR00120: E682-E698, T710-G726, D742-L767 Sodium/potassium-transporting ATPase signature PR00121: L100-I114, L127-Q147, L291-G313, L364-V385, L501-L519, I782-L803, Y849-A869, F911-1931, R945-M969 ATPASE TRANSMEMBRANE TRANSPORT PUMP MAGNESIUM PD000132: V132-Y312, W314-W426, D667-E820, K473-D742, F118-S225, V581-I629 CALCIUM PD000121: L643-N749, I587-I629 PTYPE TRANSPORTING ATPASE 1 PD111120:						N DOOL AS I SALLOUD	
es phosphorylation site 2.prf: L354-E401 on-transporting atpase signature PR00119: D211-S225, G582-A593, A604-D614, T710-M729, G582-A593, A604-D614, T710-M729, Eing ATPase (proton pump) R00120: E682-E698, T710-G726, Ssium-transporting ATPase R00121: L100-1114, L127-Q147, L364-V385, L501-L519, I782-L803, E911-1931, R945-M969 SMEMBRANE TRANSPORT PUMP D000132: V132-Y312, W314-W426, X473-D742, F118-S225, V581-I628 D0121: L643-N749, I587-I629 D0388: K828-Y1017		•				ATPases phosphoryl BL00154: V329- T367-V385, K504-C514, D588-I628, 730. G733-N766, G185-L700	BLIMPS_BLOCKS
on-transporting atpase signature PR00119: D211-S225, G582-A593, A604-D614, T710-M729, Eing ATPase (proton pump) R00120: E682-E698, T710-G726, Ssium-transporting ATPase R00121: L100-1114, L127-Q147, L364-V385, L501-L519, I782-L803, E911-1931, R945-M969 SMEMBRANE TRANSPORT PUMP D000132: V132-Y312, W314-N426, X473-D742, F118-S225, V581-I628 D0121: L643-N749, I587-I629 D0388: K828-Y1017						E1-E2 ATPases phosphorylation site atpase_e1_e2.prf: L354-E401	PROFILESCAN
Signacure Fronis: Dill-Siz, G582-A593, A604-D614, T710-M729, Eing ATPase (proton pump)  R00120: E682-E698, T710-G726, Sium-transporting ATPase R00121: L100-1114, L127-Q147, L364-V385, L501-L519, I782-L803, E911-1931, R945-M969 SMEMBRANE TRANSPORT PUMP D000132: V132-Y312, W314-W426, K473-D742, F118-S225, V581-I628 D0121: L643-N749, I587-I629 D0388: K828-Y1017						ting atpase	BLIMPS_PRINTS
ting ATPase (proton pump)  R00120: E682-E698, T710-G726,  ssium-transporting ATPase  R00121: L100-1114, L127-Q147,  L364-V385, L501-L519, I782-L803,  F911-1931, R945-M969  SMEMBRANE TRANSPORT PUMP  SM13-D742, F118-S225, V581-1628  D0121: L643-N749, I587-1629  SORTING ATPASE 1 PD111120:			•		<u> </u>	signacure FROULIS: D. 3582-A593, A604-D614,	•
g ATPase , L127-0147, 519, I782-L803, 969 RT PUMP 2, W314-N426, 225, V581-I628 I587-I629						Ling ATPase (proton pump) R00120: E682-E698, T710-G726,	BLIMPS_PRINTS
DIS, 1/82-L803, 969 RT PUMP 2, W314-N426, 225, V581-1628 I587-1629		,				Pase 27-0147,	BLIMPS_PRINTS
RT PUMP 2, W314-N426, 225, V581-1628 1587-1629 PD111120:				•		7271-6313, 1304-7363, 1301-1313, 1762-1603, 7849-A869, F911-1931, R945-M969	·
225, V581-1628 1587-1629 PD111120:			. —			PUMP W314-N426.	BLAST_PRODOM
PD111120:						0667-E820, K473-D742, F118-S225, V581-I628 CALCIUM PD000121: L643-N749, I587-I629	
74 > 1 4						PD111120:	BLAST_PRODOM

Analytical Methods and Databases	TMAP	SPSCAN	HMMER	нимек_рғам	BLIMPS_PRINTS	BLAST PRODOM	BLAST_PRODOM	ТМАР	HMMER_PFAM
ences, otifs	Transmembrane domains: A2-R29, K73-S101, P167-P190, T194-M219, T237-M261, S720-P741, G748-T776, T776-D797, I815-W841, H849-R877, C891-T911 N-terminus is cytosolic	U	Signal Peptide: M1-A32	Transient receptor: Y922-H979, R791-L851,	Transient receptor potential family PR01097: A920-T941, F942-F955	O	MELASTATIN CHROMOSOME TRANSMEMBRANE C05C12.3 T01H8.5 I F54D1.5 IV PD018035: M1-L333 PD151509: I803-L1097 PD039592: E454-T652	1 (	Mitochondrial carrier proteins: S7-K105, Y112-T202
Potential Glycosyl- ation Sites				N133 N144 N233 N298 N420 N566 N569 N763 N1054 N1245					<u>a</u> 32
Potential Phosphoryl- ation Sites				S86 S212 S235 S300 S366 S401 S518 S548 S608 S660 S719 S858 S988 S1011 S1039 S1040 S1049 S1056 S1087 S1114 S1166 S1224 S1234 S1323 S1399 S1410 T9 T422 T449 T450 T629 T637 T742 T629 T637 T742 T1268 T1297 T1387 T1389 T1449 T1462					S28 S142 S159 T144
Amino Acid Residues				1466					222
Incyte Polypep- tide ID				6046484CD1					7481427CD1
NO IN	თ			10					11

EQ ID NO:	Incyte Polypep- tide ID	Amino Acid Residues	Amino Potential Acid Phosphoryl- Residues ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
11					Mitochondrial energy transfer proteins BL00215: L13-Q37, I158-G170	BLIMPS_BLOCKS
					Mitochondrial energy transfer proteins mitoch_carrier.prf: C110-I158	PROFILESCAN
					Mitochondrial energy transfer proteins: P127-A136	MOTIFS
		·			Mitochondrial carrier protein signature PR00926: G120-D138, Y168-F186, D11-T24, T24-M38, G73-D93	BLIMPS_PRINTS
		i.			Adenine nucleotide transfer protein PR00927: F8-A20, C51-R72, T84-K96, R111- V124, R146-L167, S207-S222	BLIMPS_PRINTS
					PROTEIN TRANSPORT TRANSMEMBRANE REPEAT MITOCHONDRION CARRIER MEMBRANE INNER MITOCHONDRIAL ADP/ATP PD000117: S7-A123, Y112-S222	BLAST_PRODOM
	· ·				MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026   P02722   11-96; F12-198 DM00026   S31935   14-108; F12-T107 DM00026   P02722   116-205; L117-N201 DM00026   S31935   110-208; Q108-N201	BLAST_DOMO
					Transmembrane domains: I164-R187 N-terminus is cytosolic	ТМАР
12	7483595CD1	461	S168 S212 S233 S338 S362 S382 S439 T318 T348	N98 N163 N288 N344 N380 N381	SULFATE TRANSF TRANSMEMBRANE SULPHATE HIGH PD001755: R216	BLAST_PRODOM
					SULFATE TRANSPORTERS DM01229 P40879 5-462: T23-A140, Q124-L222 DM01229 P45380 10-468: P130-L222, T23-L138 DM01229 P50443 49-505: I136-K247, T23-S144	BLAST_DOMO
·		¥			Transmembrane domains: F6-F26, L31-S51, M62-S82, C91-A111, E128-K156, L161-R186 N-terminus is cytosolic	TMAP
13	3788427CD1	502	S146 S304 S446 S467 T25 T59 T96 T104 T164 T385 T492 T501		Mitochondrial carrier proteins: 8361-Y461, S266-Q359, T172-H264	HMMER_PFAM

Analytical Methods and Databases		s: MOTIFS	re BLIMPS_PRINTS	T BLAST_PRODOM	S BLAST_DOMO	3- TMAP	SPSCAN	5, M1- HMMER	g HMMER_PFAM	TMAP	BLIMPS_BLOCKS	3 BLAST_DOMO
Signature Sequences, Domains and Motifs	Mitochondrial energy transfer proteins mitoch_carrier.prf1: L362-1417 mitoch_carrier.prf2: S270-1315	Mitochondrial energy transfer proteins P287-1296	Mitochondrial carrier protein signature PR00926: G232-R252, V280-Q298, Y325-L343 V369-Q391	PROTEIN TRANSPORT TRANSMEMBRANE REPEAT MITOCHONDRION CARRIER MEMBRANE INNER MITOCHONDRIAL ADP/ATP PD000117: Y305-R453	MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026   Q01888   126-214: G263-1352 DM00026   P29518   233-310: I271-I352	Transmembrane domains: R176-G204, A323- K351, L424-K452 N-terminus is cytosolic	ivaç	Signal Peptide: M1-P22, M1-A23, M1-A25, G27	Bacterial extracellular solute-binding protein domain: M1-L253	Transmembrane domains: T175-A196 N-terminus is non-cytosolic	Bacterial extracellular solute binding protein signature BL01039: G52-L72, R86-Y118, L90-S101	BACTERIAL EXTRACELLULAR SOLUTE-BINDING PROTEINS, FAMILY 3 DM00557   P27676   32-261: S19-K223 DM00557   P30860   5-241: L37-W252 DM00557   P39174   15-260: L8-W252
Potential Glycosyl- ation Sites					I I	I.	N82 N172 N173 s	07.0	<u>1</u>	T.	<u> </u>	
Potential Phosphoryl- ation Sites							S6 S26 S135 S198 T2 T36 T64 T84 T92 T165 Y218					
Amino Acid Residues				·			261					
Incyte Polypep- tide ID							6972455CD1			·		
EQ NO:	13						14					

Analytical Methods and	TWAP	BLAST_DOMO	TMAP	HMMER PFAM	BLIMPS_BLOCKS	PROFILESCAN	BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_PRODOM
Signature Sequences, Domains and Motifs	N11 N342 N440 Transmembrane domains: V163-V183 N194-T214 V232-1259 I275-R302 S353-N373 A382-G402 P465-G481 N-terminus is cytosolic	do SENSITIVE; COTRANSPORTER; SODIUM; CHLORIDE; DM01337   P55011   409-906: V278-G402 DM01337   P55013   381-879: V278-G402 DM01337   A53491   381-879: V278-G402 DM01337   P55014   297-795: V278-G402	mains: F2 422 D429- 964 n-cytosol	E1-E2 ATPase domain: V268-P324	E1-E2 ATPases phosphorylation site signature BL00154: G284-L301, V442-G478, I480-L498, K624-C634, N695-M735	El-E2 ATPases phosphorylation site: 1466- F515	P-type cation-transporting atpase superfamily signature PR00119: N309-T323, C484-L498, A711-D721	Sodium/potassium-transporting ATPase signature PR00121: C477-L498, V621-V639	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP BINDING TRANSPORT PUMP CALCIUM MAGNESIUM MEMBRANE PD000132: 1230- D494
Potential Glycosyl- ation Sites	N11 N342 N440	·	N540 N669 N781 N819 N848 N867 N875 N1005						
Potential Phosphoryl- ation Sites	S40 S82 S265 S314 S349 S442 S446 S551 T50 T324 T344		S5 S103 S159 N54 S241 S249 S338 N78 S567 S587 S671 N84 S1008 S1028 T78 T97 T172 T375 T490 T664 T701 T784 T859 T871						
Amino Raid	570		1033		*				. ,
Incyte Polypep- tide ID	8077668CD1		55120485CD1						
EQ ID NO:	15		16					-	

Analytical Methods and Databases	BLAST_DOMO	MOTIFS	HWMER_PFAM	TMAP	BLAST_PRODOM	TMAP	HMMER_PFAM	HMMER_PFAM
Signature Sequences, Domains and Motifs	E1-E2 ATPASES PHOSPHORYLATION SITE DM00115 P22189 49-801: Q597-P738, W196-K322, P389-P447, S671-L723 DM00115 P37278 58-755: I192-I743, V822-K840 DM00115 P47317 26-695: F600-E749, I230-K559, K815-E853 DM00115 P54707 97-825: I232-I743, L813-E853	E1-E2 ATPases phosphorylation site: D486- T492	Reduced folate carrier domain: S10-V441	Transmembrane domains: L8-M28 N53-Y76 Y79- Q107 F111-R138 L148-S168 V174-K194 K276- D304 N316-Y336 D342-L362 A367-A395 L405- V425 P434-L454 N-terminus is cytosolic	FOLATE CARRIER PROTEIN REDUCED TRANSPORTER GLYCOPROTEIN FOLATE BINDING TRANSPORT TRANSMEMBRANE METHOTREXATE PD003967: S11-E230, G327-S493, F262-W303	Transmembrane domains: G157-Q185 V189-R217 A242-G268 K269-M297 V314-S340 E343-K369 L377-D399 L407-S433 A469-L489 G493-D521 G531-H559 N-terminus is cytosolic	Divalent cation transporter domain: 1199-8335, Y413-H559	N294 Anion-transporting ATPase domain: ) L354-S573
Potential Glycosyl- ation Sites			N45 N166 N256			N78 N466		N80 N174 N294 N491 N529 N558
Potential Phosphoryl- ation Sites			\$40 \$211 \$241 \$251 \$274 \$275 \$463 \$466 \$482 \$493 T47 T117			S8 S35 S77 S88 S106 S137 S229 S304 S321 S340 S379 T10 T27 T148 T194 T401	•	S69 S138 S198 S221 S261 S355 S473 S478 S509 S543 T24 T159 T176 T401 T406 T474 T499
Amino Acid Residues			496			573		573
Incyte Polypep- tide ID			3112883CD1			4253888CD1		7479974CD1
S C S S S C S	16		17			18		19

							70	70	Iro		
Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	MOTIFS	ТМАР	HMMER_PFAM	TMAP	BLIMPS_BLOCKS	BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_PRODOM
	PLASMID ARSENICAL PUMPDRIVING ATPASE HYDROLASE RESISTANCE ATPBINDING ARSA PD006335: D460-P568	NIFH/FRXC FAMILY DM00105 P08690 7-180: Y20-E191 DM00105 P08690 326-473: L333-K472 DM00105 P30632 17-190: K19-S167	ATP/GTP-binding site motif A (P-loop): G25-T32, G337-T344	Transmembrane demains: D8-S36, R44-I72, C96-R123, S133-I158, L171-F194 N-terminus is non-cytosolic	POT family (proton/oligopeptide symporter) domain: G56-N141	Transmembrane domains: P84-K112, Y459-N487, P528-Y553, G564-K584, L592-C612, I624-Y652 N-termincus is cytosolic	E1-E2 ATPases phosphorylation site proteins: BL00154: D231-L271, T386-S409, K131-L141	P-type cation-transporting atpase superfamily signature PR00119: A247-D257	H+-transporting ATPase (proton pump) signature PR00120: T162-A180	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP-BINDING PROTEIN PROBABLE CALCIUM TRANSPORTING CALCIUM TRANSPORT PD004657: A423-P661	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP-BINDING PROBABLE PROTEIN CALCIUM TRANSPORTING CALCIUM TRANSPORT PD149930: C363-F422
Potential Glycosyl- ation Sites						N700 N732	•				
Potential Phosphoryl- ation Sites				S42 S85 S121 S198 S215 S227 S233 S239		S5 S62 S109 S115 S185 S312 S409 S476 S556 S706 S734 T28 T30 T78 T162 T201 T227 T335 T534 T674 T695	*	į			: :
Amino Acid Residues				248		761					
Incyte Polypep- tide ID				7483850CD1		5508353CD <u>1</u>	·				
8 6 1 8	19			20	:	21					

EQ ID NO:	Incyte Polypep- tide ID	Amino Acid Residues	Potential Phosphoryl- ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
21					CALCIUM;  P32660 31	BLAST_DOMO
					ATPASE; CALCIUM; TRANSPORTING; DM02405 Q09891 206-1107: L56-N487	BLAST_DOMO
			٠		ATPASE; CALCIUM; TRANSPORTING; DM02405 S51243 356-1267: E59-F486	BLAST_DOMO
					ATPASE; CALCIUM; TRANSPORTING; DM02405 P39524 236-1049: Y58-D287, L332-N487	BLAST_DOMO
					ATP/GTP-binding site motif A (P-loop): G331-T338, A699-S706	MOTIFS
22	8543628CD1	219	S4 T148		Binding-protein-dependent transport system: A112-Y185	HMMER_PFAM
					Transmembrane domains: A20-N48, V70-S98, T156-R183, Y185-A208 N-terminus is non-cytosolic	TMAP
				•	Binding-protein-dependent transport systems inner membrane component: V105-T161	PROFILESCAN
					PROTEIN TRANSPORT TRANSMEMBRANE PERMEASE MEMBRANE AMINO ACID INNER SYSTEM TRANSPORTER ABC PD001196: E7-G111	BLAST_PRODOM
					BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388 P10345 3-214: L9-R215	SYSTEMS BLAST_DOMO
					-PROTEIN-DEPENDENT TRANSPORT SYSTEMS EMBRANE DM00388   P42399   12-220: 5	BLAST_DOMO
			·		BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388 P45023 17-232: P14-R215	BLAST_DOMO
					PROTEIN-DEPENDENT TRANSPORT MBRANE DM00388   P42200   15-226	SYSTEMS BLAST_DOMO
					protein-dependent transport systems ubrane component signature:	MOTIFS
23	7482754CD1	463	S249 S254 S413 S420 T451 T458		Transmembrane amino acid transporter protein: A48-C433	HMMER_PFAM

Analytical Methods and Databases	TMAP	BLIMPS_PRINTS	BLAST_PRODOM	HMMER						SPSCAN	HMMER_PFAM	TMAP	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_PRODOM	
Signature Sequences, Domains and Motifs	Transmembrane domains: L24-N44, G54-V74, R93-R121, D148-R168, H180-T200, S249-S274, G298-R326, W353-M373, S377-P397 N-terminus is non-cytosolic	Transmembrane four family signature PR00259: F59-V82, L73-A99, V302-V328	ACID AMINO PROTEIN TRANSPORTER PERMEASE TRANSMEMBRANE INTERGENIC REGION PUTATIVE PROLINE PD001875: F31-1320	N344 N451 Signal Peptide: N609 M1-G21						signal_cleavage: M1-G22	Ligand-gated ion channel: H574-E852	Transmembrane domains: H574-R598, P640-V667, T823-L847	14	R32184_2 IONOTROPIC GLUTAMATE RECEPTOR PD156309: A77-Y477	RECEPTOR GLUTAMATE SUBUNIT SIGNAL PRECURSOR CHANNEL IONIC TRANSMEMBRANE POSTSYNAPTIC	FDUUU50U: M5/0-E852
Potential Glycosyl- ation Sites				N69 N344 N451 N465 N609	N786											
Potential Phosphoryl- ation Sites				s153 s388	S536 S633 S700 S709 S721 S747	S867	6 T157	T421 T604	T641 T673 T696 T725 T916							
Amino Poten Acid Phosp Residues ation				1043											Y	·
Incyte Polypep- tide ID				3794818CD1		,					·					
EQ ID NO:	23			24												

EQ ID NO:	Incyte Polypep- tide ID	Amino Acid Residues	Amino Potential Acid Phosphoryl- Residues ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
24					RECEPTOR SIGNAL GLUTAMATE SUBUNIT PROTEIN TRANSMEMBRANE CHANNEL IONIC PD000273: G478-A563, G728-V817	BLAST_PRODOM
					GLUTAMATE RECEPTOR DM00247 003391 640-919: T631-S901, C964-P980 DM00247 P35436 615-886: T631-F856 DM00247 Q01098 613-882: T631-E904 DM00393 Q01097 377-614: G387-F628	BLAST_DOMO
25	4717525CD1	480	S4 S23 S56 S105 S176 S411 S418 T161 T170 T220 T302 T410 T469	·	Mitochondrial carrier proteins: M184-T276, G319-L413	HMMER_PFAM
					EF hand: Q117-H145, R13-L41, R81-L109	HMMER_PFAM
					Mitochondrial energy transfer proteins BL00215: V190-Q214, I369-G381	BLIMPS_BLOCKS
					Mitochondrial energy transfer proteins signature: I320-S371, K187-L241, V279-S331	PROFILESCAN
	·			·	Mitochondrial carrier protein signature PR00926: Q188-T201, T201-V215, G244-E264, C333-Q351, Y379-Y397, G327-Q349	BLIMPS_PRINTS
					Graves disease carrier protein signature PR00928: Q274-Q294, P205-I225, Y263-S287, I369-V389	BLIMPS_PRINTS
				, [7	TRANSPORT TRANSMEMBRANE CARRIER INNER MITOCHONDRIAL ADP/ATP PD000117: K187-S466	BLAST_PRODOM
					MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026 S57544 26-107: V190-I270 DM00026 Q01888 38-124: V190-I270	BLAST_DOMO
					EF-hand calcium-binding domain: D22-L34 D90-I102	MOTIFS

## Prosports
38 S161 N6 N171 N371 TE S253 S331 N376 S510 T378 S510 T378 S329 S421 T418 S329 S421 S171 S305 N52 N388 N455 HC S442 S469 N463 T704 Y791 TT
S510 T378  S66 S263 S329 S421 T418 T418  S171 S305 S42 S469 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S66 S263 N53 N62 N68 Tr S329 S421 T418 5171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S66 S263 N53 N62 N68 Tr T418 T418 S171 S305 N52 N388 N455 HC S442 S469 N463 T174 T198 T400 T420 T704 Y791
S66 S263 N53 N62 N68 Tr S329 S421 T418 3171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S329 S421 T418  T418  S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791 Tr
7418  5171 \$305 N52 N388 N455 HC  5442 \$469 N463  T6 T20 T56  T174 T198  T400 T420  T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S171 S305 N52 N388 N455 HC S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791
S305 N52 N388 N455 HC 2 S469 N463 120 T56 4 T198 1 T291 1 Y791
S305 N52 N388 N455 HC 2 S469 N463 120 T56 4 T198 1 T291 1 Y791 TX791
S442 S469 N463 T6 T20 T56 T174 T198 T400 T420 T704 Y791 T704 Y791
Tb T20 T50 T400 T420 T704 Y791 Transmembrane V282-G304, I403-I423, S564-L592,
T704 Y791 Transmembrane V282-G304, I403-L423, S564-L592, T704 Y795
T704 Y791  Transmembrane
Transmembrane V282-G304, I403-I423, S564-L592,
7001
0
N terminus is non-cytosolic

SEO	Incvte	Amino	Potential	Potential	Signature Sequences,	Analytical
Ĥ	Polypep-	Acid	Acid Phosphoryl-	Glycosyl-		Methods and
200	מדק הדק	Connectivity	מכדסוו סדרכם	מרדמו חדרפם	Arion contract family	Databases
0					ALION EXCHAINELS LAMLLY PORT TO TO TO TO TO THE PART OF THE PART TO THE PART OF THE PART O	BLIMES BLOCKS
					1342-F380, A382-Y429, O481-S534, A567-W608,	
					D609-E647, H653-F698, Y700-T743, L748-I787	
					Anion exchanger signature	BLIMPS_PRINTS
					PR00165: G253-I275, K284-G304, A371-S390,	
					A482-L501, I517-G537	
					ANION EXCHANGE TRANSMEMBRANE GLYCO-PROTEIN	BLAST_PRODOM
					LIPOPROTEIN PALMITATE BICARBONATE	
					COTRANSPORTER	
					PD001455: T483-T743, F752-S457,	
					R158-A237	
						BLAST_DOMO
					DM02294   P48751   601-1229: A190-E798	
					DM02294 P02730 311-908: T483-E798	
					DM02294 A42497 403-1027: F252-E798	
					DM02294 P04920 602-1237: D248-E798	
29	6972530CD1	344		N223 N299	signal_cleavage:	SPSCAN
_			<b>S333</b>		M1-A34	
			T145 T196 T286 Y330			
					Signal Peptide:	HMMER
					M9-A34	
						TMAP
					R8-L36 V39-V63 Q71-T99	
					N terminus is non-cytosolic.	

Analytical	Methods and	Ta canana	TMAP										MOTIFS										
Signature Sequences,	Domains and Motifs		N56 N218 N319 Transmembrane domains:	I965-S986, E1281-T1300,	N1670-S1692, V1836-W1864,	V2276-N2292, A2507-F2534	N terminus is non-cytosolic.						Adenosine and AMP deaminase signature	S2386-P2392									
Potential	Glycosyl-	acion Sices	N56 N218 N319	N330 N364	N396 N452	N480 N539	N607 N667	N691 N772	N828 N875	N885 N921	N949 N964	1922 S307 N988 N1135	N1153 N1166	N1241 N1259	N1273 N1305	N1323 N1372	N1479 N1525	N1528 N1618	N1624 N1892	N1969 N2134	N2342 N2428	N2510 N2596	N2625
Potential	Phosphory1-	ation Sites	S1063 S1231	S1270 S40 S1367 N330 N364	S1526 S551	S1603 S193	S1710 S1793	S223 S1824	S1866 S274	S1879 S230	S1893 S1915		S1952 S1978	S461 S2029 S382 N1241	S2045 S2090	S2230 S495	S2388 S2438	S792 S2464 S790 N1528	S2488 S2496	S2506 S901	S2537 S809	S2545 S2573	S856 S2631
Amino	Acid Phospho	Residnes	2701																				
Ĭ	Polypep-		덥																-				
EO	<u>'a</u>	NO:	30				-	_															

Analytical Methods and Databases	BLAST_DOMO	HMMER	HMMER_PFAM	TMAP	BLIMPS_BLOCKS	PROFILESCAN
Signature Sequences, Domains and Motifs	Sodium/Calcium Exchanger Chain DM05297 P48765 6-969: V263-1492 P=2.8-09	Signal Peptide: M47-P73	Sodium: solute symporter family: F45-G449	Transmembrane domains: P4-Y32, T53-G73, I84-L104, L121-Y148, G159-L179, V190-M210, R239-I256, R273-L301, S385-L405, A412-I432, S439-G459, N515-T543 N terminus is non-cytosolic.	Sodium: solute symporter BL00456: L22-S76, 198-V127, T158-G212	
Potential Glycosyl- ation Sites		N260 N481 N485 N606				
Si	T122 T158 T276 T417 T581 T774 T881 T989 T1004 T1030 T1045 T1051 S941 T1058 T1090 S953 T1307 T1559 T1610 T1658 T1786 T1887 T1945 T2171 T2213 T2266 T2328 T2409 T2415 T2459 T2459	S114 S269 S317 S375 S377 S563 S576 S602 T3 T41 T53 T74 T158 T312 T364 T483 T491 T557				
Amino Potent Acid Phosph Residues ation		610				
Incyte Polypep- tide ID		71726948CD1		-		
	O M	31				

ID Polypep- Acid Phosphoryl- NO: tide ID Residues ation Sites 31  32 7487393CD1 552 846 S60 S68 S282 S408 S47 S537 T58 T133 T311 T323 T39 T526	phoryl- Glycosyl- n Sites ation Sites	Domains and Motifs  TRANSMEMBRANE TRANSPORT PERMEASE SODIUM SYMPORT PROLINE GLYCOPROTEIN PD000991: F45-P234, L129-G449 SYMPORTER SODIUM IODIDE THYROID PD024705: A451-P552 SODIUM: SOLUTE SYMPORTER FAMILY DM00745 J7C2382   3-485:Y15-W455 DM00745 P45174   3-495:T9-W455 DM00745 P31448   1-494:F18-G449	Methods and Databases BLAST_PRODOM BLAST_PRODOM BLAST_PRODOM
7487393CD1 552 846 St S282 8 S237 T T311 T T526	ה ה ה	SODIUM	BLAST_PRODOM BLAST_PRODOM BLAST_DOMO
7487393CD1 552 S46 S( S282 8 S537 7 T311.7 T311.7 T526			BLAST_PRODOM BLAST_DOMO
7487393CD1 552 S46 S6 S143 S S282 S237 T T T T T T T T T T T T T T T T T T T			BLAST_PRODOM BLAST_DOMO
7487393CD1 552 S46 S6 S143 S S282 S S282 S S537 T T T T T T T T T T T T T T T T T T T			BLAST_PRODOM BLAST_DOMO
7487393CD1 552 846 S6 S143 S S282 S282 S537 T T311 T T526			BLAST_DOMO
7487393CD1 552 846 S6 S143 S S282 S S282 S S237 T T T T T T T T T T T T T T T T T T T			BLAST_DOMO
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Table 5

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37	4787433CB1	PGANNOT01
38	7483598CB1	BRAITUT29
39	7484823CB1	TESTNOC01
40	143935CB1	BRACDIK08
41	5923789CB1	BRAIFET02
42	6046484CB1	BRACNOK02
44	7483595CB1	TESTNOC01
45	3788427CB1	BONEUNR01
46	6972455CB1	BMARUNR02
47	8077668CB1	ADRETUE02
48	55120485CB1	BRAITUT29
49	3112883CB1	BRSTNOT03
50	4253888CB1	ADRETUE02
52	7483850CB1	LIVRDIT06
	5508353CB1	NERDTDN03
	8543628CB1	BMARUNR02
	7482754CB1	PTHYTMN05
56	3794818CB1	KIDEUNE02
57	4717525CB1	KIDEUNE02
58	5091793CB1	LUNGTUT08
59	5945527CB1	SINTNOR01
	6941124CB1	FTUBTUR01
61	6972530CB1	BMARUNR02
	6991750CB1	BRAIFER06
63	71726948CB1	KIDNNOT32

Library	Vector	Library Description
ADRETUE02	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from right adrenal tumor tissue removed from a 49-year-old Caucasian male during unilateral adrenalectomy. Pathology indicated adrenal cortical carcinoma comprising nearly the entire specimen. The tumor was attached to the adrenal gland which showed mild cortical atrophy. The tumor was encapsulated, being surrounded by a thin (1-3 mm) rim of connective tissue. The patient presented with adrenal cancer, abdominal pain, pyrexia of unknown origin, and deficiency anemia. Patient history included benign hypertension. Previous surgeries included adenotonsillectomy. Patient medications included aspirin, calcium, and iron. Family history included atherosclerotic coronary artery disease in the grandparent(s).
BMARUNR02 PIGEN	PIGEN	This random primed library was constructed using RNA isolated from an untreated SH-SY5Y cell line derived from bone marrow neuroblastoma tumor cells removed from a 4-year-old Caucasian female.
BONEUNROI	PCDNA2.1	This random primed library was constructed using pooled cDNA from two different donors. cDNA was generated using mRNA isolated from an untreated MG-63 cell line derived from an osteosarcoma tumor removed from a 14-year-old Caucasian male (donor A) and using mRNA isolated from sacral bone tumor tissue removed from an 18-year-old Caucasian female (donor B) during an exploratory laparotomy and soft tissue excision. Pathology indicated giant cell tumor of the sacrum in donor B. Donor B's history included pelvic joint pain, constipation, urinary incontinence, unspecified abdominal/pelvic symptoms, and a pelvic soft tissue malignant neoplasm. Family history included prostate cancer in donor B.
BRACDIK08	PSPORT1	This amplified and normalized library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 57-year-old Caucasian male who died from a cerebrovascular accident. Serologies were negative. Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day for 40 years).

#### Table (

Library	Vector	Library Description
BRACNOK02	PSPORT1	This amplified and normalized library was constructed using RNA isolated from posterior cingulate tissue removed from an 85-year-old Caucasian female who died from myocardial infarction and retroperitoneal hemorrhage. Pathology indicated atherosclerosis, moderate to severe, involving the circle of Willis, middle cerebral, basilar and vertebral arteries; infarction, remote, left dentate nucleus; and amyloid plaque deposition consistent with age. There was mild to moderate leptomeningeal fibrosis, especially over the convexity of the frontal lobe. There was mild generalized atrophy involving all lobes. The white matter was mildly thinned. Cortical thickness in the temporal lobes, both maximal and minimal, was slightly reduced. The substantia nigra pars compacta appeared mildly depigmented. Patient history included COPD, hypertension, and recurrent deep venous thrombosis. 6.4 million independent clones from this amplified library were normalized in one round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791.
BRAHTDK01	PSPORT1	This amplified and normalized library was constructed using pooled RNA isolated from archaecortex, anterior and posterior hippocampus tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver. 7.6x10e5 independent clones from this amplified library were normalized in 1 round using conditions adapted Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used
BRAIFER06	PCDNA2.1	This random primed library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. Serologies were negative.
BRAIFET02	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus, who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAITUT29	pINCY	Library was constructed using RNA isolated from brain tumor tissue removed from the parietal lobe of a 43-year-old female during excision of a cerebral meningeal lesion. Pathology indicated high grade glioma. Family history included acute myocardial infarction, atherosclerotic coronary artery disease, benign hypertension, and hyperlipidemia.

Library	Vector	Library Description
BRSTNOT03	PSPORT1	Library was constructed using RNA isolated from diseased breast tissue removed from a 54-year-old Caucasian female during a bilateral radical mastectomy. Pathology for the associated tumor tissue indicated residual invasive grade 3 mammary ductal adenocarcinoma. Patient history included kidney infection and condyloma acuminatum. Family history included benign hypertension, hyperlipidemia and a malignant neoplasm of the colon.
FTUBTUR01	PCDNA2.1	This random primed library was constructed using RNA isolated from fallopian tube tumor tissue removed from an 85-year-old Caucasian female during bilateral salpingo-oophorectomy and hysterectomy. Pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenocarcinoma, which was confined to the mucosa without mural involvement. Endometrioid carcinoma in situ was also present. Pathology for the associated uterus tumor indicated focal endometrioid adenocarcinoma in situ and moderately differentiated invasive adenocarcinoma arising in an endometrial polyp. Metastatic endometrioid and serous adenocarcinoma was present at the cul-de-sac tumor. Patient history included medullary carcinoma of the thyroid and myocardial infarction
KIDEUNE02	pINCY	This 5' biased random primed library was constructed using RNA isolated from an untreated transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue (Invitrogen). The cells were transformed with adenovirus 5 DNA.
KIDNNOT32	pINCY	Library was constructed using RNA isolated from kidney tissue removed from a 49-year-old Caucasian male who died from an intracranial hemorrhage and cerebrovascular accident. Patient history included tobacco abuse.
LIVRDIT06	pINCY	This library was constructed using RNA isolated from diseased liver tissue removed from a 35-year-old Caucasian male during needle biopsy of the liver. Patient history included hepatitis C.
LIVRNON08	pINCY	This normalized library was constructed from 5.7 million independent clones from a pooled liver tissue library. Starting RNA was made from pooled liver tissue removed from a 4-year-old Hispanic male who died from anoxia and a 16 week female fetus who died after 16-weeks gestation from anencephaly. Serologies were positive for cytolomegalovirus in the 4-year-old. Patient history included taking daily prenatal vitamins and mitral valve prolapse in the mother of the fetus. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

#### Table (

Library	Vector	Library Description
ruros	pINCY	Library was constructed using RNA isolated from lung tumor tissue removed from a 63-year-old Caucasian male during a right upper lobectomy with fiberoptic bronchoscopy. Pathology indicated a grade 3 adenocarcinoma. Patient history included atherosclerotic coronary artery disease, an acute myocardial infarction, rectal cancer, an asymtomatic abdominal aortic aneurysm, tobacco abuse, and cardiac dysrhythmia. Family history included congestive heart failure, stomach cancer, and lung cancer, type II diabetes, atherosclerotic coronary artery disease, and an acute myocardial infarction.
NERDTDN03	PINCY	This normalized dorsal root ganglion tissue library was constructed from 1.05 million independent clones from a dorsal root ganglion tissue library. Starting RNA was made from dorsal root ganglion tissue removed from the cervical spine of a 32-year-old Caucasian male who died from acute pulmonary edema, acute bronchopneumonia, bilateral pleural effusions, pericardial effusion, and malignant lymphoma (natural killer cell type). The patient presented with pyrexia of unknown origin, malaise, fatigue, and gastrointestinal bleeding. Patient history included probable cytomegalovirus infection, liver congestion, and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, respiratory failure, pneumonia of the left lung, natural killer cell lymphoma of the pharynx, Bell's palsy, and tobacco and alcohol abuse. Previous surgeries included colonoscopy, closed colon biopsy, adenotonsillectomy, and nasopharyngeal endoscopy and biopsy. Patient medications included Diflucan (fluconazole), Deltasone (prednisone), hydrocodone, Lortab, Alprazolam,
·		Reazodone, ProMace-Cytabom, Etoposide, Cisplatin, Cytarabine, and dexamethasone. The patient received radiation therapy and multiple blood transfusions. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
PGANNOT01 PSPORT	PSPORT1	Library was constructed using RNA isolated from paraganglionic tumor tissue removed from the intra-abdominal region of a 46-year-old Caucasian male during exploratory laparotomy. Pathology indicated a benign paraganglioma and was associated with a grade 2 renal cell carcinoma, clear cell type, which did not penetrate the capsule. Surgical margins were negative for tumor.

Library	Vector	Library Description
PROSTUS19	pINCY	This subtracted prostate tumor tissue library was constructed using 2.36 million clones from the PROSTUT13 library and
		was subjected to two round sof subtraction hybridization with 2.36 million clones from EPIPNOT01 library. The starting library for subtraction was constructed using RNA isolated from prostate tumor tissue removed from a 59-year-old
		Caucasian male during a radical prostatectomy with regional lymph node excision. Pathology indicated adenocarcinoma
	,	(Creason grace 3+5) involving the prostate peripherany with invasion of the capsule. Adenonocinations hyperplasta was present. The patient presented with elevated prostate-specific antigen (PSA). Patient history included diverticulitis of colon,
		asbestosis, and thrombophlebitis. Family history included benign hypertension, multiple myeloma, hyperlipidemia, and rheumatoid arthritis. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991)
		19:1954 and Bonaldo, et al. Genome Research (1996) 6:791.
PTHYTMN05	pINCY	Library was constructed using RNA isolated from parathyroid tissue removed from a 44-year-old Caucasian male during a
		partial paramyrouectomy, ramotogy for the materied tumor ussue indicated paramyroud caremonna (grade 1 of 4) forming a partially cystic tan mass. Both capsular and vascular invasion were present. The patient presented with unspecified
		parathyroid disorder and calcium metabolism disorder. Patient history included kidney calculus and obesity. Previous
	<u> </u>	surgeries included vasectomy and parathyroid surgery. Family history included emphysema in the mother; type II diabetes in the father: and type I diabetes and hyperlinidemia in the sibling(s).
		in the state of th
SINTNOR01	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine tissue removed from a 31-year-old
		Caucasian female during Koux-en- 1 gastric bypass. Patient history included clinical obesity.
TESTNOC01	PBLUESCRIPT	TESTNOC01   PBLUESCRIPT   This large size fractionated library was constructed using RNA isolated from testicular tissue removed from a pool of
		eleven, 10 to 61-year-old Caucasian males.

Parameter Threshold		Mismatch <50%		ESTs: Probability value= 1.0B-8 or less Full Length sequences: Probability value= 1.0B-10 or less	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater, fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater	Probability value= 1.0E-3 or less	PFAM hits: Probability value= 1.0B-3 or less Signal peptide hits: Score= 0 or greater
Reference	Applied Biosystems, Foster City, CA.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Applied Biosystems, Foster City, CA.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.
Description	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	A program that assembles nucleic acid sequences.	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, fasta, fastx, tfastx, and ssearch.	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.
Program	ABI FACTURA	ABIPARACEL FDF	ABI AutoAssembler	BLAST	FASTA	BLIMPS	HMMER

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Bnzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25.217-221	Normalized quality scorez GCG-specified "HIGH" value for that particular Prosite motif.  Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability	Ewing, B et al. (1993) Genome Res. 8 175-185, Ewing, B and P. Green. (1993) Genome Res. 8 186-194	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, TF and M.S. Waterman (1981) Adv. Appl. Math. 2.482-489, Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater, Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan .	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, B.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	7-221; age

#### What is claimed is:

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1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32,
- a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32,
- c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
- 2. An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
  - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
  - 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64.
  - A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
    - 7. A cell transformed with a recombinant polynucleotide of claim 6.
    - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
    - 9. A method of producing a polypeptide of claim 1, the method comprising:
    - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

- b) recovering the polypeptide so expressed.
- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

11. An isolated antibody which specifically binds to a polypeptide of claim 1.

- 12. An isolated polynucleotide selected from the group consisting of:
- a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
   90% identical to a polynucleotide sequence selected from the group consisting of
   SEQ ID NO:33-64,
- c) a polynucleotide complementary to a polynucleotide of a),
- d) a polynucleotide complementary to a polynucleotide of b), and
  - e) an RNA equivalent of a)-d).
  - 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.
  - 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
  - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
  - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
  - 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
  - 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- a) amplifying said target polynucleotide or fragment thereof using polymerase chain

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reaction amplification, and

- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 5 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
  - 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
  - 19. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 17.
- 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
  - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
  - b) detecting agonist activity in the sample.
- 21. A composition comprising an agonist compound identified by a method of claim 20 and pharmaceutically acceptable excipient.
  - 22. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 21.
  - 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
    - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- detecting antagonist activity in the sample.
  - 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
- 35 25. A method for treating a disease or condition associated with overexpression of

functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 24.

- 26. A method of screening for a compound that specifically binds to the polypeptide of 5 claim 1, the method comprising:
  - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
  - b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
  - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
  - 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
  - a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
  - b) detecting altered expression of the target polynucleotide, and
  - c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
    - 29. A method of assessing toxicity of a test compound, the method comprising:
    - a) treating a biological sample containing nucleic acids with the test compound,
    - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under

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conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,

- c) quantifying the amount of hybridization complex, and
  - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

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- 30. A diagnostic test for a condition or disease associated with the expression of TRICH in biological sample, the method comprising:
  - combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
  - b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
  - 31. The antibody of claim 11, wherein the antibody is:
- a) a chimeric antibody,
  - b) a single chain antibody,
  - c) a Fab fragment,
  - d) a F(ab')<sub>2</sub> fragment, or
  - e) a humanized antibody.

- 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
- 33. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 32.
  - 34. A composition of claim 32, wherein the antibody is labeled.
- 35. A method of diagnosing a condition or disease associated with the expression of in a subject, comprising administering to said subject an effective amount of the

composition of claim 34.

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36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:

- a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response.
  - b) isolating antibodies from said animal, and
- screening the isolated antibodies with the polypeptide, thereby identifying a

  polyclonal antibody which binds specifically to a polypeptide comprising an amino
  acid sequence selected from the group consisting of SEQ ID NO:1-32.
  - 37. A polyclonal antibody produced by a method of claim 36.
- 15 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
  - 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
    - immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
      - b) isolating antibody producing cells from the animal,
      - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
- 25 d) culturing the hybridoma cells, and
  - e) isolating from the culture monoclonal antibody which binds specifically to a
    polypeptide comprising an amino acid sequence selected from the group consisting
    of SEQ ID NO:1-32.
- 30 40. A monoclonal antibody produced by a method of claim 39.
  - 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
  - 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.

43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

- 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in a sample, the method comprising:
- a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in the sample.
- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 from a sample, the method comprising:
- incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
- 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 13.
  - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
  - a) labeling the polynucleotides of the sample,
    - contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
    - c) quantifying the expression of the polynucleotides in the sample.
  - 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is polynucleotide of claim 12.

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49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.

- 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
- 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
  - 52. An array of claim 48, which is a microarray.
- 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
  - 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
- 20 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.

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- 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
- 57. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:2.
- 30 58. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:3.
  - 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
  - 60. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:5.

	61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
	62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
5	63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
	64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
10	65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
	66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.
	67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.
15	68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13.
	69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14.
20	70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15.
	71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16.
	72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17.
25	73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18.
	74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19.
30	75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20.
	76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21.
	77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22.
35	78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23.

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	79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24.
	80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25.
5	81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26.
	82. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27.
10	83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28.
10	84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29.
	85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30.
15	86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31.
	87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32.
20	88. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:33.
	89. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:34.
25	90. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:35.
	91. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:36.
30	92. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:37.

93. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

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NO:38.

NO:50.

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	94. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:39.
	95. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:40.
	96. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:41.
	97. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:42.
	98. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:43.
:	99. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:44.
•	100. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:45.
	101. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:46.
	102. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:47.
	103. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:48.
	104. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:49.
	105. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

106. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:51.
107. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:52.
108. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:53.
109. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:54.
110. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:55.
111. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:56.
112. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:57.

- 113. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:58.
- 25 114. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:59.
  - 115. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:60.
  - 116. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:61.
  - 117. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:62.

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118. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:63.

119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.

# **CORRECTED VERSION**

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(54) Title: TRANSPORTERS AND ION CHANNELS

(57) Abstract: The invention provides human transporters and ion channels (TRICH) and polynucleotides which identify and encode TRICH. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with aberrant expression of TRICH.

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#### TRANSPORTERS AND ION CHANNELS

This invention relates to nucleic acid and amino acid sequences of transporters and ion channels and to the use of these sequences in the diagnosis, treatment, and prevention of transport, neurological, muscle, immunological and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

#### **BACKGROUND OF THE INVENTION**

hydrophobic lipid bilayer membranes which are highly impermeable to most polar molecules. Cells and organelles require transport proteins to import and export essential nutrients and metal ions including K\*, NH<sub>4</sub>\*, P<sub>i</sub>, SO<sub>4</sub><sup>2</sup>, sugars, and vitamins, as well as various metabolic waste products. Transport proteins also play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). Transport can occur by a passive concentration-dependent mechanism, or can be linked to an energy source such as ATP hydrolysis or an ion gradient. Proteins that function in transport include carrier proteins, which bind to a specific solute and undergo a conformational change that translocates the bound solute across the membrane, and channel proteins, which form hydrophilic pores that allow specific solutes to diffuse through the membrane down an electrochemical solute gradient.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with

25 simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems driven by the sodium gradient that exists across the plasma membrane. Sodium moves into the cell down its electrochemical gradient and brings the solute into the cell with it. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na\*/K\* ATPase system. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). All three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging techniques and for specific targeting of radioisotopes to the thyroid gland (Levy,

O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

One of the largest families of transporters is the major facilitator superfamily (MFS), also 5 called the uniporter-symporter-antiporter family. MFS transporters are single polypeptide carriers that transport small solutes in response to ion gradients. Members of the MFS are found in all classes of living organisms, and include transporters for sugars, oligosaccharides, phosphates, nitrates, nucleosides, monocarboxylates, and drugs. MFS transporters found in eukaryotes all have a structure comprising 12 transmembrane segments (Pao, S.S. et al. (1998) Microbiol. Molec. Biol. 10 Rev. 62:1-34). The largest family of MFS transporters is the sugar transporter family, which includes the seven glucose transporters (GLUT1-GLUT7) found in humans that are required for the transport of glucose and other hexose sugars. These glucose transport proteins have unique tissue distributions and physiological functions. GLUT1 provides many cell types with their basal glucose requirements and transports glucose across epithelial and endothelial barrier tissues; GLUT2 15 facilitates glucose uptake or efflux from the liver; GLUT3 regulates glucose supply to neurons; GLUT4 is responsible for insulin-regulated glucose disposal; and GLUT5 regulates fructose uptake into skeletal muscle. Defects in glucose transporters are involved in a recently identified neurological syndrome causing infantile seizures and developmental delay, as well as glycogen storage disease, Fanconi-Bickel syndrome, and non-insulin-dependent diabetes mellitus (Mueckler, 20 M. (1994) Eur. J. Biochem. 219:713-725; Longo, N. and L.J. Elsas (1998) Adv. Pediatr. 45:293-313).

Monocarboxylate anion transporters are proton-coupled symporters with a broad substrate specificity that includes L-lactate, pyruvate, and the ketone bodies acetate, acetoacetate, and beta-hydroxybutyrate. At least seven isoforms have been identified to date. The isoforms are predicted to have twelve transmembrane (TM) helical domains with a large intracellular loop between TM6 and TM7, and play a critical role in maintaining intracellular pH by removing the protons that are produced stoichiometrically with lactate during glycolysis. The best characterized H\*-monocarboxylate transporter is that of the erythrocyte membrane, which transports L-lactate and a wide range of other aliphatic monocarboxylates. Other cells possess H\*-linked monocarboxylate transporters with differing substrate and inhibitor selectivities. In particular, cardiac muscle and tumor cells have transporters that differ in their K<sub>m</sub> values for certain substrates, including stereoselectivity for L- over D-lactate, and in their sensitivity to inhibitors. There are Na\*-monocarboxylate cotransporters on the luminal surface of intestinal and kidney epithelia, which allow the uptake of lactate, pyruvate, and ketone bodies in these tissues. In addition, there are specific and selective transporters for organic cations and organic anions in organs including the

kidney, intestine and liver. Organic anion transporters are selective for hydrophobic, charged molecules with electron-attracting side groups. Organic cation transporters, such as the ammonium transporter, mediate the secretion of a variety of drugs and endogenous metabolites, and contribute to the maintenance of intercellular pH (Poole, R.C. and A.P. Halestrap (1993) Am. J. Physiol. 264:C761-C782; Price, N.T. et al. (1998) Biochem. J. 329:321-328; and Martinelle, K. and I. Haggstrom (1993) J. Biotechnol. 30:339-350).

ATP-binding cassette (ABC) transporters are members of a superfamily of membrane proteins that transport substances ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. ABC 10 transporters consist of four modules: two nucleotide-binding domains (NBD), which hydrolyze ATP to supply the energy required for transport, and two membrane-spanning domains (MSD), each containing six putative transmembrane segments. These four modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product contains a single NBD and MSD. These "half-15 molecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system. Several genetic diseases are attributed to defects in ABC transporters, such as the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel), adrenoleukodystrophy (adrenoleukodystrophy protein, ALDP), Zellweger syndrome (peroxisomal membrane protein-70, PMP70), and hyperinsulinemic 20 hypoglycemia (sulfonylurea receptor, SUR). Overexpression of the multidrug resistance (MDR) protein, another ABC transporter, in human cancer cells makes the cells resistant to a variety of cytotoxic drugs used in chemotherapy (Taglicht, D. and S. Michaelis (1998) Meth. Enzymol. 292:130-162).

A number of metal ions such as iron, zinc, copper, cobalt, manganese, molybdenum,

selenium, nickel, and chromium are important as cofactors for a number of enzymes. For example,
copper is involved in hemoglobin synthesis, connective tissue metabolism, and bone development,
by acting as a cofactor in oxidoreductases such as superoxide dismutase, ferroxidase
(ceruloplasmin), and lysyl oxidase. Copper and other metal ions must be provided in the diet, and
are absorbed by transporters in the gastrointestinal tract. Plasma proteins transport the metal ions to
the liver and other target organs, where specific transporters move the ions into cells and cellular
organelles as needed. Imbalances in metal ion metabolism have been associated with a number of
disease states (Danks, D.M. (1986) J. Med. Genet. 23:99-106).

P-type ATPases comprise a class of cation-transporting transmembrane proteins. They are integral membrane proteins which use an aspartyl phosphate intermediate to move cations across a membrane. Features of P-type ATPases include: (i) a cation channel; (ii) a stalk, formed by

extensions of the transmembrane  $\alpha$ -helices into the cytoplasm; (iii) an ATP binding domain; (iv) a phosphorylated aspartic acid; (v) an adjacent transduction domain; (vi) a phosphatase domain, which removes the phosphate from the aspartic acid as part of the reaction cycle; and (vii) six or more transmembrane domains. Included in this class are heavy metal-transporting ATPases as well as aminophospholipid transporters.

The transport of phosphatidylserine and phosphatidylethanolamine by aminophospholipid translocase results in the movement of these molecules from one side of a bilayer to another. This transport is conducted by a newly identified subfamily of P-type ATPases which are proposed to be amphipath transporters. Amphipath transporters move molecules having both a hydrophilic and a hydrophobic region. As many as seventeen different genes belong to this P-type ATPases subfamily, being grouped into several distinct classes and subclasses (Halleck, M.S. et al., (1999) Physiol. Genomics 1:139-150; Vulpe, C. Et al., (1993) Nat. Genet. 3:7-13).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process. Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates uptake of long-chain fatty acids (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

The lipocalin superfamily constitutes a phylogenetically conserved group of more than forty proteins that function as extracellular ligand-binding proteins which bind and transport small hydrophobic molecules. Members of this family function as carriers of retinoids, odorants, chromophores, pheromones, allergens, and sterols, and in a variety of processes including nutrient transport, cell growth regulation, immune response, and prostaglandin synthesis. A subset of these proteins may be multifunctional, serving as either a biosynthetic enzyme or as a specific enzyme inhibitor. (Tanaka, T. et al. (1997) J. Biol. Chem. 272:15789-15795; and van't Hof, W. et al. (1997) J. Biol. Chem. 272:1837-1841.)

Members of the lipocalin family display unusually low levels of overall sequence conservation. Pairwise sequence identity often falls below 20%. Sequence similarity between family members is limited to conserved cysteines which form disulfide bonds and three motifs which form a juxtaposed cluster that functions as a target cell recognition site. The lipocalins share an eight stranded, anti-parallel beta-sheet which folds back on itself to form a continuously hydrogen-bonded beta-barrel. The pocket formed by the barrel functions as an internal ligand binding site. Seven loops (L1 to L7) form short beta-hairpins, except loop L1 which is a large

omega loop that forms a lid to partially close the internal ligand-binding site (Flower (1996) Biochem. J. 318:1-14).

Lipocalins are important transport molecules. Each lipocalin associates with a particular ligand and delivers that ligand to appropriate target sites within the organism. Retinol-binding protein (RBP), one of the best characterized lipocalins, transports retinol from stores within the liver to target tissues. Apolipoprotein D (apo D), a component of high density lipoproteins (HDLs) and low density lipoproteins (LDLs), functions in the targeted collection and delivery of cholesterol throughout the body. Lipocalins are also involved in cell regulatory processes. Apo D, which is identical to gross-cystic-disease-fluid protein (GCDFP)-24, is a progesterone/pregnenolone-binding 10 protein expressed at high levels in breast cyst fluid. Secretion of apo D in certain human breast cancer cell lines is accompanied by reduced cell proliferation and progression of cells to a more differentiated phenotype. Similarly, apo D and another lipocalin, α<sub>1</sub>-acid glycoprotein (AGP), are involved in nerve cell regeneration. AGP is also involved in anti-inflammatory and immunosuppressive activities. AGP is one of the positive acute-phase proteins (APP); circulating 15 levels of AGP increase in response to stress and inflammatory stimulation. AGP accumulates at sites of inflammation where it inhibits platelet and neutrophil activation and inhibits phagocytosis. The immunomodulatory properties of AGP are due to glycosylation. AGP is 40% carbohydrate. making it unusually acidic and soluble. The glycosylation pattern of AGP changes during acutephase response, and deglycosylated AGP has no immunosuppressive activity (Flower (1994) FEBS 20 Lett. 354:7-11; Flower (1996) supra).

The lipocalin superfamily also includes several animal allergens, including the mouse major urinary protein (mMUP), the rat α-2-microgloobulin (rA2U), the bovine β-lactoglobulin (βlg), the cockroach allergen (Bla g4), bovine dander allergen (Bos d2), and the major horse allergen, designated Equus caballus allergen 1 (Equ c1). Equ c1 is a powerful allergen responsible for about 80% of anti-horse IgE antibody response in patients who are chronically exposed to horse allergens. It appears that lipocalins may contain a common structure that is able to induce the IgE response (Gregoire, C. et al., (1996) J. Biol. Chem. 271:32951-32959).

Lipocalins are used as diagnostic and prognostic markers in a variety of disease states. The plasma level of AGP is monitored during pregnancy and in diagnosis and prognosis of conditions including cancer chemotherapy, renal disfunction, myocardial infarction, arthritis, and multiple sclerosis. RBP is used clinically as a marker of tubular reabsorption in the kidney, and apo D is a marker in gross cystic breast disease (Flower (1996) supra). Additionally, the use of lipocalin animal allergens may help in the diagnosis of allergic reactions to horses (Gregoire supra), pigs, cockroaches, mice and rats.

35 Mitochondrial carrier proteins are transmembrane-spanning proteins which transport ions

and charged metabolites between the cytosol and the mitochondrial matrix. Examples include the ADP, ATP carrier protein; the 2-oxoglutarate/malate carrier; the phosphate carrier protein; the pyruvate carrier; the dicarboxylate carrier which transports malate, succinate, fumarate, and phosphate; the tricarboxylate carrier which transports citrate and malate; and the Grave's disease 5 carrier protein, a protein recognized by IgG in patients with active Grave's disease, an autoimmune disorder resulting in hyperthyroidism. Proteins in this family consist of three tandem repeats of an approximately 100 amino acid domain, each of which contains two transmembrane regions (Stryer, L. (1995) Biochemistry, W.H. Freeman and Company, New York NY, p. 551; PROSITE PDOC00189 Mitochondrial energy transfer proteins signature; Online Mendelian Inheritance in 10 Man (OMIM) \*275000 Graves Disease).

This class of transporters also includes the mitochondrial uncoupling proteins, which create proton leaks across the inner mitochondrial membrane, thus uncoupling oxidative phosphorylation from ATP synthesis. The result is energy dissipation in the form of heat. Mitochondrial uncoupling proteins have been implicated as modulators of thermoregulation and metabolic rate, and have been 15 proposed as potential targets for drugs against metabolic diseases such as obesity (Ricquier, D. et al. (1999) J. Int. Med. 245:637-642).

#### Ion Channels

The electrical potential of a cell is generated and maintained by controlling the movement of ions across the plasma membrane. The movement of ions requires ion channels, which form ionselective pores within the membrane. There are two basic types of ion channels, ion transporters and gated ion channels. Ion transporters utilize the energy obtained from ATP hydrolysis to actively transport an ion against the ion's concentration gradient. Gated ion channels allow passive flow of an ion down the ion's electrochemical gradient under restricted conditions. Together, these types of ion channels generate, maintain, and utilize an electrochemical gradient that is used in 1) electrical 25 impulse conduction down the axon of a nerve cell, 2) transport of molecules into cells against concentration gradients, 3) initiation of muscle contraction, and 4) endocrine cell secretion.

# Ion Transporters

Ion transporters generate and maintain the resting electrical potential of a cell. Utilizing the energy derived from ATP hydrolysis, they transport ions against the ion's concentration gradient. These transmembrane ATPases are divided into three families. The phosphorylated (P) class ion transporters, including Na\*-K\* ATPase, Ca2+-ATPase, and H+-ATPase, are activated by a phosphorylation event. P-class ion transporters, also known as E1-E2 type ATPases, are responsible for maintaining resting potential distributions such that cytosolic concentrations of Na<sup>+</sup> and Ca<sup>2+</sup> are low and cytosolic concentration of K+ is high. The vacuolar (V) class of ion transporters includes 35 H<sup>+</sup> pumps on intracellular organelles, such as lysosomes and Golgi. V-class ion transporters are

responsible for generating the low pH within the lumen of these organelles that is required for function. The coupling factor (F) class consists of H<sup>+</sup> pumps in the mitochondria. F-class ion transporters utilize a proton gradient to generate ATP from ADP and inorganic phosphate (P<sub>i</sub>).

The P-ATPases are hexamers of a 100 kD subunit with ten transmembrane domains and several large cytoplasmic regions that may play a role in ion binding (Scarborough, G.A. (1999) Curr. Opin. Cell Biol. 11:517-522). The V-ATPases are composed of two functional domains: the V<sub>1</sub> domain, a peripheral complex responsible for ATP hydrolysis; and the V<sub>0</sub> domain, an integral complex responsible for proton translocation across the membrane. The F-ATPases are structurally and evolutionarily related to the V-ATPases. The F-ATPase F<sub>0</sub> domain contains 12 copies of the c subunit, a highly hydrophobic protein composed of two transmembrane domains and containing a single buried carboxyl group in TM2 that is essential for proton transport. The V-ATPase V<sub>0</sub> domain contains three types of homologous c subunits with four or five transmembrane domains and the essential carboxyl group in TM4 or TM3. Both types of complex also contain a single a subunit that may be involved in regulating the pH dependence of activity (Forgac, M. (1999) J. Biol. Chem. 274:12951-12954).

The resting potential of the cell is utilized in many processes involving carrier proteins and gated ion channels. Carrier proteins utilize the resting potential to transport molecules into and out of the cell. Amino acid and glucose transport into many cells is linked to sodium ion co-transport (symport) so that the movement of Na<sup>+</sup> down an electrochemical gradient drives transport of the other molecule up a concentration gradient. Similarly, cardiac muscle links transfer of Ca<sup>2+</sup> out of the cell with transport of Na<sup>+</sup> into the cell (antiport).

## **Gated Ion Channels**

Gated ion channels control ion flow by regulating the opening and closing of pores. The ability to control ion flux through various gating mechanisms allows ion channels to mediate such diverse signaling and homeostatic functions as neuronal and endocrine signaling, muscle contraction, fertilization, and regulation of ion and pH balance. Gated ion channels are categorized according to the manner of regulating the gating function. Mechanically-gated channels open their pores in response to mechanical stress; voltage-gated channels (e.g., Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>2+</sup>, and Cl channels) open their pores in response to changes in membrane potential; and ligand-gated channels (e.g., acetylcholine-, serotonin-, and glutamate-gated cation channels, and GABA- and glycine-gated chloride channels) open their pores in the presence of a specific ion, nucleotide, or neurotransmitter. The gating properties of a particular ion channel (i.e., its threshold for and duration of opening and closing) are sometimes modulated by association with auxiliary channel proteins and/or post translational modifications, such as phosphorylation.

Mechanically-gated or mechanosensitive ion channels act as transducers for the senses of

touch, hearing, and balance, and also play important roles in cell volume regulation, smooth muscle contraction, and cardiac rhythm generation. A stretch-inactivated channel (SIC) was recently cloned from rat kidney. The SIC channel belongs to a group of channels which are activated by pressure or stress on the cell membrane and conduct both Ca<sup>2+</sup> and Na<sup>+</sup> (Suzuki, M. et al. (1999) J. Biol. Chem. 274:6330-6335).

The pore-forming subunits of the voltage-gated cation channels form a superfamily of ion channel proteins. The characteristic domain of these channel proteins comprises six transmembrane domains (S1-S6), a pore-forming region (P) located between S5 and S6, and intracellular amino and carboxy termini. In the Na<sup>+</sup> and Ca<sup>2+</sup> subfamilies, this domain is repeated four times, while in the K<sup>+</sup> channel subfamily, each channel is formed from a tetramer of either identical or dissimilar subunits. The P region contains information specifying the ion selectivity for the channel. In the case of K<sup>+</sup> channels, a GYG tripeptide is involved in this selectivity (Ishii, T.M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11651-11656).

Voltage-gated Na<sup>+</sup> and K<sup>+</sup> channels are necessary for the function of electrically excitable

cells, such as nerve and muscle cells. Action potentials, which lead to neurotransmitter release and muscle contraction, arise from large, transient changes in the permeability of the membrane to Na<sup>+</sup> and K<sup>+</sup> ions. Depolarization of the membrane beyond the threshold level opens voltage-gated Na<sup>+</sup> channels. Sodium ions flow into the cell, further depolarizing the membrane and opening more voltage-gated Na<sup>+</sup> channels, which propagates the depolarization down the length of the cell.

Depolarization also opens voltage-gated potassium channels. Consequently, potassium ions flow outward, which leads to repolarization of the membrane. Voltage-gated channels utilize charged residues in the fourth transmembrane segment (S4) to sense voltage change. The open state lasts only about 1 millisecond, at which time the channel spontaneously converts into an inactive state that cannot be opened irrespective of the membrane potential. Inactivation is mediated by the

channel's N-terminus, which acts as a plug that closes the pore. The transition from an inactive to a closed state requires a return to resting potential.

Voltage-gated Na<sup>+</sup> channels are heterotrimeric complexes composed of a 260 kDa pore-forming  $\alpha$  subunit that associates with two smaller auxiliary subunits,  $\beta 1$  and  $\beta 2$ . The  $\beta 2$  subunit is a integral membrane glycoprotein that contains an extracellular Ig domain, and its association with  $\alpha$  and  $\beta 1$  subunits correlates with increased functional expression of the channel, a change in its gating properties, as well as an increase in whole cell capacitance due to an increase in membrane surface area (Isom, L.L. et al. (1995) Cell 83:433-442).

Non voltage-gated Na<sup>+</sup> channels include the members of the amiloride-sensitive Na<sup>+</sup> channel/degenerin (NaC/DEG) family. Channel subunits of this family are thought to consist of two transmembrane domains flanking a long extracellular loop, with the amino and carboxyl termini

located within the cell. The NaC/DEG family includes the epithelial Na<sup>+</sup> channel (ENaC) involved in Na<sup>+</sup> reabsorption in epithelia including the airway, distal colon, cortical collecting duct of the kidney, and exocrine duct glands. Mutations in ENaC result in pseudohypoaldosteronism type 1 and Liddle's syndrome (pseudohyperaldosteronism). The NaC/DEG family also includes the recently characterized H<sup>+</sup>-gated cation channels or acid-sensing ion channels (ASIC). ASIC subunits are expressed in the brain and form heteromultimeric Na<sup>+</sup>-permeable channels. These channels require acid pH fluctuations for activation. ASIC subunits show homology to the degenerins, a family of mechanically-gated channels originally isolated from C. elegans. Mutations in the degenerins cause neurodegeneration. ASIC subunits may also have a role in neuronal function, or in pain perception, since tissue acidosis causes pain (Waldmann, R. and M. Lazdunski (1998) Curr. Opin. Neurobiol. 8:418-424; Eglen, R.M. et al. (1999) Trends Pharmacol. Sci. 20:337-342).

K<sup>+</sup> channels are located in all cell types, and may be regulated by voltage, ATP concentration, or second messengers such as Ca<sup>2+</sup> and cAMP. In non-excitable tissue, K<sup>+</sup> channels are involved in protein synthesis, control of endocrine secretions, and the maintenance of osmotic equilibrium across membranes. In neurons and other excitable cells, in addition to regulating action potentials and repolarizing membranes, K<sup>+</sup> channels are responsible for setting the resting membrane potential. The cytosol contains non-diffusible anions and, to balance this net negative charge, the cell contains a Na<sup>+</sup>-K<sup>+</sup> pump and ion channels that provide the redistribution of Na<sup>+</sup>, K<sup>+</sup>, and Cl<sup>-</sup>. The pump actively transports Na<sup>+</sup> out of the cell and K<sup>+</sup> into the cell in a 3:2 ratio. Ion channels in the plasma membrane allow K<sup>+</sup> and Cl<sup>-</sup> to flow by passive diffusion. Because of the high negative charge within the cytosol, Cl<sup>-</sup> flows out of the cell. The flow of K<sup>+</sup> is balanced by an electromotive force pulling K<sup>+</sup> into the cell, and a K<sup>+</sup> concentration gradient pushing K<sup>+</sup> out of the cell. Thus, the resting membrane potential is primarily regulated by K<sup>+</sup> flow (Salkoff, L. and T. Jegla (1995) Neuron 15:489-492).

Potassium channel subunits of the <u>Shaker</u>-like superfamily all have the characteristic six transmembrane/1 pore domain structure. Four subunits combine as homo- or heterotetramers to form functional K channels. These pore-forming subunits also associate with various cytoplasmic β subunits that alter channel inactivation kinetics. The <u>Shaker</u>-like channel family includes the voltage-gated K<sup>+</sup> channels as well as the delayed rectifier type channels such as the human ether-ago-go related gene (HERG) associated with long QT, a cardiac dysrythmia syndrome (Curran, M.E. (1998) Curr. Opin. Biotechnol. 9:565-572; Kaczorowski, G.J. and M.L. Garcia (1999) Curr. Opin. Chem. Biol. 3:448-458).

A second superfamily of K<sup>+</sup> channels is composed of the inward rectifying channels (Kir).

Kir channels have the property of preferentially conducting K<sup>+</sup> currents in the inward direction.

These proteins consist of a single potassium selective pore domain and two transmembrane domains, which correspond to the fifth and sixth transmembrane domains of voltage-gated K<sup>+</sup> channels. Kir subunits also associate as tetramers. The Kir family includes ROMK1, mutations in which lead to Bartter syndrome, a renal tubular disorder. Kir channels are also involved in regulation of cardiac pacemaker activity, seizures and epilepsy, and insulin regulation (Doupnik, C.A. et al. (1995) Curr. Opin. Neurobiol. 5:268-277; Curran, supra).

The recently recognized TWIK K\* channel family includes the mammalian TWIK-1, TREK-1 and TASK proteins. Members of this family possess an overall structure with four transmembrane domains and two P domains. These proteins are probably involved in controlling the resting potential in a large set of cell types (Duprat, F. et al. (1997) EMBO J 16:5464-5471).

The voltage-gated Ca <sup>2+</sup> channels have been classified into several subtypes based upon their electrophysiological and pharmacological characteristics. L-type Ca <sup>2+</sup> channels are predominantly expressed in heart and skeletal muscle where they play an essential role in excitation-contraction coupling. T-type channels are important for cardiac pacemaker activity, while N-type and P/Q-type channels are involved in the control of neurotransmitter release in the central and peripheral nervous system. The L-type and N-type voltage-gated Ca <sup>2+</sup> channels have been purified and, though their functions differ dramatically, they have similar subunit compositions. The channels are composed of three subunits. The α<sub>1</sub> subunit forms the membrane pore and voltage sensor, while the α<sub>2</sub>δ and β subunits modulate the voltage-dependence, gating properties, and the current amplitude of the channel. These subunits are encoded by at least six α<sub>1</sub>, one α<sub>2</sub>δ, and four β genes. A fourth subunit, γ, has been identified in skeletal muscle (Walker, D. et al. (1998) J. Biol. Chem. 273:2361-2367; McCleskey, E.W. (1994) Curr. Opin. Neurobiol. 4:304-312).

The high-voltage-activated Ca <sup>2+</sup> channels that have been characterized biochemically include complexes of a pore-forming alpha1 subunit of approximately 190-250 kDa; a

25 transmembrane complex of alpha2 and delta subunits; an intracellular beta subunit; and in some cases a transmembrane gamma subunit. A variety of alpha1 subunits, alpha2delta complexes, beta subunits, and gamma subunits are known. The Cav1 family of alpha1 subunits conduct L-type Ca <sup>2+</sup> currents, which initiate muscle contraction, endocrine secretion, and gene transcription, and are regulated primarily by second messenger-activated protein phosphorylation pathways. The Cav2

30 family of alpha1 subunits conduct N-type, P/Q-type, and R-type Ca <sup>2+</sup> currents, which initiate rapid synaptic transmission and are regulated primarily by direct interaction with G proteins and SNARE proteins and secondarily by protein phosphorylation. The Cav3 family of alpha1 subunits conduct T-type Ca <sup>2+</sup> currents, which are activated and inactivated more rapidly and at more negative membrane potentials than other Ca <sup>2+</sup> current types. The distinct structures and patterns of regulation of these three families of Ca <sup>2+</sup> channels provide an array of Ca <sup>2+</sup> entry pathways in

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response to changes in membrane potential and a range of possibilities for regulation of Ca 2+ entry by second messenger pathways and interacting proteins (Catterall, W.A. (2000) Annu. Rev. Cell Dev. Biol. 16:521-555).

The alpha-2 subunit of the voltage-gated Ca 2+-channel may include one or more Cache 5 domains. An extracellular Cache domain may be fused to an intracellular catalytic domain, such as the histidine kinase, PP2C phosphatase, GGDEF (a predicted diguanylate cyclase), HD-GYP (a predicted phosphodiesterase) or adenylyl cyclase domain, or to a noncatalytic domain, like the methyl-accepting, DNA-binding winged helix-turn-helix, GAF, PAS or HAMP (a domain found in istidine kinases, denylyl cyclases, ethyl-binding proteins and phosphatases). Small molecules are 10 bound via the Cache domain and this signal is converted into diverse outputs depending on the intracellular domains (Anantharaman, V. and Aravind, L.(2000) Trends Biochem. Sci. 25:535-537).

The transient receptor family (Trp) of calcium ion channels are thought to mediate capacitative calcium entry (CCE). CCE is the Ca2+ influx into cells to resupply Ca2+ stores depleted by the action of inositol triphosphate (IP3) and other agents in response to numerous hormones and 15 growth factors. Trp and Trp-like were first cloned from Drosophila and have similarity to voltage gated Ca 2+ channels in the S3 through S6 regions. This suggests that Trp and/or related proteins may form mammalian CCE channels (Zhu, X. et al. (1996) Cell 85:661-671; Boulay, G. et al. (1997) J. Biol. Chem. 272:29672-29680). Melastatin is a gene isolated in both the mouse and human, whose expression in melanoma cells is inversely correlated with melanoma aggressiveness in vivo. 20 The human cDNA transcript corresponds to a 1533-amino acid protein having homology to members of the Trp family. It has been proposed that the combined use of malastatin mRNA expression status and tumor thickness might allow for the determination of subgroups of patients at both low and high risk for developing metastatic disease (Duncan, L.M. et al (2001) J. Clin. Oncol. 19:568-576).

Chloride channels are necessary in endocrine secretion and in regulation of cytosolic and organelle pH. In secretory epithelial cells, Cl enters the cell across a basolateral membrane through an Na<sup>+</sup>, K<sup>+</sup>/Cl<sup>-</sup> cotransporter, accumulating in the cell above its electrochemical equilibrium concentration. Secretion of Cl from the apical surface, in response to hormonal stimulation, leads to flow of Na + and water into the secretory lumen. The cystic fibrosis transmembrane conductance 30 regulator (CFTR) is a chloride channel encoded by the gene for cystic fibrosis, a common fatal genetic disorder in humans. CFTR is a member of the ABC transporter family, and is composed of two domains each consisting of six transmembrane domains followed by a nucleotide-binding site. Loss of CFTR function decreases transepithelial water secretion and, as a result, the layers of mucus that coat the respiratory tree, pancreatic ducts, and intestine are dehydrated and difficult to clear.

35 The resulting blockage of these sites leads to pancreatic insufficiency, "meconium ileus", and

devastating "chronic obstructive pulmonary disease" (Al-Awqati, Q. et al. (1992) J. Exp. Biol. 172:245-266).

The voltage-gated chloride channels (CLC) are characterized by 10-12 transmembrane domains, as well as two small globular domains known as CBS domains. The CLC subunits 5 probably function as homotetramers. CLC proteins are involved in regulation of cell volume, membrane potential stabilization, signal transduction, and transepithelial transport. Mutations in CLC-1, expressed predominantly in skeletal muscle, are responsible for autosomal recessive generalized myotonia and autosomal dominant myotonia congenita, while mutations in the kidney channel CLC-5 lead to kidney stones (Jentsch, T.J. (1996) Curr. Opin. Neurobiol. 6:303-310).

Ligand-gated channels open their pores when an extracellular or intracellular mediator binds to the channel. Neurotransmitter-gated channels are channels that open when a neurotransmitter binds to their extracellular domain. These channels exist in the postsynaptic membrane of nerve or muscle cells. There are two types of neurotransmitter-gated channels. Sodium channels open in response to excitatory neurotransmitters, such as acetylcholine, glutamate, and serotonin. This opening causes an influx of Na+ and produces the initial localized depolarization that activates the voltage-gated channels and starts the action potential. Chloride channels open in response to inhibitory neurotransmitters, such as γ-aminobutyric acid (GABA) and glycine, leading to hyperpolarization of the membrane and the subsequent generation of an action potential. Neurotransmitter-gated ion channels have four transmembrane domains and probably 20 function as pentamers (Jentsch, supra). Amino acids in the second transmembrane domain appear to be important in determining channel permeation and selectivity (Sather, W.A. et al. (1994) Curr. Opin. Neurobiol. 4:313-323).

Ligand-gated channels can be regulated by intracellular second messengers. For example, calcium-activated K+ channels are gated by internal calcium ions. In nerve cells, an influx of calcium during depolarization opens K\* channels to modulate the magnitude of the action potential (Ishi et al., supra). The large conductance (BK) channel has been purified from brain and its subunit composition determined. The  $\alpha$  subunit of the BK channel has seven rather than six transmembrane domains in contrast to voltage-gated K+ channels. The extra transmembrane domain is located at the subunit N-terminus. A 28-amino-acid stretch in the C-terminal region of the subunit (the 30 "calcium bowl" region) contains many negatively charged residues and is thought to be the region responsible for calcium binding. The β subunit consists of two transmembrane domains connected by a glycosylated extracellular loop, with intracellular N- and C-termini (Kaczorowski, supra; Vergara, C. et al. (1998) Curr. Opin. Neurobiol. 8:321-329).

Cyclic nucleotide-gated (CNG) channels are gated by cytosolic cyclic nucleotides. The best 35 examples of these are the cAMP-gated Na + channels involved in olfaction and the cGMP-gated

cation channels involved in vision. Both systems involve ligand-mediated activation of a G-protein coupled receptor which then alters the level of cyclic nucleotide within the cell. CNG channels also represent a major pathway for Ca2+ entry into neurons, and play roles in neuronal development and plasticity. CNG channels are tetramers containing at least two types of subunits, an a subunit which 5 can form functional homomeric channels, and a β subunit, which modulates the channel properties. All CNG subunits have six transmembrane domains and a pore forming region between the fifth and sixth transmembrane domains, similar to voltage-gated K+ channels. A large C-terminal domain contains a cyclic nucleotide binding domain, while the N-terminal domain confers variation among channel subtypes (Zufall, F. et al. (1997) Curr. Opin. Neurobiol. 7:404-412).

The activity of other types of ion channel proteins may also be modulated by a variety of intracellular signalling proteins. Many channels have sites for phosphorylation by one or more protein kinases including protein kinase A, protein kinase C, tyrosine kinase, and casein kinase II, all of which regulate ion channel activity in cells. Kir channels are activated by the binding of the Gβγ subunits of heterotrimeric G-proteins (Reimann, F. and F.M. Ashcroft (1999) Curr. Opin. Cell. 15 Biol. 11:503-508). Other proteins are involved in the localization of ion channels to specific sites in the cell membrane. Such proteins include the PDZ domain proteins known as MAGUKs (membrane-associated guanylate kinases) which regulate the clustering of ion channels at neuronal synapses (Craven, S.E. and D.S. Bredt (1998) Cell 93:495-498).

### **Disease Correlation**

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The etiology of numerous human diseases and disorders can be attributed to defects in the transport of molecules across membranes. Defects in the trafficking of membrane-bound transporters and ion channels are associated with several disorders, e.g., cystic fibrosis, glucose-galactose malabsorption syndrome, hypercholesterolemia, von Gierke disease, and certain forms of diabetes mellitus. Single-gene defect diseases resulting in an inability to transport small 25 molecules across membranes include, e.g., cystinuria, iminoglycinuria, Hartup disease, and Fanconi disease (van't Hoff, W.G. (1996) Exp. Nephrol. 4:253-262; Talente, G.M. et al. (1994) Ann. Intern. Med. 120:218-226; and Chillon, M. et al. (1995) New Engl. J. Med. 332:1475-1480).

Human diseases caused by mutations in ion channel genes include disorders of skeletal muscle, cardiac muscle, and the central nervous system. Mutations in the pore-forming subunits of 30 sodium and chloride channels cause myotonia, a muscle disorder in which relaxation after voluntary contraction is delayed. Sodium channel myotonias have been treated with channel blockers. Mutations in muscle sodium and calcium channels cause forms of periodic paralysis, while mutations in the sarcoplasmic calcium release channel, T-tubule calcium channel, and muscle sodium channel cause malignant hyperthermia. Cardiac arrythmia disorders such as the long QT 35 syndromes and idiopathic ventricular fibrillation are caused by mutations in potassium and sodium

channels (Cooper, E.C. and L.Y. Jan (1998) Proc. Natl. Acad. Sci. USA 96:4759-4766). All four known human idiopathic epilepsy genes code for ion channel proteins (Berkovic, S.F. and I.E. Scheffer (1999) Curr. Opin. Neurology 12:177-182). Other neurological disorders such as ataxias, hemiplegic migraine and hereditary deafness can also result from mutations in ion channel genes (Jen, J. (1999) Curr. Opin. Neurobiol. 9:274-280; Cooper, supra).

Ion channels have been the target for many drug therapies. Neurotransmitter-gated channels have been targeted in therapies for treatment of insomnia, anxiety, depression, and schizophrenia. Voltage-gated channels have been targeted in therapies for arrhythmia, ischemic stroke, head trauma, and neurodegenerative disease (Taylor, C.P. and L.S. Narasimhan (1997) Adv. Pharmacol. 39:47-98). Various classes of ion channels also play an important role in the perception of pain, and thus are potential targets for new analgesics. These include the vanilloid-gated ion channels, which are activated by the vanilloid capsaicin, as well as by noxious heat. Local anesthetics such as lidocaine and mexiletine which blockade voltage-gated Na<sup>+</sup> channels have been useful in the treatment of neuropathic pain (Eglen, supra). Calcium-channel protein expression is altered in metastatic melanomas (Enklaar, T. et al. (2000) Genomics 67:179-187.

Ion channels in the immune system have recently been suggested as targets for immunomodulation. T-cell activation depends upon calcium signaling, and a diverse set of T-cell specific ion channels has been characterized that affect this signaling process. Channel blocking agents can inhibit secretion of lymphokines, cell proliferation, and killing of target cells. A peptide antagonist of the T-cell potassium channel Kv1.3 was found to suppress delayed-type hypersensitivity and allogenic responses in pigs, validating the idea of channel blockers as safe and efficacious immunosuppressants (Cahalan, M.D. and K.G. Chandy (1997) Curr. Opin. Biotechnol. 8:749-756).

The discovery of new transporters and ion channels, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of transport, neurological, muscle, immunological and cell proliferative disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of transporters and ion channels.

#### SUMMARY OF THE INVENTION

The invention features purified polypeptides, transporters and ion channels, referred to collectively as "TRICH" and individually as "TRICH-1," "TRICH-2," "TRICH-3," "TRICH-4," "TRICH-5," "TRICH-6," "TRICH-7," "TRICH-8," "TRICH-9," "TRICH-10," "TRICH-11," "TRICH-12," "TRICH-13," "TRICH-14," "TRICH-15," "TRICH-16," "TRICH-17," "TRICH-18," "TRICH-19," "TRICH-20," "TRICH-21," "TRICH-22," "TRICH-23," "TRICH-24," "TRICH-25,"

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"TRICH-26," "TRICH-27," "TRICH-28," "TRICH-29," "TRICH-30," "TRICH-31," and "TRICH-32." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-32. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:33-64.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide.

30 In another alternative, the invention provides a transgenic organism comprising the recombinant polynucleotide.

The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of

SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous

nucleotides.

The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional TRICH,

comprising administering to a patient in need of such treatment the composition.

Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide 5 comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) exposing a sample 10 comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional TRICH, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide 20 having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the

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absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20° contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) 15 a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide 25 complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

# BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the present invention.

35 Table 2 shows the GenBank identification number and annotation of the nearest GenBank

homolog for polypeptides of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

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#### DESCRIPTION OF THE INVENTION

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a,"
"an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for
example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an
antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in
the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

#### **DEFINITIONS**

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"TRICH" refers to the amino acid sequences of substantially purified TRICH obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of TRICH. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

An "allelic variant" is an alternative form of the gene encoding TRICH. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding TRICH include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as TRICH or a polypeptide with at least one functional characteristic of TRICH. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding TRICH, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding TRICH. The encoded protein may also be "altered," and may contain deletions. insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent TRICH. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of TRICH is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar 30 side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally

occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

5 Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of TRICH. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of TRICH either by directly interacting with TRICH or by acting on components of the biological pathway in which TRICH participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')<sub>2</sub>, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind TRICH polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an in vitro evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No.

5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH<sub>2</sub>), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules,

e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker. (See, e.g., Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13.)

The term "intramer" refers to an aptamer which is expressed in vivo. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothicates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic TRICH, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that annual by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding TRICH or

fragments of TRICH may be employed as hybridization probes. The probes may be stored in freeze-

dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, 10 Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows 15 amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
20	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
25	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Пе	Leu, Val
30	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
35	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical 40 conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the

absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which 5 retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an 15 exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of TRICH or the polynucleotide encoding TRICH which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a 25 molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:33-64 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:33-64, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:33-64 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:33-64 from related polynucleotide sequences. The precise length of a fragment of SEQ 35 ID NO:33-64 and the region of SEQ ID NO:33-64 to which the fragment corresponds are routinely

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determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-32 is encoded by a fragment of SEQ ID NO:33-64. A fragment of SEQ ID NO:1-32 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-32. For example, a fragment of SEQ ID NO:1-32 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-32. The precise length of a fragment of SEQ ID NO:1-32 and the region of SEQ ID NO:1-32 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default

20 parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e
sequence alignment program. This program is part of the LASERGENE software package, a suite of
molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in
Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992)

CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters

are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted"
residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the
"percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local

Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2

Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool

5 Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

10 Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50.

Expect: 10

Word Size: 11

· Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap

penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10
Word Size: 3

Filter: on

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Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas

wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 µg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T<sub>m</sub>) for the specific sequence at a defined ionic strength and pH. The T<sub>m</sub> is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T<sub>m</sub> and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., Cot or Rot analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate 30 substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect

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cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of TRICH which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide

fragment of TRICH which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of TRICH. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of TRICH.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an TRICH may involve lipidation, glycosylation,
30 phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in
the art. These processes may occur synthetically or biochemically. Biochemical modifications will
vary by cell type depending on the enzymatic milieu of TRICH.

"Probe" refers to nucleic acid sequences encoding TRICH, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule.

Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes.

"Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences.

10 Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2<sup>nd</sup> ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple

sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence
that is made by an artificial combination of two or more otherwise separated segments of sequence.
This artificial combination is often accomplished by chemical synthesis or, more commonly, by the
artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques
such as those described in Sambrook, supra. The term recombinant includes nucleic acids that have
been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently,
a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter
sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to
transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing TRICH, nucleic acids encoding TRICH, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or

cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by

infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence 10 having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, or at least 98%, or greater 15 sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the 20 reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide 25 base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9

(May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, or at least 98%, or greater sequence identity over a certain defined length of one of the polypeptides.

#### 35 THE INVENTION

The invention is based on the discovery of new human transporters and ion channels (TRICH), the polynucleotides encoding TRICH, and the use of these compositions for the diagnosis, treatment, or prevention of transport, neurological, muscle, immunological and cell proliferative disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide 10 sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the 15 polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are transporters and ion channels. For example, SEQ ID NO:1 is 83% identical to human sodium-hydrogen exchanger 6 (GenBank ID g2944233) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.1e-242, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:1 also contains a sodium/hydrogen

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exchanger family domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, and other BLAST analyses provide further corroborative evidence that SEQ ID NO:1 is a sodium-hydrogen exchange transporter. In another example, SEQ ID NO:7 is 85% identical to Rattus norvegicus Na<sup>+</sup>/,K<sup>+</sup>-ATPase alpha subunit (GenBank ID g619915) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:7 also contains an E1-E2 ATPase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-10 based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:7 is a cation-transporting ATPase. In yet another example, SEQ ID NO:13 is 77% identical to a human carrier-like protein (GenBank ID g3694661) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.5e-209, which indicates 15 the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:13 also contains a mitochondrial energy transfer protein domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) The presence of this domain is confirmed by BLIMPS, MOTIFS, and PROFILESCAN analyses, providing further corroborative evidence that 20 SEQ ID NO:13 is a transporter. Further, SEQ ID NO:16 is 41% identical to human novel ATPase (GenBank ID g8979801) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.1e-165, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:16 also contains an E1-E2 ATPase domain as determined by searching for statistically significant matches in the hidden 25 Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:16 is a cation-transporting ATPase. In a further example, SEQ ID NO:19 is 43% identical to Sinorhizobium sp. As4 ArsA, the catalytic subunit of the arsenic oxyanion-translocating ATPase (GenBank ID g5802945) as determined by the Basic Local Alignment Search Tool 30 (BLAST). (See Table 2.) The BLAST probability score is 7.7e-125, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:19 also contains an anion-transporting ATPase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from additional BLAST and PROFILESCAN analyses provide

35 further corroborative evidence that SEQ ID NO:19 is an anion-transporting ATPase. In yet a further

example, SEQ ID NO:21 is 54% identical to a murine putative E1-E2 ATPase (GenBank ID g28577) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.2e-190, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:21 also contains six transmembrane 5 domains as determined using TMAP, a program which delineates transmembrane segments. (See Table 3.) Data from BLIMPS, and MOTIFS, analyses provide further corroborative evidence that SEQ ID NO:21 is an ATPase. In a futher example, SEQ ID NO:24 is 52% identical, from residue A77 to residue L1007, to rat NMDAR-L (GenBank ID g2160125) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.5e-262, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEO ID NO:24 also contains a ligand gated ion channel domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS analysis provide further corroborative evidence that SEQ ID NO:24 is a glutamate receptor. SEQ ID NO:2-6, SEQ ID NO:8-15 12, SEQ ID NO:14-15, SEQ ID NO:17-18, SEQ ID NO:20, SEQ ID NO:22-23, and SEQ ID NO:25-32 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-32 are described in Table 7.

As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide sequences of the invention, and of fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:33-64 or that distinguish between SEQ ID NO:33-64 and related polynucleotide sequences.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank cDNAs or ESTs which contributed to the assembly of the full length polynucleotide sequences. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences

including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence 5 identified as FL\_XXXXXX\_N<sub>1</sub>\_N<sub>2</sub>\_YYYYY\_N<sub>3</sub>\_N<sub>4</sub> represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and  $N_{I,2,3...}$ , if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons 10 brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as FLXXXXXX\_gAAAAA\_gBBBBB\_1\_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs
GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).
INCY	Full length transcript and exon prediction from mapping of EST
	sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in

Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant
Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotide

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sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses TRICH variants. A preferred TRICH variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the TRICH amino acid sequence, and which contains at least one functional or structural characteristic of TRICH.

The invention also encompasses polynucleotides which encode TRICH. In a particular 10 embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:33-64, which encodes TRICH. The polynucleotide sequences of SEQ ID NO:33-64, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses a variant of a polynucleotide sequence encoding TRICH. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding TRICH. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID 20 NO:33-64 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:33-64. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of TRICH.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant 25 of a polynucleotide sequence encoding TRICH. A splice variant may have portions which have significant sequence identity to the polynucleotide sequence encoding TRICH, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% 30 polynucleotide sequence identity to the polynucleotide sequence encoding TRICH over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide sequence encoding TRICH. Any one of the splice variants described above can encode an amino acid sequence which contains at least one functional or 35 structural characteristic of TRICH.

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It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding TRICH, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring TRICH, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode TRICH and its variants are generally capable

of hybridizing to the nucleotide sequence of the naturally occurring TRICH under appropriately
selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding
TRICH or its derivatives possessing a substantially different codon usage, e.g., inclusion of nonnaturally occurring codons. Codons may be selected to increase the rate at which expression of the
peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with
which particular codons are utilized by the host. Other reasons for substantially altering the
nucleotide sequence encoding TRICH and its derivatives without altering the encoded amino acid
sequences include the production of RNA transcripts having more desirable properties, such as a
greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode TRICH and TRICH derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding TRICH or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of
hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID
NO:33-64 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and
S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol.
152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in
"Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence

preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

10 The nucleic acid sequences encoding TRICH may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 15 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. 20 (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and 25 PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

to the template at temperatures of about 68°C to 72°C.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotidespecific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the 5 emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode TRICH may be cloned in recombinant DNA molecules that direct expression of TRICH, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express TRICH.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter TRICH-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, 20 oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of TRICH, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with

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fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding TRICH may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, TRICH itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular 10 Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of TRICH, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring 15 polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active TRICH, the nucleotide sequences encoding TRICH or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in 25 polynucleotide sequences encoding TRICH. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding TRICH. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding TRICH and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no 30 additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ.

20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding TRICH and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989)

Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express 10 sequences encoding TRICH. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. 20 Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 25 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding TRICH. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding TRICH can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding TRICH into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M.

Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of TRICH are needed, e.g. for the production of antibodies, vectors which direct high level expression of TRICH may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of TRICH. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 10 1995, supra; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of TRICH. Transcription of sequences encoding TRICH may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO 15 J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding TRICH may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to 25 obtain infective virus which expresses TRICH in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of 30 DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression 35 of TRICH in cell lines is preferred. For example, sequences encoding TRICH can be transformed

into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine

10 phosphoribosyltransferase genes, for use in tk and apr cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dlift confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding TRICH is inserted within a marker gene sequence, transformed cells containing sequences encoding TRICH can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding TRICH under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding TRICH and that express TRICH may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

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Immunological methods for detecting and measuring the expression of TRICH using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on TRICH is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols,

10 Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding TRICH include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled 15 nucleotide. Alternatively, the sequences encoding TRICH, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by 20 Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding TRICH may be cultured under 25 conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression. vectors containing polynucleotides which encode TRICH may be designed to contain signal sequences which direct secretion of TRICH through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. 35 Different host cells which have specific cellular machinery and characteristic mechanisms for

post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding TRICH may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric TRICH protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of TRICH activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available 10 affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the TRICH encoding sequence and the heterologous protein sequence, so that TRICH may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled TRICH may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, 35S-methionine.

TRICH of the present invention or fragments thereof may be used to screen for compounds that specifically bind to TRICH. At least one and up to a plurality of test compounds may be screened for specific binding to TRICH. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of TRICH, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) Current Protocols in Immunology 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which TRICH binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case,

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the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express TRICH, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, Drosophila, or E. coli. Cells expressing TRICH or cell membrane fractions which contain TRICH 5 are then contacted with a test compound and binding, stimulation, or inhibition of activity of either TRICH or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with TRICH, either in 10 solution or affixed to a solid support, and detecting the binding of TRICH to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

TRICH of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of TRICH. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for TRICH activity, wherein TRICH is combined with at least one test compound, and the activity of TRICH in the presence of a test compound is compared with the activity of TRICH in the absence 20 of the test compound. A change in the activity of TRICH in the presence of the test compound is indicative of a compound that modulates the activity of TRICH. Alternatively, a test compound is combined with an in vitro or cell-free system comprising TRICH under conditions suitable for TRICH activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of TRICH may do so indirectly and need not come in direct contact with the 25 test compound. At least one and up to a plurality of test compounds may be screened.

In another embodiment, polynucleotides encoding TRICH or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 30 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo, Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous 35 recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or

developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding TRICH may also be manipulated in vitro in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding TRICH can also be used to create "knockin" humanized animals (pigs) or transgenic animals (nuce or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding TRICH is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress TRICH, e.g., by secreting TRICH in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

## **THERAPEUTICS**

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of TRICH and transporters and ion channels. In addition, examples of tissues expressing TRICH can be found in Table 6. Therefore, TRICH appears to play a role in transport, neurological, muscle, immunological and cell proliferative disorders. In the treatment of disorders associated with increased TRICH expression or activity, it is desirable to decrease the expression or activity of TRICH. In the treatment of disorders associated with decreased TRICH expression or activity, it is desirable to increase the expression or activity of TRICH.

Therefore, in one embodiment, TRICH or a fragment or derivative thereof may be

30 administered to a subject to treat or prevent a disorder associated with decreased expression or
activity of TRICH. Examples of such disorders include, but are not limited to, a transport disorder
such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's
muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes
insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis,

35 normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug

resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long OT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, 5 mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, 10 Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital horn syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup 15 disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, 25 encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular

dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy,

lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, 5 epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplègia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune 10 thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple 15 sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and 20 helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, 25 bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing TRICH or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified TRICH in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of TRICH may be

administered to a subject to treat or prevent a disorder associated with decreased expression or activity of TRICH including, but not limited to, those listed above.

In a further embodiment, an antagonist of TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH. Examples of such disorders include, but are not limited to, those transport, neurological, muscle, immunological and cell proliferative disorders described above. In one aspect, an antibody which specifically binds TRICH may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express TRICH.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding TRICH may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of TRICH including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of TRICH may be produced using methods which are generally known in the art. In particular, purified TRICH may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind TRICH. Antibodies to TRICH may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with TRICH or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to,

Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as

lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to
TRICH have an amino acid sequence consisting of at least about 5 amino acids, and generally will

consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of TRICH amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to TRICH may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBVhybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and 15 Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce TRICH-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-20 10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for TRICH may also be generated. For example, such fragments include, but are not limited to, F(ab')2 fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the F(ab')2 fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. 30 et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between TRICH and its 35 specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies

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reactive to two non-interfering TRICH epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for TRICH. Affinity is expressed as an 5 association constant, K<sub>a</sub>, which is defined as the molar concentration of TRICH-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K, determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple TRICH epitopes, represents the average affinity, or avidity, of the antibodies for TRICH. The K<sub>2</sub> determined for a preparation of monoclonal antibodies, which are monospecific 10 for a particular TRICH epitope, represents a true measure of affinity. High-affinity antibody preparations with K, ranging from about 109 to 1012 L/mole are preferred for use in immunoassays in which the TRICH-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K<sub>n</sub> ranging from about 10<sup>6</sup> to 10<sup>7</sup> L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of TRICH, 15 preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For 20 example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of TRICH-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding TRICH, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding TRICH. Such technology is well known in the art, and antisense 30 oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding TRICH. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered 35 intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence

complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding TRICH may be used for 10 somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase 15 (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 20 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida 25 <u>albicans</u> and <u>Paracoccidioides brasiliensis</u>; and protozoan parasites such as <u>Plasmodium falciparum</u> and Trypanosoma cruzi). In the case where a genetic deficiency in TRICH expression or regulation causes disease, the expression of TRICH from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in

TRICH are treated by constructing mammalian expression vectors encoding TRICH and introducing these vectors by mechanical means into TRICH-deficient cells. Mechanical transfer technologies for use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev.

Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr.

Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of TRICH include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). TRICH may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding TRICH from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to TRICH expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding TRICH under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant")

discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4<sup>+</sup> T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding TRICH to cells which have one or more genetic abnormalities with respect to the expression of TRICH. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding TRICH to target cells which have one or more genetic abnormalities with 20 respect to the expression of TRICH. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing TRICH to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. 25 Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also 30 taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and 35 propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to

those of ordinary skill in the art.

In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding TRICH to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based 5 on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for TRICH into 10 the alphavirus genome in place of the capsid-coding region results in the production of a large number of TRICH-coding RNAs and the synthesis of high levels of TRICH in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the 15 gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of TRICH into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding TRICH.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by

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scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding TRICH. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6.

Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2'O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding TRICH.

25 Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased TRICH expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding TRICH may be therapeutically useful, and in the treatment of disorders associated with decreased TRICH expression or activity, a compound which specifically promotes expression of the polynucleotide encoding TRICH may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in

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altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding TRICH is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding 10 TRICH are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding TRICH. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a 15 change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa 20 cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable

excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of TRICH, antibodies to TRICH, and mimetics, agonists, antagonists, or inhibitors of 5 TRICH.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar 15 region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active 20 ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising TRICH or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of 25 the macromolecule. Alternatively, TRICH or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example TRICH or fragments thereof, antibodies of TRICH, and agonists, antagonists or inhibitors of

TRICH, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED<sub>50</sub> (the dose therapeutically effective in 50% of the population) or LD<sub>50</sub> (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is 5 the therapeutic index, which can be expressed as the LD<sub>50</sub>/ED<sub>50</sub> ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED50 with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, 10 the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the 15 subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1  $\mu$ g to 100,000  $\mu$ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and 20 methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

## DIAGNOSTICS

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In another embodiment, antibodies which specifically bind TRICH may be used for the diagnosis of disorders characterized by expression of TRICH, or in assays to monitor patients being treated with TRICH or agonists, antagonists, or inhibitors of TRICH. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for TRICH include methods which utilize the antibody and a label to detect 30 TRICH in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring TRICH, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of TRICH expression.

Normal or standard values for TRICH expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to TRICH under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of TRICH expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding TRICH may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of TRICH may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of TRICH, and to monitor regulation of TRICH levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding TRICH or closely related molecules may be used to identify nucleic acid sequences which encode TRICH. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding TRICH, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the TRICH encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:33-64 or from genomic sequences including promoters, enhancers, and introns of the TRICH gene.

Means for producing specific hybridization probes for DNAs encoding TRICH include the cloning of polynucleotide sequences encoding TRICH or TRICH derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes <u>in vitro</u> by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as <sup>32</sup>P or <sup>35</sup>S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding TRICH may be used for the diagnosis of disorders associated with expression of TRICH. Examples of such disorders include, but are not limited to, a transport disorder such as akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic

fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, prostate cancer, cardiac disorders associated with transport, e.g., angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, polymyositis, neurological disorders associated with transport, e.g., 10 Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia, and other disorders associated with transport, e.g., neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, 15 glucose-galactose malabsorption syndrome, glycogen storage disease, hypercholesterolemia, adrenoleukodystrophy, Zellweger syndrome, Menkes disease, occipital hom syndrome, von Gierke disease, pseudohypoaldosteronism type 1, Liddle's syndrome, cystinuria, iminoglycinuria, Hartup disease, Fanconi disease, and Bartter syndrome; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders. amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system 25 disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, hemiplegic migraine, tardive

35 dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive

supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; a muscle disorder such as cardiomyopathy, myocarditis, Duchenne's muscular dystrophy, Becker's muscular dystrophy, myotonic dystrophy, central core disease, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, infectious myositis, polymyositis, dermatomyositis, 5 inclusion body myositis, thyrotoxic myopathy, ethanol myopathy, angina, anaphylactic shock, arrhythmias, asthma, cardiovascular shock, Cushing's syndrome, hypertension, hypoglycemia, myocardial infarction, migraine, pheochromocytoma, and myopathies including encephalopathy, epilepsy, Kearns-Sayre syndrome, lactic acidosis, myoclonic disorder, ophthalmoplegia, acid maltase deficiency (AMD, also known as Pompe's disease), generalized myotonia, and myotonia 10 congenita; an immunological disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, 15 diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis. polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, 20 Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease 25 (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, 30 thymus, thyroid, and uterus. The polynucleotide sequences encoding TRICH may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered TRICH expression. Such qualitative or quantitative methods are well known in the art.

In a particular aspect, the nucleotide sequences encoding TRICH may be useful in assays

that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding TRICH may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding TRICH in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of TRICH, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding TRICH, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding TRICH may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding TRICH, or a fragment of a polynucleotide complementary to the polynucleotide encoding TRICH, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent

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conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic 5 disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding TRICH are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause 10 differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in highthroughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by 15 comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of TRICH include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The 30 microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this

information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, TRICH, fragments of TRICH, or antibodies specific for TRICH may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression <u>in vivo</u>, as in the case of a tissue or biopsy sample, or <u>in vitro</u>, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is

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useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National 5 Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the 10 treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared . with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance 20 under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, 25 supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are 30 compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence 35 data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for TRICH to quantify the levels of TRICH expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. 5 Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, 15 due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount 30 of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad.

Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in DNA Microarrays: A Practical Approach, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

In another embodiment of the invention, nucleic acid sequences encoding TRICH may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during 10 chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. 15 (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding TRICH on a physical map and a specific disorder, or a predisposition to a specific disorder, may 25 help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for 35 further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide

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sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, TRICH, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between TRICH and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT

10 application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with TRICH, or fragments thereof, and washed. Bound TRICH is then detected by methods well known in the art. Purified TRICH can also be coated directly onto plates for use in the aforementioned drug screening techniques.

Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding TRICH specifically compete with a test compound for binding TRICH. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with TRICH.

In additional embodiments, the nucleotide sequences which encode TRICH may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below, including U.S. Ser. No. 60/254,303, U.S. Ser. No. 60/256,190, U.S. Ser. No. 60/257,504, U.S. Ser. No. 60/261,546, U.S. Ser. No. 60/262,832, U.S. Ser. No. 60/264,377, and U.S. Ser. No. 60/266,019, are expressly incorporated by reference herein.

### **EXAMPLES**

### I. Construction of cDNA Libraries

35 Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD

database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the 15 UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected 20 (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid 25 (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Life Technologies.

### II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids

were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

### III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows.

Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing

vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and
25 programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis.

The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae,
30 Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs,

stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:33-64. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

### IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative transporters and ion channels were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range

of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode transporters and ion channels, the encoded polypeptides were analyzed by querying against PFAM models for transporters and ion channels. Potential transporters and ion channels were also identified by homology to Incyte cDNA sequences that had been annotated as transporters and ion channels. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

### V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example 20 III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm 30 in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases.

genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

### "Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST

analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs

(HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

### VI. Chromosomal Mapping of TRICH Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:33-64 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:33-64 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously

identified disease genes map within or in proximity to the intervals indicated above.

### VII. Analysis of Polynucleotide Expression

Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which

5 RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, supra, ch. 7;

Ausubel (1995) supra, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

15

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding TRICH are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous

system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding TRICH. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

### 10 VIII. Extension of TRICH Encoding Polynucleotides

Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment.

One primer was synthesized to initiate 5'extension of the known fragment, and the other primer was synthesized to initiate 3'extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg<sup>2+</sup>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech),

ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step

The concentration of DNA in each well was determined by dispensing 100 µl PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II

35 (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the

30 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

concentration of DNA. A 5  $\mu$ l to 10  $\mu$ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates,

digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and
sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For
shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%)
agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended
clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector

(Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction
site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on
antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C im
384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase

(Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

### IX. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:33-64 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μCi of [γ-32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech).

35 An aliquot containing 107 counts per minute of the labeled probe is used in a typical membrane-

based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 5 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

### X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena (1999), supra). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon 15 wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; 20 Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides 25 in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element 30 on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

### Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)\* RNA is purified using the oligo-(dT) cellulose method. Each poly(A)\* RNA sample is 35 reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X

first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)<sup>+</sup> RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)<sup>+</sup> RNAs are synthesized by in vitro

5 transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

### **Microarray Preparation**

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5  $\mu$ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1  $\mu$ l of the array element DNA, at an average concentration of 100 ng/ $\mu$ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

### 5 Hybridization

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Hybridization reactions contain 9 μl of sample mixture consisting of 0.2 μg each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

### 10 Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-todigital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-

compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

### XI. Complementary Polynucleotides

Sequences complementary to the TRICH-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring TRICH. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of TRICH. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the TRICH-encoding transcript.

### XII. Expression of TRICH

Expression and purification of TRICH is achieved using bacterial or virus-based expression systems. For expression of TRICH in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac* (*tac*) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express TRICH upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of TRICH in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding TRICH by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription.

Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or

human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, TRICH is synthesized as a fusion protein with, e.g., glutathione

S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from <a href="Schistosoma japonicum">Schistosoma japonicum</a>, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from

TRICH at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, <a href="supra">supra</a>, ch. 10 and 16). Purified TRICH obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII where applicable.

### XIII. Functional Assays

TRICH function is assessed by expressing the sequences encoding TRICH at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA 20 expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10  $\mu$ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2  $\mu g$  of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein 25 provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. 30 FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular 35 proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane

composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of TRICH on gene expression can be assessed using highly purified

5 populations of cells transfected with sequences encoding TRICH and either CD64 or CD64-GFP.

CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding TRICH and other genes of interest can be analyzed by northern analysis or microarray techniques.

### XIV. Production of TRICH Specific Antibodies

TRICH substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the TRICH amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, <a href="mailto:supra">supra</a>.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-TRICH activity by, for example, binding the peptide or TRICH to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

### XV. Purification of Naturally Occurring TRICH Using Specific Antibodies

Naturally occurring or recombinant TRICH is substantially purified by immunoaffinity chromatography using antibodies specific for TRICH. An immunoaffinity column is constructed by covalently coupling anti-TRICH antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

35 Media containing TRICH are passed over the immunoaffinity column, and the column is

washed under conditions that allow the preferential absorbance of TRICH (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/TRICH binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and TRICH is collected.

### 5 XVI. Identification of Molecules Which Interact with TRICH

Molecules which interact with TRICH may include transporter substrates, agonists or antagonists, modulatory proteins such as Gβγ proteins (Reimann, supra) or proteins involved in TRICH localization or clustering such as MAGUKs (Craven, supra). TRICH, or biologically active fragments thereof, are labeled with <sup>125</sup>I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M.

Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH.

multi-well plate are incubated with the labeled TRICH, washed, and any wells with labeled TRICH complex are assayed. Data obtained using different concentrations of TRICH are used to calculate values for the number, affinity, and association of TRICH with the candidate molecules.

Alternatively, proteins that interact with TRICH are isolated using the yeast 2-hybrid system (Fields, S. and O. Song (1989) Nature 340:245-246). TRICH, or fragments thereof, are expressed as fusion proteins with the DNA binding domain of Gal4 or lexA, and potential interacting proteins are expressed as fusion proteins with an activation domain. Interactions between the TRICH fusion protein and the TRICH interacting proteins (fusion proteins with an activation domain) reconstitute a transactivation function that is observed by expression of a reporter gene. Yeast 2-hybrid systems are commercially available, and methods for use of the yeast 2-hybrid system with ion channel proteins are discussed in Niethammer, M. and M. Sheng (1998, Meth. Enzymol. 293:104-122).

TRICH may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

Potential TRICH agonists or antagonists may be tested for activation or inhibition of TRICH ion channel activity using the assays described in section XVIII.

### XVII. Demonstration of TRICH Activity

Ion channel activity of TRICH is demonstrated using an electrophysiological assay for ion conductance. TRICH can be expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector encoding TRICH. Eukaryotic expression vectors are commercially available, and the techniques to introduce them into cells are well known to those skilled in the art. A second plasmid which expresses any one of a number of marker genes, such as ß-galactosidase, is co-transformed into the cells to allow rapid identification of those cells which have taken up and expressed the foreign DNA. The cells are incubated for 48-72 hours after

transformation under conditions appropriate for the cell line to allow expression and accumulation of TRICH and  $\beta$ -galactosidase.

Transformed cells expressing \(\textit{\beta}\)-galactosidase are stained blue when a suitable colorimetric substrate is added to the culture media under conditions that are well known in the art. Stained cells are tested for differences in membrane conductance by electrophysiological techniques that are well known in the art. Untransformed cells, and/or cells transformed with either vector sequences alone or \(\theta\)-galactosidase sequences alone, are used as controls and tested in parallel. Cells expressing TRICH will have higher anion or cation conductance relative to control cells. The contribution of TRICH to conductance can be confirmed by incubating the cells using antibodies specific for TRICH. The antibodies will bind to the extracellular side of TRICH, thereby blocking the pore in the ion channel, and the associated conductance.

Alternatively, ion channel activity of TRICH is measured as current flow across a TRICH-containing Xenopus laevis oocyte membrane using the two-electrode voltage-clamp technique (Ishi et al., supra; Jegla, T. and L. Salkoff (1997) J. Neurosci. 17:32-44). TRICH is subcloned into an appropriate Xenopus oocyte expression vector, such as pBF, and 0.5-5 ng of mRNA is injected into mature stage IV oocytes. Injected oocytes are incubated at 18 °C for 1-5 days. Inside-out macropatches are excised into an intracellular solution containing 116 mM K-gluconate, 4 mM KCl, and 10 mM Hepes (pH 7.2). The intracellular solution is supplemented with varying concentrations of the TRICH mediator, such as cAMP, cGMP, or Ca<sup>+2</sup> (in the form of CaCl<sub>2</sub>), where appropriate.

Electrode resistance is set at 2-5 MΩ and electrodes are filled with the intracellular solution lacking mediator. Experiments are performed at room temperature from a holding potential of 0 mV. Voltage ramps (2.5 s) from -100 to 100 mV are acquired at a sampling frequency of 500 Hz. Current measured is proportional to the activity of TRICH in the assay. In particular, the activity of TRICH-1 is measured as Na<sup>+</sup> conductance and the activity of TRICH-3 is measured as Ca<sup>2+</sup> conductance.

Transport activity of TRICH is assayed by measuring uptake of labeled substrates into Xenopus laevis oocytes. Oocytes at stages V and VI are injected with TRICH mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl 2, 1mM MgCl 2, 1mM Na<sub>2</sub>HPO<sub>4</sub>, 5 mM Hepes, 3.8 mM NaOH, 50µg/ml gentamycin, pH 7.8) to allow expression of TRICH. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl 2, 1mM MgCl 2, 10 mM Hepes/Tris pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, ions, and neurotransmitters) is initiated by adding labeled substrate (e.g. radiolabeled with <sup>3</sup>H, fluorescently labeled with rhodamine, etc.) to the oocytes. After incubating for 30 minutes, uptake is terminated by washing the oocytes three times in Na\*-free medium, measuring the incorporated label, and comparing with controls. TRICH

activity is proportional to the level of internalized labeled substrate. Test substrates include ran-GTP for TRICH-2, glucose for TRICH-5, amino acids for TRICH-6 and TRICH-14, cations for TRICH-7 and TRICH-16, Na+, K+ and Cl-ions for TRICH-15, reduced folate or analogues such as methotrexate for TRICH-17, divalent cations for TRICH-18, anions such as arsenate and antimonite 5 for TRICH-19, and nitrate or oligopeptides for TRICH-20.

ATPase activity associated with TRICH can be measured by hydrolysis of radiolabeled ATP-[γ-<sup>32</sup>P], separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered <sup>32</sup>P using a scintillation counter. The reaction mixture contains ATP-[y-<sup>32</sup>P] and varying amounts of TRICH in a suitable buffer incubated at 37°C for a suitable period of time. The 10 reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base. and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of <sup>32</sup>P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of TRICH in the assay.

Lipocalin activity of TRICH is measured by ligand fluorescence enhancement spectrofluorometry (Lin et al. (1997) Molecular Vision 3:17). Examples of ligands include retinol (Sigma, St. Louis MO) and 16-anthryloxy-palmitic acid (16-AP) (Molecular Probes Inc., Eugene OR). Ligand is dissolved in 100% ethanol and its concentration is estimated using known extinction coefficents (retinol: 46,000 A/M/cm at 325 nm; 16-AP: 8,200 A/M/cm at 361 nm). A 20 700 μl aliquot of 1 μM TRICH in 10 mM Tris (pH 7.5), 2 mM EDTA, and 500 mM NaCl is placed in a 1 cm path length quartz cuvette and 1  $\mu$ l aliquots of ligand solution are added. Fluorescence is measured 100 seconds after each addition until readings are stable. Change in fluorescence per unit change in ligand concentration is proportional to TRICH activity.

### XVIII. Identification of TRICH Agonists and Antagonists

TRICH is expressed in a eukaryotic cell line such as CHO (Chinese Hamster Ovary) or HEK (Human Embryonic Kidney) 293. Ion channel activity of the transformed cells is measured in the presence and absence of candidate agonists or antagonists. Ion channel activity is assayed using patch clamp methods well known in the art or as described in Example XVII. Alternatively, ion channel activity is assayed using fluorescent techniques that measure ion flux across the cell membrane (Velicelebi, G. et al. (1999) Meth. Enzymol. 294:20-47; West, M.R. and C.R. Molloy (1996) Anal. Biochem. 241:51-58). These assays may be adapted for high-throughput screening using microplates. Changes in internal ion concentration are measured using fluorescent dyes such as the Ca2+ indicator Fluo-4 AM, sodium-sensitive dyes such as SBFI and sodium green, or the Clindicator MQAE (all available from Molecular Probes) in combination with the FLIPR fluorimetric 35 plate reading system (Molecular Devices). In a more generic version of this assay, changes in

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membrane potential caused by ionic flux across the plasma membrane are measured using oxonyl dyes such as DiBAC<sub>4</sub> (Molecular Probes). DiBAC<sub>4</sub> equilibrates between the extracellular solution and cellular sites according to the cellular membrane potential. The dye's fluorescence intensity is 20-fold greater when bound to hydrophobic intracellular sites, allowing detection of DiBAC<sub>4</sub> entry into the cell (Gonzalez, J.E. and P.A. Negulescu (1998) Curr. Opin. Biotechnol. 9:624-631). Candidate agonists or antagonists may be selected from known ion channel agonists or antagonists, peptide libraries, or combinatorial chemical libraries.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

### Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide
				Д
7484831	1	7484831CD1	33	7484831CB1
2477266	2	2477266CDI	34	2477266CB1
3552033	3	3552033CD1	35	3552033CB1
4778139	4	4778139CD1	36	4778139CB1
4787433	5	4787433CD1	37	4787433CB1
7483598	9	7483598CD1	38	7483598CB1
7484823	7	7484823CD1	39	7484823CB1
143935	8	143935CD1	40	143935CB1
5923789	6	5923789CD1	41	5923789CB1
6046484	10	6046484CD1	42	6046484CB1
7481427	11	7481427CD1	43	7481427CB1
7483595	12	7483595CD1	44	7483595CB1
3788427	13	3788427CD1	45	3788427CB1
6972455	14	6972455CD1	46	6972455CB1
8077668	15	8077668CD1	47	8077668CB1
55120485	. 16	55120485CD1	48	55120485CB1
3112883	17	3112883CD1	49	3112883CB1
4253888	18	4253888CD1	50	4253888CB1
7479974	19	7479974CD1	51	7479974CB1
7483850	20	7483850CD1	52	7483850CB1
5508353	21	5508353CD1	53	5508353CB1
8543628	22	8543628CD1	54	8543628CB1
7482754	23	7482754CD1	55	7482754CB1
3794818	24	3794818CD1	95	3794818CB1
4717525	25	4717525CD1		4717525CB1
5091793		5091793CD1		5091793CB1
5945527	27	5945527CD1	59	5945527CB1
6941124	28	6941124CD1	09	6941124CB1
6972530	29	6972530CD1	61	6972530CB1

Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte
	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide
				A
6991750	30	6991750CD1	79	6991750CB1
71726948	31	71726948CD1	63	71726948CB1
7487393	32	7487393CD1	94	7487393CB1

1 7484831CD1 2 2477266CD1 3 3552033CD1 5 4778139CD1 6 7483598CD1 7 7484823CD1		g2944233 g2102696 g3243075 g8131903 g2337865	5.1e-242 1.0e-37 0.0 5.1e-107 2.3e-251	
		92102696 93243075 98131903 92337865	1.0e-37 0.0 5.1e-107 2.3e-251	K  ~   K
		93243075 98131903 92337865 36978016	0.0 5.1e-107 2.3e-251	, S [ ~ []
		93243075 98131903 92337865 96978016	0.0 5.1e-107 2.3e-251	
		32337865 36978016	5.1e-107 2.3e-251	~   H
		48131903 42337865 46978016	5.1e-107 2.3e-251	[Mus musculus] transient receptor potential-related protein
		32337865 36978016	2.3e-251	
		36978016		(Homo sapiens) organic cation transporter
7484823CDI			1.9e-32	[Rattus norvegicus] neuronal glutamine transporter
7484823CD1				Varoqui,H. et al. (2000) J. Biol. Chem. 275:4049-4054
		g619915	0.0	[Rattus norvegicus] Na,K-ATPase alpha subunit
				Shamraj, O.I., and Lingrel, J.B. (1994)Proc. Natl. Acad. Sci. USA 91:12952-12956
143935CD1		g179304	7.8e-116	B12 protein [Homo sapiens] (Wolf, F.W. et al. (1992) J. Biol. Chem. 267:1317-1326)
5923789CD1		g1552526	0.0	sodium-calcium exchanger form 3 [Rattus norvegicus] (Nicoll, D.A. et al. (1996) J. Biol. Chem. 271:24914-24921)
6046484CD1	•	g3243075	0.0	melastatin 1 [Homo sapiens] (Hunter, J.J. (1998) Genomics 54:116-123)
7481427CD1		9178661	9.8e-93	adenine nucleotide translocator-2 [Homo sapiens] (Ku, D.H. et al. (1990) J. Biol.

Table 2 (cont.)

10 NO:   Polypeptide ID   ID NO:   7483595CD1   g9453726   3.7e-61   1		Incyte	GenBank	Probability	GenBank Homolog
7483595CD1   G9453726   3.7e-61     3788427CD1   G4155688   1.7e-24     6972455CD1   G4155688   1.7e-24     8077668CD1   G8979801   1.1e-165     3112883CD1   G3979801   1.1e-165     4253888CD1   G3925431   1.6e-29     7479974CD1   G5802945   7.7e-125     7483850CD1   G11933414   6.3e-11     5508353CD1   G457270   5.2e-190     68543628CD1   G6967939   3.7e-45	d	Polypeptide ID	ID NO:	score	
3788427CD1	12	7483595CD1	g9453726	3.7e-61	bA48209.2 (Novel sulphate transporter family member) [Homo sapiens]
6972455CD1       g4155688       1.7e-24         8077668CD1       g5081312       6.6e-47         55120485CD1       g8979801       1.1e-165         3112883CD1       g3115983       4.0e-128         4253888CD1       g3925431       1.6e-29         7479974CD1       g5802945       7.7e-125         7483850CD1       g11933414       6.3e-11         5508353CD1       g6457270       5.2e-190         8543628CD1       g6967939       3.7e-45	13	3788427CD1	g3694661	5.5e-209	carrier protein-like; similar to Q01888 (PID:g266574) [Homo sapiens]
8077668CD1       g5081312       6.6e-47         55120485CD1       g8979801       1.1e-165         3112883CD1       g3115983       4.0e-128         4253888CD1       g3925431       1.6e-29         7479974CD1       g5802945       7.7e-125         7483850CD1       g11933414       6.3e-11         5508353CD1       g6457270       5.2e-190         8543628CD1       g6967939       3.7e-45	14	6972455CD1	g4155688	1.7e-24	(Helicobacter pylori J99) AMINO ACID ABC TRANSPORTER, BINDING PROTEIN PRECURSOR
55120485CD1 g8979801 1.1e-165 3112883CD1 g3115983 4.0e-128 4253888CD1 g3925431 1.6e-29 7479974CD1 g5802945 7.7e-125 7483850CD1 g11933414 6.3e-11 5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45	15	8077668CD1	g5081312	6.6e-47	(Rattus norvegicus) bumetanide- sensitive Na-K-2Cl cotransporter Anzai, N., et al. (1999) Roles of
55120485CD1       g8979801       1.1e-165         3112883CD1       g3115983       4.0e-128         4253888CD1       g3925431       1.6e-29         7479974CD1       g5802945       7.7e-125         7483850CD1       g11933414       6.3e-11         5508353CD1       g6457270       5.2e-190         8543628CD1       g6967939       3.7e-45					vasopressin and hypertonicity in basolateral Na/K/2Cl cotransporter
55120485CD1       g8979801       1.1e-165         3112883CD1       g3115983       4.0e-128         4253888CD1       g3925431       1.6e-29         7479974CD1       g5802945       7.7e-125         7483850CD1       g11933414       6.3e-11         5508353CD1       g6457270       5.2e-190         8543628CD1       g6967939       3.7e-45					expression in rat kidney inner
55120485CD1       g8979801       1.1e-165         3112883CD1       g3115983       4.0e-128         4253888CD1       g3925431       1.6e-29         7479974CD1       g5802945       7.7e-125         7483850CD1       g11933414       6.3e-11         5508353CD1       g6457270       5.2e-190         8543628CD1       g6967939       3.7e-45					duct cells. Jpn. J. Physiol. 49, 201-206
3112883CD1 g3115983 4.0e-128 425388CD1 g3925431 1.6e-29 7479974CD1 g5802945 7.7e-125 7483850CD1 g11933414 6.3e-11 5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45	16	55120485CD1	g8979801	1.1e-165	
4253888CD1 g3925431 1.6e-29 7479974CD1 g5802945 7.7e-125 7483850CD1 g11933414 6.3e-11 5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45	17	3112883CD1	g3115983	4.0e-128	dJ206D15.1 (Reduced Folate Carier protein RFC LIKE) [Homo sapiens]
7479974CD1 g5802945 7.7e-125 7483850CD1 g11933414 6.3e-11 5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45	18	4253888CD1	g3925431	1.6e-29	[Caenorhabditis elegans] (282084) contains similarity to Pfam domain:
7479974CD1 g5802945 7.7e-125 7483850CD1 g11933414 6.3e-11 .5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45					PF01769 (Divalent cation transporter), Score=211.5, E- value=4.2e-60. N=2
7483850CD1 g11933414 6.3e-11 .5508353CD1 g6457270 5.2e-190 .8543628CD1 g6967939 3.7e-45	19	7479974CD1	g5802945	7.7e-125	[Sinorhizobium sp. As4] ArsA
7483850CD1 g11933414 6.3e-11 5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45					oxyanion-translocating ATPase)
.5508353CD1 g6457270 5.2e-190 8543628CD1 g6967939 3.7e-45	20	7483850CD1	g11933414	6.3e-11	[Glycine max] nitrate transporter NRT1-5
8543628CD1 g6967939 3.7e-45	21	.5508353CD1	g6457270	5.2e-190	[Mus musculus] putative E1-E2 ATPase Halleck, M.S. et al., (1999) Physiol. Genomics (Online) 1:139-150
ABC transporter integral memb protein Takamori S. et al., (2000)	22	8543628CD1	g6967939	3.7e-45	MEDLINE : 20473714 [Campylobacter jejuni] amino-acid
					ABC transporter integral membrane
					i S. et al., 407:189-94

Table 2 (cont.)

GenBank Homolog	[Homo sapiens] amino acid transporter system Al Wang H. et al., (2000) Biochem. Biophys. Res. Commun. 273:1175-9	NWDAR-L [Rattus norvegicus] Sucher, N.J. et al. (1995) J. Neurosci. 15:6509-6520	calcium-binding transporter [Homo	Similarity to multidrug resistance protein (SW:BMR1_BACSU)	[Caenorhabditis elegans] The C. elegans Sequencing Consortium (1998) Science 282: 2012-2018	ol 3-phos	anion exchanger 3 brain isoform [Homo sapiens] Yannoukakos, D. et al. (1994) Circ.	1010 ≿ 0	cardiac sodium-calcium exchanger [Oncorhynchus mykiss] Xue,X.H. et al. (1999) Am. J. Physiol 277.6693-6700	10 .	anion transpor et al. (2000) 5:4507-4512
Probability score	1.9e-20	1.5e-262	7.8e-111	1.1e-51		1.5e-161	6.8e-66	3.5e-16.	5.5e-11	1.0e-152	2.2e-118
GenBank ID NO:	g11640743	g2160125	g6841066	93880532		g7543982	g476222	g10175963	g6273849	91628579	97707622
Incyte Polypeptide ID	7482754CD1	3794818CD1	4717525CD1	5091793CD1		5945527CD1	6941124CD1	6972530CD1	6991750CD1	71726948CD1	7487393CD1
Polypeptide SEQ ID NO:	23	24	25	26		27	28	29	30	31	32

Table 3

Analytical Methods and Databases	HMMER		ТМАР	HMMER-PFAM	BLIMPS-PRINTS	BLIMPS-PRINTS	BLIMPS-PRODOM	BLAST-PRODOM	BLAST-PRODOM	BLAST-DOMO
	Signal peptide: M1-A37		Transmembrane domains: P18-A39, R68-I88, R95-I115, K175-H203, L208-L236, D245-A269, A282-Q306, A319-L347, L360-L388, Y428-G456, H482-T502, Q508-L536 N-terminus is non-cytosolic	yen exchanger family:	Na+/H+ exchanger isoform signatures PR01088: S44-A63, E64-I88, W89-I107, Y108-Q134, S299-D316, A318-M37, G588-D606, P612-O640, V641-D668	res PR01084: I211-T219,	transport exchanger PD01672: 182-M230, A319-V344, F381-F414, 419-S465, F466-T512	gen exchanger 6, myeloblast G527-E547	nembrane transport antiporter, 100631: L74-G125	Beta Na exchanger: DM02572 P48764 10-734: D171-R539,R21-L116 DM02572 Q01345 12-703: D179-D563,L22-R117 DM02572 P50482 16-723: E181-F558,G16-S124 DM02572 P26434 14-716: D179-D563, A48-T120,
Potential Si Glycosyl- Do ation Sites			H	So	, PR	Na	Na+ V. F.	OS OS	Na ex	
Potential Phosphoryl- ation Sites	S11 S52 S66 S124 S147 S198 S244 S260 S546	S585 S689 S694 S695 S712 T59 T133 T154 T177 T581 T658 T665						·		
Amino Acid Residues	726									
Incyte Polypep- tide ID	7484831CD1	······································								
2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	1									

Ğ	Incyte	Amino	Potential	Potential	Signature Compense	
Ω 10 10	Polypep- tide ID	Acid Residues	Pho	Glycosyl- ation Sites		Analytical Methods and
2	2477266cn1	1081	070 770	١		Databases
	1000001120		S180 S192 S315	989N C9TN	Transmembrane domains: P197-L213 R255-V275 E498-P520	TMAP
			S493 S615 S639 S697 S1011 T18		I839-V863 N terminus cytosolic	
			,61			
			T328			
			T386 T/86 T813 T871 T881 T907			
					IMPORTIN SUBINIT KARYODHERIN DROMETN	MOROGE HOW IN
					r repeat	PLAST_FRODOM
					F691-D1033	
					b: A458-S615, Q3	
					7-L198	MOTIFS
					Phospholipase A2 histidine active site C725-C732	MOTIFS
	3552033CD1	1172		N233 N420	Transient receptor:	HMMER_PFAM
			1618 S687	N576 N579		
			S1060	096N		
•			S1076			
			27 42			
			917			
			1984 11031 T1112 T1118 T1132 T1155			
			7857			
_	•	•			rane domains: G51-I75 D397-R425	TMAP
•			*		F/12-V/40 E/80-K806 V822-G842 M852-A872 W934-m962	
					N terminus cytosolic	
				0	receptor potential family	BLIMPS_PRINTS
T				7	A941-T962, F963-F976, V990-M1003	
					ы	BLAST_PRODOM
				. 14	E464-E660	

Table 3 (cont.)

Incyte Polype tide I	Incyte Polypep- tide ID	Amino Acid Residues	Potential Phosphoryl- ation Sites	Potential Glycosyl- ation Sites		Analytical Methods and Databases
					ANK MOTIF REPEAT DM03196 P34586 38-822: 1819-C1009, L583-G619, L702-L807, D114-N144, T9-Q48	BLAST_DOMO
					Aminoacyl-transfer RNA synthetases class-II MC signature 2 All11-L1120	MOTIFS
4778139CD1	)CD1	742	\$104 \$203	N258 N650	OTEIN 2K	BLAST_PRODOM
				T 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7	CALCIUM/CALMODULINDEPENDENT EUKAKYOTIC PD011701: K536-R709	
			S491 S735 T371			
			T619			
4787433CD1	3CD1	577	S70 S119 S176 S319 S337 S544 S550 S560 T135 T356 T521 T534	N31 N57 N65 N68 N108 N345 N352 N546 N558	Sugar (and other) transporter: K120-E538 HM	HMMER_PFAM
			T569 Y10			
					Transmembrane domains: G17-G45 R150-R178 TM L185-Y205 F214-I234 S243-L263 L269-F293 I355-S377 N390-D410 T416-P436 L442-Y462	TMAP
			:		A488-L516 N + Dermining Antonolin	
					anger isoform PR010870 I32-V46	BLIMPS PRINTS
						11 1
7483598CD1	98CD1	462	S24 S56 S90 S242 S243 S393 T282 T391	N100 N331 N436 N441 N457	rter	HMMER_PFAM
					Transmembrane domains: C33-L53 G65-K85 TM T102-A130 N148-L168 A175-V195 W213-Y241 A250-F278 F328-A348 L358-P378 T391-N419 N terminus non-cytosolic	TMAP
*					ACID AMINO PROTEIN TRANSPORTER PERMEASE TRANSMEMBRANE INTERGENIC REGION PUTATIVE PROLINE PD001875: F39-T209	BLAST_PRODOM
ŀ						

Analytical Methods and Databases		SPSCAN	HMMER	нимек_рғам		BLIMPS_PRINTS	BLAST PRODOM	BLAST_PRODOM		ТМАР	HIMBE PFAM
uences, otifs	Transmembrane domains: A2-R29, K73-S101, P167-P190, T194-M219, T237-M261, S720-P741, G748-T776, T776-D797, I815-W841, H849-R877, C891-T911 N-terminus is cytosolic	signal_cleavage: M1-A30	Signal Peptide: Ml-A32	Transient receptor: Y922-H979, R791-L851, D552-W598	: :	Transient receptor potential family PR01097: A920-T941, F942-F955	MELASTATIN 1 PD183973: L1097-C1466		PD018035: M1-L333 PD151509: I803-L1097 PD039592: E454-T652	Transmembrane domains: G51-I75, D397-R425, L602-Y630, F692-V712, W717-I737, N763-N783, Y789-F809, Y818-A846, W913-T941, L970-L987 N-terminus is cytosolic	<b>∥-⊣</b>
Potential Glycosyl- ation Sites				N133 N144 N233 N298 N420 N566 N569 N763 N1054 N1245							
Potential Phosphoryl- ation Sites				\$86 \$212 \$235 \$300 \$366 \$401 \$518 \$548 \$608 \$719 \$858 \$1011 \$1049 \$1040 \$1087 \$1114 \$1166 \$1234 \$1339 \$1410 \$1422 \$1449 \$1450 \$1629 \$1637 \$1140 \$1629 \$1637 \$1140 \$1629 \$1637 \$1140 \$1629 \$1637 \$1140 \$1160	703 T129 T138						S28 S142 S159 T144
Amino Acid Residues				1466	•						222
Incyte Polypep- tide ID				6046484CD1							7481427CD1
ON CO	ი			0							11

\$212 \$233 \$362 \$382 \$318 T348 \$304 \$446	Incyte Polypep- tide ID	Amino Acid Residue	Amino Potential Acid Phosphoryl- Residues ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and
461 S168 S212 S233 S338 S362 S382 S439 T318 T348 S439 T318 T348 S562 S382 S457 T25 T25 T59					Mitochondrial energy transfer proteins BL00215: L13-Q37, I158-G170	BLIMPS_BLOCKS
461 S168 S212 S233 S439 T318 T348 S439 T318 T348 S62 S362 S382 S439 T318 T348 S436 S304 S446 S467 T25 T59					Mitochondrial energy transfer proteins mitoch_carrier.prf: C110-I158	PROFILESCAN
461 S168 S212 S233 S338 S362 S382 S439 T318 T348 S62 S382 S439 T318 T348 S62 S362 S362 S365 S365 S365 S365 S365 S365 S365 S365		·			Mitochondrial energy transfer proteins: P127-A136	MOTIFS
461 S168 S212 S233 S439 T318 T348 S439 T318 T348 S502 S367 T25 T59 S467 T25 T59					Mitochondrial carrier protein signature PR00926: G120-D138, Y168-F186, D11-T24, T24-M38; G73-D93	BLIMPS_PRINTS
461 S168 S212 S233 S439 T318 T348 S439 T318 T348 S502 S362 S362 S362 S362 S362 S364 S446 S467 T25 T59	·				Adenine nucleotide transfer protein PR00927: F8-A20, C51-R72, T84-K96, R111- V124, R146-L167, S207-S222	BLIMPS_PRINTS
461 S168 S212 S233 S338 S362 S382 S439 T318 T348 T348 S502 S146 S304 S446 S467 T25 T59					PROTEIN TRANSPORT TRANSMEMBRANE REPEAT MITOCHONDRION CARRIER MEMBRANE INNER MITOCHONDRIAL ADP/ATP PD000117: S7-A123, Y112-S222	BLAST_PRODOM
461 S168 S212 S233 S338 S362 S382 S439 T318 T348 S62 S146 S304 S446 S467 T25 T59	•		: :		ENERGY TR 2 11-96: F 14-108: 2 116-205: 5 110-208:	BLAST_DOMO
461 S168 S212 S233 S338 S362 S382 S439 T318 T348 502 S146 S304 S446 S467 T25 T59					Transmembrane domains: I164-R187 N-terminus is cytosolic	ТМАР
502 S146 S304 S467 T25 T	83595CD		S212 S362 T318	N98 N163 N288 N344 N380 N381	N288 SULFATE TRANSPORTER PROTEIN TRANSPORT TRANSMEMBRANE AFFINITY GLYCOPROTEIN SULPHATE HIGH DISEASE PD001755: R216-D272, V414-D459	BLAST_PRODOM
502 S146 S304 S467 T25 T				·	SULFATE TRANSPORTERS DM01229 P40879 5-462: T23-A140, Q124-L222 DM01229 P45380 10-468: P130-L222, T23-L138 DM01229 P50443 49-505: I136-K247, T23-S144	BLAST_DOMO
502 S146 S304 S467 T25 T					Transmembrane domains: F6-F26, L31-S51, M62-S82, C91-A111, E128-K156, L161-R186 N-terminus is cytosolic	TMAP
40	88427CD	N. T.		-	Mitochondrial carrier proteins: S361-Y461, S266-Q359, T172-H264	HMMER_PFAM

) EO	Incyte	Amino	Potential	Potential	Signature Seguences,	Analytical
	Doll, men.	7.04	Dhoenhom! -	- ניים היינים	Domaine and Motife	Mother of the
NO:	tide ID	Residues ation	ation Sites	ation Sites	Domains and Mocies	Methods and Databases
13					Mitochondrial energy transfer proteins mitoch_carrier.prf1: L362-1417 mitoch_carrier.prf2: S270-1315	PROFILESCAN
					Mitochondrial energy transfer proteins: P287-1296	MOTIFS
					Mitochondrial carrier protein signature PR00926: G232-R252, V280-Q298, Y325-L343, V369-Q391	BLIMPS_PRINTS
				·	PROTEIN TRANSPORT TRANSMEMBRANE REPEAT MITOCHONDRION CARRIER MEMBRANE INNER MITOCHONDRIAL ADP/ATP PD000117: Y305-R453	BLAST_PRODOM
					MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026 Q01888 126-214: G263-I352 DM00026 P29518 233-310: I271-I352	BLAST_DOMO
					Transmembrane domains: R176-G204, A323- K351, L424-K452 N-terminus is cytosolic	TMAP
14	6972455CD1	261	S6 S26 S135 S198 T2 T36 T64 T84 T92 T165 Y218	N82 N172	N173 signal_cleavage: M1-A23	SPSCAN
					Signal Peptide: M1-P22, M1-A23, M1-A25, M1- G27	HMMER
					Bacterial extracellular solute-binding protein domain: M1-L253	HMMER_PFAM
					Transmembrane domains: T175-A196 N-terminus is non-cytosolic	ТМАР
					Bacterial extracellular solute binding protein signature BL01039: G52-L72, R86-Y118, L90-S101	BLIMPS_BLOCKS
					BACTERIAL EXTRACELLULAR SOLUTE-BINDING PROTEINS, FAMILY 3 DM00557 P27676 32-261: S19-K223 DM00557 P30860 5-241: L37-W252 DM00557 P39174 15-260: L8-W252 DM00557 P45678 11-258: A11-W252	BLAST_DOMO

Analytical			BLAST_DOMO	TMAP.	HMMER PFAM	BLIMPS_BLOCKS	PROFILESCAN	BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_PRODOM
Signature Sequences,	Domains and Motifs	ונה	do SENSITIVE; COTRANSPORTER; SODIUM; CHLORIDE; DM01337 P55011 409-906; V278-G402 DM01337 P55013 381-879; V278-G402 DM01337 A53491 381-879; V278-G402 DM01337 P55014 297-795; V278-G402	brane do 3 L394-Y 1 L937-E us is no	E1-E2 ATPase domain: V268-P324	E1-E2 ATPases phosphorylation site signature BL00154: G284-L301, V442-G478, I480-L498, K624-C634, N695-M735	E1-E2 ATPases phosphorylation site: 1466- F515	P-type cation-transporting atpase superfamily signature PR00119: N309-T323, C484-L498, A711-D721	Sodium/potassium-transporting ATPase signature PR00121: C477-L498, V621-V639	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP BINDING TRANSPORT PUMP CALCIUM MAGNESIUM MEMBRANE PD000132: 1230- D494
Potential	Glycosyl- ation Sites	N11 N342 N440		N540 N669 N781 N819 N848 N867 N875 N1005				٠		
Potential	Phosphoryl- ation Sites	S40 S82 S265 S314 S349 S442 S446 S551 T50 T324 T344		SS S103 S159 S241 S249 S338 S567 S587 S671 S798 S833 S850 S1008 S1028 T78 T97 T172 T375 T490 T664 T701 T784 T859 T871						
Amino	Acid Phosp Residues ation	570		1033						
Incyte	Polypep- tide ID	8077668CD1	•	55120485CD1						
ĒΟ	N S	15		16						

Incyte Polypep- tide ID	Amino Acid Residues	Potential Phosphoryl- ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
			3	E1-E2 ATPASES PHOSPHORYLATION SITE DM00115 P22189 49-801: Q597-P738, W196-K322, P389-P447, S671-L723 DM00115 P37278 58-755: I192-I743, V822-K840 DM00115 P47317 26-695: F600-E749, I230-K559, K815-E853 DM00115 P54707 97-825: I232-I743, L813-E853	BLAST_DOMO
			·	E1-E2 ATPases phosphorylation site: D486- T492	MOTIFS
3112883CD1	496	S40 S211 S241 S251 S274 S275 S463 S466 S482 S493 T47 T117	N45 N166 N256	N256 Reduced folate carrier domain: S10-V441	HMMER_PFAM
				153-Y76 Y79- (194 K276- 1395 L405-	ТМАР
				PROTEIN REDUCED TRANSPORTER OLATE BINDING TRANSPORT METHOTREXATE -E230, G327-S493, F262-W303	BLAST_PRODOM
4253888CD1	573	S8 S35 S77 S88 S106 S137 S229 S304 S321 S340 S379 T10 T27 T148 T194 T401 Y115	N78 N466	85 V189-R217 E343-K369 G493-D521	TMAP
				Divalent cation transporter domain: L199-S335, Y413-H559	HMMER_PFAM
7479974CD1	573	S69 S138 S198 S221 S261 S355 S473 S478 S509 S543 T24 T159 T176 T401 T406 T474 T499	N80 N174 N294, N491 N529 N558	Pase domain:	HMMER_PFAM

Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	MOTIFS	ТМАР	HIMER_PFAM	, TMAP	BLIMPS_BLOCKS	BLIMPS_PRINTS	BLIMPS_PRINTS	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	PLASMID ARSENICAL PUMPDRIVING ATPASE HYDROLASE RESISTANCE ATPBINDING ARSA PD006335: D460-P568	NIFH/FRXC FAMILY DM00105 P08690 7-180: Y20-E191 DM00105 P08690 326-473: L333-K472 DM00105 P30632 17-190: K19-S167	ATP/GTP-binding site motif A (P-loop): G25-T32, G337-T344	Transmembrane domains: D8-S36, R44-I72, C96-R123, S133-I158, L171-F194 N-terminus is non-cytosolic		Transmembrane domains: P84-K112, Y459-W487, P528-Y553, G564-K584, L592-C612, I624-Y652 N-termincus is cytosolic	E1-E2 ATPases phosphorylation site proteins: BL00154: D231-L271, T386-S409, K131-L141	P-type cation-transporting atpase superfamily signature PR00119: A247-D257	H+-transporting ATPase (proton pump) signature PR00120: T162-A180	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP-BINDING PROTEIN PROBABLE CALCIUM TRANSPORTING CALCIUM TRANSPORT PD004657: A423-P661	ATPASE HYDROLASE TRANSMEMBRANE PHOSPHORYLATION ATP-BINDING PROBABLE PROTEIN CALCIUM TRANSPORTING CALCIUM TRANSPORT PD149930: C363-F422
Forential Glycosyl- ation Sites						N700 N732					
Potential Phosphoryl- ation Sites	n			S42 S85 S121 S198 S215 S227 S233 S239	·	S5 S62 S109 S115 S185 S312 S409 S476 S556 S706 S734 T28 T20 T78 T162 T201 T227 T335 T534 T674 T695	·		-		
Amino Acid Residues			,	248		7,61					,
incyte Polypep- tide ID				7483850CD1		5508353CD1					
3 1 1 1 1 1 1 1 1	19			20		21					

Analytical Methods and Databases	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	MOTIFS	HMMER_PFAM	TMAP		PROFILESCAN	BLAST_PRODOM	BLAST_DOMO	SYSTEMS BLAST_DOMO:	BLAST_DOMO	BLAST_DOMO	MOTIFS	HMMER_PFAM
Signature Sequences, Domains and Motifs	_		ATPASE; CALCIUM; TRANSPORTING; DM02405 S51243 356-1267: E59-F486	ATPASE; CALCIUM; TRANSPORTING; DM02405 P39524 236-1049: Y58-D287, L332-N487	ATP/GTP-binding site motif A (P-1oop): G331-T338, A699-S706	Binding-protein-dependent transport system: A112-Y185	Transmembrane domains: A20-N48, V70-S98, T156-R183, Y185-A208	N-terminus is non-cytosolic	Binding-protein-dependent transport systems inner membrane component: V105-T161	PROTEIN TRANSPORT TRANSMEMBRANE PERMEASE MEMBRANE AMINO ACID INNER SYSTEM TRANSPORTER ABC PD001196: E7-G111	BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388 P10345 3-214: L9-R215	BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388   P42399   12-220: S12-R215	BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388 P45023 17-232: P14-R215	BINDING-PROTEIN-DEPENDENT TRANSPORT SYSTEMS INNER MEMBRANE DM00388   P42200   15-226: P14-R215	Binding-protein-dependent transport systems MOTIFS inner membrane component signature: L113-P141	Transmembrane amino acid transporter protein: A48-C433
Potential Glycosyl- ation Sites									·							
Potential Phosphoryl- ation Sites				ek :	- 1	S4 T148			-							S249 S254 S413 S420 T451 T458
Amino Poten Acid Phosp Residues ation						219										463
Incyte Polypep- tide ID						8543628CD1										7482754CD1
0.19 10.00 10.00	21					22										23

ON OI OI OI	Incyte Polypep- tide ID	Amino Acid Residues	Potential Phosphoryl- ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
1				·	Transmembrane domains: L24-N44, G54-V74, R93-R121, D148-R168, H180-T200, S249-S274, G298-R326, W353-M373, S377-P397 N-terminus is non-cytosolic	тмар
- }					Transmembrane four family mignature PR00259: F59-V82, L73-A99, V302-V328	BLIMPS_PRINTS
J						BLAST_PRODOM
	3794818CD1	1043	\$36 \$153 \$206 \$372 \$388 \$402 \$536 \$633 \$700	N69 N344 N451	eptide:	нммек
			S721 S867	000		
			S891 S901 S1014 S1036 T157 T192 T210 T421 T491			
		·	T604 T673 T916			
					signal_cleavage: M1-G22	SPSCAN
I					Ligand-gated ion channel: H574-E852	HMMER_PFAM
			•		Transmembrane domains: H574-R598, P640-V667, T823-L847 N terminus is non-cytosolic	ТМАР
				1	NMDA receptor signature PR00177: F493-L521, T577-G602, L644-D671, F831-F855	BLIMPS_PRINTS
1				<b></b>	R32184_2 IONOTROPIC GLUTAMATE RECEPTOR PD156309: A77-Y477	BLAST_PRODOM
				<u> </u>	RECEPTOR GLUTAMATE SUBUNIT SIGNAL PRECURSOR I CHANNEL IONIC TRANSMEMBRANE POSTSYNAPTIC PD000500: M570-E852	BLAST_PRODOM
ij						

	Incyte Polypep- tide ID	Amino Acid Residues	Potential Phosphoryl- ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
1					RECEPTOR SIGNAL GLUTAMATE SUBUNIT PROTEIN TRANSMEMBRANE CHANNEL IONIC PD000273: G478-A563, G728-V817	BLAST_PRODOM
}					GLUTAMATE RECEPTOR DM00247 Q03391 640-919: T631-S901, C964-P980	BLAST_DOMO
					DM00247 P35436 615-886: T631-F856 DM00247 Q01098 613-882: T631-E904 DM00393 Q01097 377-614: G387-F628	
	4717525CD1	480	S4 S23 S56 S105		Mitochondrial carrier proteins:	HMMER_PFAM
			T170 T410		0150-0100 (0191-501)	
					EF hand: Q117-H145, R13-L41, R81-L109	HMMER_PFAM
					Mitochondrial energy transfer proteins BL00215: V190-Q214, I369-G381	BLIMPS_BLOCKS
					al	PROFILESCAN
					I320-S371, K187-L241, V279-S331	
					Mitochondrial carrier protein signature PR00926: Q188-T201, T201-V215, G244-E264, C333-Q351, Y379-Y397, G327-Q349	BLIMPS_PRINTS
					Graves disease carrier protein signature PR00928: Q274-Q294, P205-I225, Y263-S287, I369-V389	BLIMPS_PRINTS
	-				TRANSPORT TRANSMEMBRANE CARRIER INNER MITOCHONDRIAL ADP/ATP PD000117: K187-S466	BLAST_PRODOM
					MITOCHONDRIAL ENERGY TRANSFER PROTEINS DM00026 S57544 26-107: V190-I270 DM00026 Q01888 38-124: V190-I270	BLAST_DOMO
			·		EF-hand calcium-binding domain: D22-L34 D90-1102	MOTIFS
						Cathorne Company

Table 3 (cont.)

Analytical Wethods and Databases	тмар	TMAP	BLIMPS_BLOCKS	BLAST_DOMO	HWMER_PFAM	TWAP
Signature Sequences, Domains and Motifs	Transmembrane domains:	Transmembrane domains:     V9-H37, E85-S105, L114-L134,     L197-T225, G290-W317, V342-A362,     X379-G399, T433-L453, N460-L480     N terminus is cytosolic.	glpT family of transport BL00942: T29-K41, N82-L124, W171-V190, F211-P247, E281-Y321, L339-D356	GLPT FAMILY OF TRANSPORTERS  DM02439 P37948   1-403: K84-H244,  L305-L446  DM02439   P09836   1-401: L87-E234,  P295-A426, S16-K41  DM02439   P08194   1-403: L87-D256,  Q251-A444, R22-145	N455 HCO3- transporter family: A571-L745, H240-F546, R119-L208	Transmembrane domains:     V282-G304, L330-I358, T373-M393,     I403-I423, H478-Y499, V514-E542,     S564-L592, T604-A630, S672-Y700,     L748-I776  N terminus is non-cytosolic.
Potential Glycosyl- ation Sites	N6 N171 N371 N376	N53 N62 N68		5	N52 N388 N455 F N463	T
cential sphoryl- on Sites	s8 S38 S161 S240 S253 S331 S389 S510 T378	S39 S66 S263 S267 S329 S421 T338 T418			S21 S171 S305 S399 S442 S469 S564 T6 T20 T56 T130 T174 T198 T200 T400 T420 T438 T704 Y791	
no d idues	518	501			801	
	5091793CD1	5945527CD1		·	6941124CD1	
EQ NO:		27		·	28	

Domains and Motifs Anion exchangers far
BL00219: S448-T483, P243-V282, T289-D312, L342-F380, A382-Y429, Q481-S534, A567-W608, D609-E647, H653-F698, Y700-T743, L748-I787
Anion exchanger signature PR00165: G253-I275, K284-G304, A371-S390,
A482-L501, I517-G537
ANION EXCHANGE TRANSMEMBRANE GLYCO-PROTEIN
LIPOPROTEIN PALMITATE BICARBONATE
PD001455: T483-T743, F752-S457,
152M-061A
DM02294 P48751 601-1229: A190-E798 DM02294 P02730 311-908: T483-E798
DM02294 A42497 403-1027: F252-E798
DM02294   P04920   602-1237:
signal_cleavage:
M1-A34
Signal Peptide: M9-A34
Transmembrane domains:
R8-L36 V39-V63 Q71-T99
N terminus is non-cytosolic.

EO	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
a ç	Polypep-	Acid	Acid Phosphoryl-	Glycosyl- ation Sites	Domains and Motifs	Methods and Databases
30	6991750CD1	2701		N56 N218 N319	N56 N218 N319 Transmembrane domains:	TMAP
			S40 S1367	N330.N364	1965-S986, E1281-T1300,	
			S551	N396 N452	N1670-S1692, V1836-W1864,	
			S1603 S193	N480 N539	V2276-N2292, A2507-F2534	
			S1710 S1793	N607 N667	N terminus is non-cytosolic.	
			1824	N691 N772		
			S1866 S274	N828 N875		
			S1879 S230	N885 N921		
			S1893 S1915	N949 N964		
			S651 S1922 S307	51922 S307 N988 N1135		
			S1952 S1978	N1153 N1166	Adenosine and AMP deaminase signature	MOTIFS
			S461 S2029 S382 N1241	N1241 N1259	S2386-P2392	
	•		52090.	N1273 N1305		
			S495	N1323 N1372		
			8 52438	N1479 N1525		
			S792 S2464 S790 N1528	N1528 N1618		
			S2496	N1624 N1892		
				N1969 N2134		
			S809	N2342 N2428	•	
			3	N2510 N2596		
			S856 S2631	N2625		

EQ ID NO:	Incyte Polypep- tide ID	Amino Acid Residues	Amino Potential Acid Phosphoryl- Residues ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
30			8 1 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6 6		Sodium/Calcium Exchanger Chain DM05297 P48765 6-969: V263-I492 P=2.8-09	BLAST_DOMO
31	71726948CD1	610	S114 S269 S317 S375 S377 S563 S576 S602 T3 T41 T53 T74 T158 T312 T364 T483 T491 T557	N260 N481 N485 N606	Signal Peptide: M47-P73	HMER
					Sodium: solute symporter family: F45-G449	HMMER_PFAM
	·				Transmembrane domains: P4-Y32, T53-G73, I84-L104, L121-Y148, G159-L179, V190-M210, R239-I256, R273-L301, S385-L405, A412-1432, S439-G459, N515-T543 N terminus is non-cytosolic.	TMAP
					Sodium: solute symporter BL00456: L22-S76, I98-V127, T158-G212	BLIMPS_BLOCKS
					Sodium: solute symporter family signatures: N155-A201	PROFILESCAN

EQ ID NO:	Incyte Polypep- tide ID	Amino Acid Residues	Amino Potential Acid Phosphoryl- Residues ation Sites	Potential Glycosyl- ation Sites	Signature Sequences, Domains and Motifs	Analytical Methods and Databases
31					TRANSMEMBRANE TRANSPORT PERMEASE SODIUM SYMPORT PROLINE GLYCOPROTEIN PD000991: F45-P234, L129-G449	BLAST_PRODOM
					SYMPORTER SODIUM IODIDE THYROID PD024705: A451-P552	BLAST_PRODOM
					SODIUM: SOLUTE SYMPORTER FAMILY DM00745 JC2382 3-485:Y15-W455	BLAST_DOMO
					DM00745 P45174 3-495:T9-W455 DM00745 P31448 1-494:F18-G449 DM00745 P44963 1-483:V23-Y463	
32	7487393CD1	552	S60 S68 S167 S276 S408 S475	N39 N56 N62 N102 N377	Sugar (and other) transporter: I18-V530	HMMER_PFAM
			5537 T58 T133 T311 T323 T391 T526			
					Transmembrane domains: V10-E38, K145-G164, I174-L202, M232-A252, Q262-S282, K345-I368,	ТМАР
					G3/3-L39/, F412-L440, S4/3-L496, P497-L514 N terminus is non-cytosolic.	
					ORGANIC TRANSPORTERLIKE TRANSPORT PROTEIN RENAL ANION TRANSPORTER CATIONIC KIDNEY SPECIFIC SOLUTE PD151320: N102-K145	BLAST_PRODOM

Polynucleotide SEQ ID NO:/ Incyte ID/ Sequence Length	Sequence Fragments
33/7484831CB1/2365	1-325, 94-240, 106-242, 106-244, 106-245, 106-514, 106-515, 106-582, 106-593, 106-597, 106-612, 109-245, 110-245, 128-770, 160-516, 164-267, 176-549, 181-440, 184-581, 234-317, 234-409, 252-770, 275-469, 275-473, 275-570, 275-636, 275-770, 336-1038, 338-1038, 377-543, 413-622, 414-1038, 495-1038, 504-764, 504-1009, 521-952, 521-1034, 521-1038, 523-680, 704-974, 781-1246, 824-1377, 884-1127, 904-1254, 916-1533, 940-1038, 958-1226, 978-1038, 990-1038, 1018-1637, 1031-1722, 1064-1463, 1070-1211, 1086-1149, 1110-1336, 1110-1386, 1110-1387, 1110-1391, 1110-1462, 1113-1382, 1139-1622, 1144-1622, 1153-1629, 1178-1434, 1178-1749, 1210-1694, 1262-1309, 1445-1969, 1445-2000, 1455-1830, 1463-1630, 1478-1701, 1519-1790, 1526-1952, 1582-1838, 1669-1886, 1676-1846, 1772-2205, 1846-2117, 1847-2060, 1890-2209, 1934-2041, 1938-2315, 1950-2365
34/2477266CB1/3400	1-89, 1-306, 13-348, 13-665, 16-468, 23-270, 23-271, 23-404, 23-512, 27-306, 35-386, 35-589, 37-693, 124-795, 240-510, 317-524, 366-999, 459-996, 473-1144, 492-1041, 493-1060, 505-1019, 761-1360, 798-1025, 798-1268, 872-1360, 875-1143, 988-1278, 996-1252, 996-1612, 1024-1265, 1032-1623, 1054-1507, 1078-1341, 1192-1726, 1213-1608, 1218-1469, 1253-1798, 1332-1567, 1332-1800, 1345-1908, 1387-1693, 1419-1981, 1434-1737, 1436-1726, 1518-2165, 1524-2132, 1562-1836, 1775-2024, 1779-2022, 1797-2084, 1890-2052, 1890-2134, 1890-2438, 1913-2196, 1942-2197, 1949-2528, 1952-2509, 1954-2029, 1981-2621, 1994-2252, 1994-2252, 1994-2259, 2039-2147, 2040-2319, 2062-2315, 2142-2732, 2205-2493, 2238-2536, 2265-2795, 2275-2544, 2277-2528, 2302-2718, 2337-2622, 2380-2920, 2444-2841, 2467-2711, 2472-2728, 2472-2955, 2484-2753, 2492-2641, 2503-2746, 2512-3142, 2517-3095, 2555-3131, 2568-3246, 2590-2931, 2605-2903, 2643-2973, 2646-2931, 2656-2902, 2673-3284, 2725-3400, 22673-3388, 2795-3082, 2880-3395, 2818-3355, 2818-3355, 2832-3051, 2832-3088, 2851-3007, 2851-3015, 2851-3028, 2854-3120, 2868-3336, 2871-3133, 2871-3195, 2884-3181, 2892-3104
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Polynucleotide	Sequence Fragments
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	2451-3023, 2455-3024, 2470-3024, 2479-3024, 2480-3024, 2482-3024, 2484-3024, 2488-3024, 2489-3024, 2491-3024, 2501-3024,
	2508-3024, 2511-3024, 2522-3024, 2582-3024, 2583-3024, 2587-3024, 2590-3024, 2592-2877, 2592-3022, 2592-3024, 2596-3024,
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	2770-3021, 2773-3203, 2802-3279, 2812-3004, 2831-3008, 2831-3037, 2831-3117, 2831-3220, 2831-3247, 2831-3275, 2831-3300,
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	3427-4044, 3444-3811, 3463-4149, 3472-4150, 3485-4080, 3498-3527, 3510-3811, 3521-4030, 3547-3782, 3552-4035, 3552-4039,
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	4219-4458, 4295-4458
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	874, 467-874, 481-874, 498-874, 508-874, 563-874, 580-874, 1464-1728, 1464-1973, 1660-1896, 1660-1919, 1935-2155, 2166-2453,
	2166-2722, 2178-2456
37/4787433CB1/1924	1-77, 1-100, 1-480, 154-1023, 513-772, 719-980, 719-1247, 729-984, 805-1016, 843-1529, 848-1456, 849-1459, 902-1530, 902-1552,
	1527, 1075-1646, 1076-1628, 1082-1734, 1091-1693, 1095-1701, 1099-1339, 1104-1681, 1104-1707, 1106-1674, 1112-1332, 1119-
	1583, 1123-1765, 1129-1683, 1169-1608, 1200-1626, 1207-1745, 1218-1832, 1230-1611, 1232-1669, 1244-1741, 1256-1758, 1265-
· ·	1584, 1273-1758, 1280-1557, 1284-1765, 1303-1924, 1356-1592, 1356-1924, 1359-1585, 1359-1833, 1366-1917, 1382-1911, 1387-
	1913, 1518-1924, 1575-1909, 1776-1886
38/7483598CB1/1797	1-808, 8-302, 13-417, 14-808, 179-808, 490-808, 491-808, 732-1017, 881-1226, 881-1227, 891-1105, 1157-1429, 1157-1700, 1184-
	11707, 1295-1760, 1324-1759, 1359-1744, 1359-1749, 1359-1760, 1362-1760

Polynucleotide	Sequence Fragments
SEO ID NO:/	
Incyte ID/ Sequence	
Length	
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	1651, 1506-1732, 1683-2339, 1700-2334, 1724-2174, 1724-2423, 1792-2400, 1804-2530, 1822-2278, 1834-2323, 1840-2516, 1878-
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	2530, 1976-2515, 1977-2497, 1980-2431, 2012-2660, 2020-2596, 2046-2671, 2049-2576, 2083-2578, 2095-2752, 2095-2763, 2098-
	2659, 2105-2794, 2114-2787, 2129-2908, 2162-2720, 2163-2689, 2225-3024, 2229-2763, 2241-2925, 2358-3057, 2374-3044, 2444-
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40/143935CB1/2773	1-217, 1-462, 1-942, 16-601, 34-217, 39-566, 176-217, 206-667, 217-756, 217-769, 274-800, 288-919, 316-814, 316-956, 328-967, 334-
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:	3113-3264, 3114-3261
64/7487393CB1/1659	64/7487393CB1/1659 1-402, 1-1659, 307-506, 416-506, 547-860, 547-863, 547-867, 1147-1598

Polynucleotide SEQ	Incyte Project TD:	Representative Library
ID NO:	,,,	Troprosonative Dioral y
33	7484831CB1	LIVRNON08
34	2477266CB1	LIVRNON08
35	3552033CB1	BRAHTDK01
36	4778139CB1	PROSTUS19
37	4787433CB1	PGANNOT01
38	7483598CB1	BRAITUT29
39	7484823CB1	TESTNOC01
40	143935CB1	BRACDIK08
41	5923789CB1	BRAIFET02
42	6046484CB1	BRACNOK02
44	7483595CB1	TESTNOC01
45	3788427CB1	BONEUNR01
46	6972455CB1	BMARUNR02
47	8077668CB1	ADRETUE02
	55120485CB1	BRAITUT29
49	3112883CB1	BRSTNOT03
	425388SCB1	ADRETUE02
	74S3850CB1	LIVRDIT06
	5508353CB1	NERDTDN03
	8543628CB1	BMARUNR02
	74S2754CB1	PTHYTMN05
	3794818CB1	KIDEUNE02
	4717525CB1	KIDEUNE02
	5091793CB1	LUNGTUT08 ~
	5945527CB1	SINTNOR01
	6941124CB1	FTUBTUR01
	6972530CB1	BMARUNR02
	6991750CB1	BRAIFER06
53		KIDNNOT32

Library	Vector	Library Description
ADRETUE02	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from right adrenal tumor tissue removed from a 49-year-old Caucasian male during unilateral adrenalectomy. Pathology indicated adrenal cortical carcinoma comprising nearly the entire specimen. The tumor was attached to the adrenal gland which showed mild cortical atrophy. The tumor was encapsulated, being surrounded by a thin (1-3 mm) rim of connective tissue. The patient presented with adrenal cancer, abdominal pain, pyrexia of unknown origin, and deficiency anemia. Patient history included benign hypertension. Previous surgeries included adenotonsillectomy. Patient medications included aspirin, calcium, and iron. Family history included atherosclerotic coronary artery disease in the grandparent(s).
BMARUNR02 PIGEN	PIGEN	This random primed library was constructed using RNA isolated from an untreated SH-SY5Y cell line derived from bone marrow neuroblastoma tumor cells removed from a 4-year-old Caucasian female.
BONEUNR01	PCDNA2.1	This random primed library was constructed using pooled cDNA from two different donors. cDNA was generated using mRNA isolated from an untreated MG-63 cell line derived from an osteosarcoma tumor removed from a 14-year-old Caucasian male (donor A) and using mRNA isolated from sacral bone tumor tissue removed from an 18-year-old Caucasian female (donor B) during an exploratory laparotomy and soft tissue excision. Pathology indicated giant cell tumor of the sacrum in donor B. Donor B's history included pelvic joint pain, constipation, urinary incontinence, unspecified abdominal/pelvic symptoms, and a pelvic soft tissue malignant neoplasm. Family history included prostate cancer in donor B.
BRACDIK08	PSPORT1	This amplified and normalized library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 57-year-old Caucasian male who died from a cerebrovascular accident. Serologies were negative. Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day for 40 years).

### Table (

Library	Vector	Library Description
BRACNOK02	PSPORT1	This amplified and normalized library was constructed using RNA isolated from posterior cingulate tissue removed from an 85-year-old Caucasian female who died from myocardial infarction and retroperitoneal hemorrhage. Pathology indicated atherosclerosis, moderate to severe, involving the circle of Willis, middle cerebral, basilar and vertebral arteries; infarction, remote, left dentate nucleus; and amyloid plaque deposition consistent with age. There was mild to moderate leptomeningeal fibrosis, especially over the convexity of the frontal lobe. There was mild generalized atrophy involving all lobes. The white matter was mildly thinned. Cortical thickness in the temporal lobes; both maximal and minimal, was slightly reduced. The substantia nigra pars compacta appeared mildly depigmented. Patient history included COPD, hypertension, and recurrent deep venous thrombosis. 6.4 million independent clones from this amplified library were normalized in one round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791.
BRAHTDK01	PSPORT1	This amplified and normalized library was constructed using pooled RNA isolated from archaecortex, anterior and posterior hippocampus tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver. 7.6x10e5 independent clones from this amplified library were normalized in 1 round using conditions adapted Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used
BRAIFER06	PCDNA2.1	This random primed library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. Serologies were negative.
BRAIFET02	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus, who was stillborn with a hypoplastic left heart at 23 weeks' gestation.
BRAITUT29	pINCY	Library was constructed using RNA isolated from brain tumor tissue removed from the parietal lobe of a 43-year-old female during excision of a cerebral meningeal lesion. Pathology indicated high grade glioma. Family history included acute myocardial infarction, atherosclerotic coronary artery disease, benign hypertension, and hyperlipidemia.

### Table (

Library	Vector	Library Description
BRSTNOT03	PSPORT1	Library was constructed using RNA isolated from diseased breast tissue removed from a 54-year-old Caucasian female during a bilateral radical mastectomy. Pathology for the associated tumor tissue indicated residual invasive grade 3 mammary ductal adenocarcinoma. Patient history included kidney infection and condyloma acuminatum. Family history included benign hypertension, hyperlipidemia and a malignant neoplasm of the colon.
FTUBTUR01	PCDNA2.1	This random primed library was constructed using RNA isolated from fallopian tube tumor tissue removed from an 85-year-old Caucasian female during bilateral salpingo-oophorectomy and hysterectomy. Pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenocarcinoma, which was confined to the mucosa without mural involvement. Endometrioid carcinoma in situ was also present. Pathology for the associated uterus tumor indicated focal endometrioid adenocarcinoma in situ and moderately differentiated invasive adenocarcinoma arising in an endometrial polyp. Metastatic endometrioid and serous adenocarcinoma was present at the cul-de-sac tumor. Patient history included medullary carcinoma of the thyroid and myocardial infarction.
KIDEUNE02	pINCY	This 5' biased random primed library was constructed using RNA isolated from an untreated transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue (Invitrogen). The cells were transformed with adenovirus 5 DNA.
KIDNNOT32	pINCY	Library was constructed using RNA isolated from kidney tissue removed from a 49-year-old Caucasian male who died from an intracranial hemorrhage and cerebrovascular accident. Patient history included tobacco abuse.
LIVRDIT06	pINCY	This library was constructed using RNA isolated from diseased liver tissue removed from a 35-year-old Caucasian male during needle biopsy of the liver. Patient history included hepatitis C.
LIVRNON08	pINCY	This normalized library was constructed from 5.7 million independent clones from a pooled liver tissue library. Starting RNA was made from pooled liver tissue removed from a 4-year-old Hispanic male who died from anoxia and a 16 week female fetus who died after 16-weeks gestation from anencephaly. Serologies were positive for cytolomegalovirus in the 4-year-old. Patient history included asthma in the 4-year-old. Family history included taking daily prenatal vitamins and mitral valve prolapse in the mother of the fetus. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

### Table (

Library	Vector	Library Description
LUNGTUTO8	pINCY	Library was constructed using RNA isolated from lung tumor tissue removed from a 63-year-old Caucasian male during a right upper lobectomy with fiberoptic bronchoscopy. Pathology indicated a grade 3 adenocarcinoma. Patient history included atherosclerotic coronary artery disease, an acute myocardial infarction, rectal cancer, an asymtomatic abdominal aortic aneurysm, tobacco abuse, and cardiac dysrhythmia. Family history included congestive heart failure, stomach cancer, and lung cancer, type II diabetes, atherosclerotic coronary artery disease, and an acute myocardial infarction.
NERDTDN03	pINCY	This normalized dorsal root ganglion tissue library was constructed from 1.05 million independent clones from a dorsal root ganglion tissue library. Starting RNA was made from dorsal root ganglion tissue removed from the cervical spine of a 32-year-old Caucasian male who died from acute pulmonary edema, acute bronchopneumonia, bilateral pleural effusions, pericardial effusion, and malignant lymphoma (natural killer cell type). The patient presented with pyrexia of unknown origin, malaise, fatigue, and gastrointestinal bleeding. Patient history included probable cytomegalovirus infection, liver congestion, and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, respiratory failure, pneumonia of the left lung, natural killer cell lymphoma of the pharynx, Bell's palsy, and tobacco and alcohol abuse. Previous surgeries included colonoscopy, closed colon biopsy, adenotonsillectomy, and nasopharyngeal endoscopy and biopsy. Patient medications included Diflucan (fluconazole), Deltasone (prednisone), hydrocodone, Lortab, Alprazolam,
		Reazodone, ProMace-Cytabom, Etoposide, Cisplatin, Cytarabine, and dexamethasone. The patient received radiation therapy and multiple blood transfusions. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
PGANNOT01 PSPORT1	PSPORTI	Library was constructed using RNA isolated from paraganglionic tumor tissue removed from the intra-abdominal region of a 46-year-old Caucasian male during exploratory laparotomy. Pathology indicated a benign paraganglioma and was associated with a grade 2 renal cell carcinoma, clear cell type, which did not penetrate the capsule. Surgical margins were negative for tumor.

Library	Vector	Library Description
PROSTUS19	pINCY	This subtracted prostate tumor tissue library was constructed using 2.36 million clones from the PROSTUT13 library and was subjected to two round sof subtraction hybridization with 2.36 million clones from EPIPNOT01 library. The starting library for subtraction was constructed using RNA isolated from prostate tumor tissue removed from a 59-year-old Caucasian male during a radical prostatectomy with regional lymph node excision. Pathology indicated adenocarcinoma (Gleason grade 3+3) involving the prostate peripherally with invasion of the capsule. Adenofibromatous hyperplasia was present. The patient presented with elevated prostate-specific antigen (PSA). Patient history included diverticulitis of colon, asbestosis, and thrombophlebitis. Family history included benign hypertension, multiple myeloma, hyperlipidemia, and rheumatoid arthritis. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991) 19:1954 and Bonaldo, et al. Genome Research (1996) 6:791.
PTHYTMN05 pINCY	pINCY	Library was constructed using RNA isolated from parathyroid tissue removed from a 44-year-old Caucasian male during a partial parathyroidectomy. Pathology for the matched tumor tissue indicated parathyroid carcinoma (grade 1 of 4) forming a partially cystic tan mass. Both capsular and vascular invasion were present. The patient presented with unspecified parathyroid disorder and calcium metabolism disorder. Patient history included kidney calculus and obesity. Previous surgeries included vasectomy and parathyroid surgery. Family history included emphysema in the mother; type II diabetes in the father; and type I diabetes and hyperlipidemia in the sibling(s).
SINTNOR01	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine tissue removed from a 31-year-old Caucasian female during Roux-en-Y gastric bypass. Patient history included clinical obesity.
TESTNOC01	PBLUESCRIPT	PBLUESCRIPT This large size fractionated library was constructed using RNA isolated from testicular tissue removed from a pool of eleven, 10 to 61-year-old Caucasian males.

Parometer Threshold	s, Foster City, CA.	s, Foster City, CA; Mismatch <50% ena, CA.	s, Foster City, CA.	Altschul, S.F. et al. (1990) J. Mol. Biol. ESTs: Probability value= 1.0E-8 215:403-410; Altschul, S.F. et al. (1997) or less Nucleic Acids Res. 25:3389-3402. Full Length sequences: Probability value= 1.0E-10 or less	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489. Full Length sequences: fastx score=100 or greater fastx score=100 or greater	Henikoff, S. and J.G. Henikoff (1991) Nucleic Probability value= 1.0E-3 or less Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, B.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a greater
Reference	Applied Biosystems, Foster City, CA.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Applied Biosystems, Foster City, CA.	Altschul, S.F. et al. (1990) J. Mol. 215:403-410; Altschul, S.F. et al. Nucleic Acids Res. 25:3389-3402.	Pearson, W.R. and D.J. Lipm Natl. Acad Sci. USA 85:2444 W.R. (1990) Methods Enzym and Smith, T.F. and M.S. Wa Adv. Appl. Math. 2:482-489.	Henikoff, S. and J.G. Henikoff (1991) Acids Res. 19:6565-6572; Henikoff, J S. Henikoff (1996) Methods Enzymol 266:88-105, and Attwood, T.K. et al. Chem. Inf. Comput. Sci. 37:417-424.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Sonnhammer, B.L.L. e (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World Vie
Description	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	A Fast Data Finder useful in comparing and annotating armino acid or nucleic acid sequences.	A program that assembles nucleic acid sequences.	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.
Program	ABI FACTURA	ABIPARACEL FDF	ABI AutoAssembler	BLAST	FASTA	BLIMPS	HMMER

Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality scorez GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater, Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Intl. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	
Motifs	A program that searches armino acid sequences for pattems that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	7-221; ige

### What is claimed is:

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1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32,
- b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-32,
- a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
- An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the
   group consisting of SEQ ID NO:1-32.
  - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
  - 4. An isolated polynucleotide encoding a polypeptide of claim 2.
  - 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64.
- 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.
  - 7. A cell transformed with a recombinant polynucleotide of claim 6.
  - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
  - 9. A method of producing a polypeptide of claim 1, the method comprising:
  - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

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- b) recovering the polypeptide so expressed.
- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

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- 11. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 12. An isolated polynucleotide selected from the group consisting of:
- a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:33-64,
- b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO.33-64.
- c) a polynucleotide complementary to a polynucleotide of a),
- d) a polynucleotide complementary to a polynucleotide of b), and
- e) an RNA equivalent of a)-d).
- 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.

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- 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
- b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

- 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- a) amplifying said target polynucleotide or fragment thereof using polymerase chain

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reaction amplification, and

- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 5 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
  - 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.

19. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment the composition of claim 17.

- 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
  - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
  - b) detecting agonist activity in the sample.
- 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
  - 22. A method for treating a disease or condition associated with decreased expression of functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 21.
  - 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
    - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- 30 b) detecting antagonist activity in the sample.
  - 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
- 35 25. A method for treating a disease or condition associated with overexpression of

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functional TRICH, comprising administering to a patient in need of such treatment a composition of claim 24.

26. A method of screening for a compound that specifically binds to the polypeptide of 5 claim 1, the method comprising:

- combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
- b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
  - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
  - 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
  - exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
  - b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
  - 29. A method of assessing toxicity of a test compound, the method comprising:
  - a) treating a biological sample containing nucleic acids with the test compound,
  - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under

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conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,

- c) quantifying the amount of hybridization complex, and
  - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

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- 30. A diagnostic test for a condition or disease associated with the expression of TRICH in biological sample, the method comprising:
  - a) combining the biological sample with an antibody of claim 11, under conditions
     suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
  - b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
  - 31. The antibody of claim 11, wherein the antibody is:
- 20 a) a chimeric antibody,
  - b) a single chain antibody,
  - c) a Fab fragment,
  - d) a F(ab')<sub>2</sub> fragment, or
  - e) a humanized antibody.

- 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
- 33. A method of diagnosing a condition or disease associated with the expression of TRICH in a subject, comprising administering to said subject an effective amount of the composition of claim 32.
  - 34. A composition of claim 32, wherein the antibody is labeled.
- 35. A method of diagnosing a condition or disease associated with the expression of in a subject, comprising administering to said subject an effective amount of the

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composition of claim 34.

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36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:

- a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
  - b) isolating antibodies from said animal, and
- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
  - 37. A polyclonal antibody produced by a method of claim 36.
- 15 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
  - 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
    - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-32, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
      - b) isolating antibody producing cells from the animal,
      - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
- 25 d) culturing the hybridoma cells, and
  - e) isolating from the culture monoclonal antibody which binds specifically to a
    polypeptide comprising an amino acid sequence selected from the group consisting
    of SEQ ID NO:1-32.
- 30 40. A monoclonal antibody produced by a method of claim 39.
  - 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
  - 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.

43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

- 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in a sample, the method comprising:
- a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 in the sample.
- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32 from a sample, the method comprising:
- a) incubating the antibody of claim 11 with a sample under conditions to allow specific binding of the antibody and the polypeptide, and
- b) separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-32.
- 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 13.
  - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
  - a) labeling the polynucleotides of the sample,
  - b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
  - c) quantifying the expression of the polynucleotides in the sample.
  - 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is polynucleotide of claim 12.

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- 49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.
- 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
- 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
  - 52. An array of claim 48, which is a microarray.
- 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
  - 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
  - 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
    - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
      - 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
- 30 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
  - 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
  - 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.

35

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	61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
	62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
5	63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
	64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
10	65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10
	66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11
	67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12
15	68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13
	69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14
20	70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15.
20	71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16.
	72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17.
25	73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18.
	74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19.
30	75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20
	76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21.
	77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22.
35	78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23.

•	79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24
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5	81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26.
	82. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27.
10	83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28.
	84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29.
	85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30.
15	86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31.
	87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32.
20	88. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:33.
	89. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:34.
25	90. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:35.
20	91. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:36.
30	92. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:37.

93. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

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NO:38.

	NO:39.
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15	99. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:44.
20	100. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:45.
	101. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:46.
25	102. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:47.
	103. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:48.
30	104. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:49.
35	105. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:50.

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5	107. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:52.
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25	114. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:59.
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30	116. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:61.
	117. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

NO:62.

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118. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:63.

119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.

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      KHAN, Farrah A.
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 Thr Ile Pro Tyr Thr Pro Arg Phe Leu Glu Val Phe Leu Ile Tyr
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 Gly Glu Phe Arg Lys Tyr Asn Asn Asn Gly Asp Glu Ile Thr
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 Pro Thr Asn Thr Leu Glu Glu Leu Met Leu Ala Phe Ser His Trp
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 Thr Tyr Glu Tyr Thr Arg Gly Glu Leu Leu Val Leu Asp Leu Gln
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 Gly Val Gly Glu Asn Leu Thr Asp Pro Ser Val Ile Lys Pro Glu
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 Glu Asp Ala Ile Arg Asn Phe Ile Ala Lys His His Trp Asn Ser
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 Cys Cys Arg Lys Leu Lys Leu Pro Asp Leu Lys Arg Asn Asp Tyr
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Val Val Phe His Asn His Ser Asn Trp Ser Leu Glu Asp Thr Gly
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Ala Leu Leu Ser Ser Gly Gln Lys Asp Tyr Val Thr Val Gln Leu
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Gln Asn Gly Glu Ile Trp Glu Leu Ser Arg Cys Ser Arg Asn Lys
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Arg Glu Asn Thr Ser Ser Leu Gly Tyr Glu Tyr Thr Gly Ser Lys
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Lys Glu Phe Pro Cys Val Asp Gly Tyr Ile Tyr Asp Gln Asn Thr
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Trp Lys Ser Thr Ala Val Thr Gln Trp Asn Leu Val Cys Asp Arg
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Lys Trp Leu Ala Met Leu Ile Gln Pro Leu Phe Met Phe Gly Val
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Leu Leu Gly Ser Val Thr Phe Gly Tyr Phe Ser Asp Arg Leu Gly
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Arg Arg Val Val Leu Trp Ala Thr Ser Ser Ser Met Phe Leu Phe
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Trp Ala Ser Val His Leu His Ser Phe Phe Ala Val Gly Thr Leu
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Leu Val Ala Leu Thr Gly Tyr Leu Val Arg Thr Trp Trp Leu Tyr
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Gln Met Ile Leu Ser Thr Val Thr Val Pro Phe Ile Leu Cys Cys
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Trp Val Leu Pro Glu Thr Pro Phe Trp Leu Leu Ser Glu Gly Arg
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Tyr Glu Glu Ala Gln Lys Ile Val Asp Ile Met Ala Lys Trp Asn
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Arg Ala Ser Ser Cys Lys Leu Ser Glu Leu Leu Ser Leu Asp Leu
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Gln Gly Pro Val Ser Asn Ser Pro Thr Glu Val Gln Lys His Asn
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Leu Ser Tyr Leu Phe Tyr Asn Trp Ser Ile Thr Lys Arg Thr Leu
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Phe Ser Leu Asn Ser Val Asn Leu Gly Gly Asn Glu Tyr Leu Asn
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Cys Ile Ala Met Asp Lys Val Gly Arg Arg Thr Val Leu Ala Tyr
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Pro Gln Lys His Tyr Ile Leu Gly Val Val Thr Ala Met Val Gly
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Lys Phe Ala Ile Gly Ala Ala Phe Gly Leu Ile Tyr Leu Tyr Thr
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Val Asp Leu Ser Ser Ile Trp Ile Phe Ile Pro Gln Leu Phe Val
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Gly Thr Met Ala Leu Leu Ser Gly Val Leu Thr Leu Lys Leu Pro
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Glu Thr Leu Gly Lys Arg Leu Ala Thr Thr Trp Glu Glu Ala Ala
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Lys Leu Glu Ser Glu Asn Glu Ser Lys Ser Ser Lys Leu Leu Leu
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Ser His Gln Arg Ala Lys Glu Ile Leu Thr Arg Gly Gly Pro Asn
Thr Val Thr Pro Pro Pro Thr Thr Pro Glu Trp Val Lys Phe Cys
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Lys Gln Leu Phe Gly Gly Phe Ser Leu Leu Leu Trp Thr Gly Ala
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Ile Leu Cys Phe Val Ala Tyr Ser Ile Gln Ile Tyr Phe Asn Glu
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Val Val Ile Val Thr Gly Cys Phe Ser Tyr Tyr Gln Glu Ala Lys
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Ser Ser Lys Ile Met Glu Ser Phe Lys Asn Met Val Pro Gln Gln
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Ala Leu Val Ile Arg Gly Gly Glu Lys Met Gln Ile Asn Val Gln
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Glu Val Val Leu Gly Asp Leu Val Glu Ile Lys Gly Gly Asp Arg
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Val Pro Ala Asp Leu Arg Leu Ile Ser Ala Gln Gly Cys Lys Val
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Asp Asn Ser Ser Leu Thr Gly Glu Ser Glu Pro Gln Ser Arg Ser
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Pro Asp Phe Thr His Glu Asn Pro Leu Glu Thr Arg Asn Ile Cys
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Phe Phe Ser Thr Asn Cys Val Glu Gly Thr Ala Arg Gly Ile Val
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Ile Ala Thr Gly Asp Ser Thr Val Met Gly Arg Ile Ala Ser Leu
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Thr Ser Gly Leu Ala Val Gly Gln Thr Pro Ile Ala Ala Glu Ile
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Glu His Phe Ile His Leu Ile Thr Val Val Ala Val Phe Leu Gly
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Val Thr Phe Phe Ala Leu Ser Leu Leu Gly Tyr Gly Trp Leu
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                                     310
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Glu Ala Ile Ile Phe Leu Ile Gly Ile Ile Val Ala Asn Val Pro
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Glu Gly Leu Leu Ala Thr Val Thr Val Cys Leu Thr Leu Thr Ala
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Lys Arg Met Ala Arg Lys Asn Cys Leu Val Lys Asn Leu Glu Ala
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Val Glu Thr Leu Gly Ser Thr Ser Thr Ile Cys Ser Asp Lys Thr
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                365
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Gly Thr Leu Thr Gln Asn Arg Met Thr Val Ala His Met Trp Phe
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Asp Met Thr Val Tyr Glu Ala Asp Thr Thr Glu Glu Gln Thr Gly
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	Thr			410					415					Arg 420
	Ala			425					430					Glu 435
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	Ala			455					460					465
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	Gln			500					505					510
	Glu			515					520					525
	Asn			530					535					540
	Gly			545					550					555
	Pro Asn			560					565					570
	Ile			575					580					585
	Arg			590					595	-				600
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	Ile			620					625					630
	Gly			635					640					645
PIO	Ile	Ser	rys	650	Asp	Ala	Ser	Ala	655	Lys	Ala	He	Val	Val 660
	Gly			665					670					675
	Leu			680					685					690
	Gln			695					700					705
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	Lys			725					730					Asp 735
	Ser			740					745					750
	Ser			755					760					765
	Leu			770					775					780
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	Leu			800					805					810
	Val			815					820					825
	Met			830					835					840
	His			845					850		•			Ile 855
Gln	Ala	Leu	Ala	860	Phe	Phe	Thr	Tyr	Phe 865	Val	Ile	Leu	Ala	Glu 870

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Glu Asp Lys Tyr Leu Asn Asp Leu Glu Asp Ser Tyr Gly Gln Gln
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Trp Thr Tyr Glu Gln Arg Lys Val Val Glu Phe Thr Cys Gln Thr
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Ala Phe Phe Val Thr Ile Val Val Val Gln Trp Ala Asp Leu Ile
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Ile Ser Lys Thr Arg Arg Asn Ser Leu Phe Gln Gln Gly Met Arg
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Asn Lys Val Leu Ile Phe Gly Ile Leu Glu Glu Thr Leu Leu Ala
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Ala Phe Leu Ser Tyr Thr Pro Gly Met Asp Val Ala Leu Arg Met
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Tyr Pro Leu Lys Ile Thr Trp Trp Leu Cys Ala Ile Pro Tyr Ser
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Ile Leu Ile Phe Val Tyr Asp Glu Ile Arg Lys Leu Leu Ile Arg
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Thr Met Gln Thr Leu Thr Lys Gln Asp Thr Met Leu Lys Ala Met
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Phe Ser Gly Arg Met Glu Val Leu Thr Asp Ser Glu Gly Trp Ile
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Leu Ile Asp Arg Cys Gly Lys His Phe Gly Thr Ile Leu Asn Tyr
Leu Arg Asp Gly Ala Val Pro Leu Pro Glu Ser Arg Arg Glu Ile
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Glu Glu Leu Leu Ala Glu Ala Lys Tyr Tyr Leu Val Gln Gly Leu
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Val Glu Glu Cys Gln Ala Ala Leu Gln Asn Lys Asp Thr Tyr Glu
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Pro Phe Cys Lys Val Pro Val Ile Thr Ser Ser Lys Glu Glu Gln
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Lys Leu Ile Ala Thr Ser Asn Lys Pro Ala Val Lys Leu Leu Tyr
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Asn Arg Ser Asn Asn Lys Tyr Ser Tyr Thr Ser Asn Ser Asp Asp
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Asn Met Leu Lys Asn Ile Glu Leu Phe Asp Lys Leu Ser Leu Arg
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Phe Asn Gly Arg Val Leu Phe Ile Lys Asp Val Ile Gly Asp Glu
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Ile Cys Cys Trp Ser Phe Tyr Gly Gln Gly Arg Lys Ile Ala Glu
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Val Cys Cys Thr Ser Ile Val Tyr Ala Thr Glu Lys Lys Gln Thr
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Lys Val Glu Phe Pro Glu Ala Arg Ile Tyr Glu Glu Thr Leu Asn
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Ile Leu Leu Tyr Glu Ala Gln Asp Gly Arg Gly Pro Asp Asn Ala
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Leu Leu Glu Ala Thr Gly Gly Ala Ala Gly Arg Ser His His Leu
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Pro Ile Trp Tyr Pro Glu Asn Pro Ser Leu Gly Asp Lys Ile Ala
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Arg Val Ile Val Tyr Phe Val Ala Leu Ile Tyr Met Phe Leu Gly
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Val Ser Ile Ile Ala Asp Arg Phe Met Ala Ser Ile Glu Val Ile
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Thr Ser Gln Glu Arg Glu Val Thr Ile Lys Lys Pro Asn Gly Glu
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Thr Ser Thr Thr Ile Arg Val Trp Asn Glu Thr Val Ser Asn
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Gly Pro Ser Thr Ile Val Gly Ser Ala Ala Phe Asn Met Phe Ile
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Ile Ile Gly Ile Cys Val Tyr Val Ile Pro Asp Gly Glu Thr Arg
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Lys Ile Lys His Leu Arg Val Phe Phe Ile Thr Ala Ala Trp Ser
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Ile Phe Ala Tyr Ile Trp Leu Tyr Met Ile Leu Ala Val Phe Ser
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Pro Gly Val Val Gln Val Trp Glu Gly Leu Leu Thr Leu Phe Phe
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Phe Pro Val Cys Val Leu Leu Ala Trp Val Ala Asp Lys Arg Leu
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Leu Phe Tyr Lys Tyr Met His Lys Lys Tyr Arg Thr Asp Lys His
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Arg Gly Ile Ile Ile Glu Thr Glu Gly Asp His Pro Lys Gly Ile
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Glu Met Asp Gly Lys Met Met Asn Ser His Phe Leu Asp Gly Asn
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Ala Asp Ala Ser Ile Gly Asn Val Thr Gly Ser Asn Ala Val Asn
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Trp Ala Leu Gln Gly Gln Glu Phe His Val Ser Ala Gly Thr Leu
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Ala Phe Ser Val Thr Leu Phe Thr Ile Phe Ala Phe Val Cys Ile
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Ser Val Leu Leu Tyr Arg Arg Arg Pro His Leu Gly Gly Glu Leu
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Gly Gly Pro Arg Gly Cys Lys Leu Ala Thr Thr Trp Leu Phe Val
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Leu Lys Gln Val Phe Gly Lys Gly Leu Ile Lys Ala Ala Met Thr
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Thr Gly Ala Trp Ile Phe Thr Gly Gly Val Ser Thr Gly Val Ile
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Ser His Val Gly Asp Ala Leu Lys Asp His Ser Ser Lys Ser Arg
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Gly Arg Val Cys Ala Ile Gly Ile Ala Pro Trp Gly Ile Val Glu
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Asn Lys Glu Asp Leu Val Gly Lys Asp Val Thr Arg Val Tyr Gln
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Thr Met Ser Asn Pro Leu Ser Lys Leu Ser Val Leu Asn Asn Ser
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His Thr His Phe Ile Leu Ala Asp Asn Gly Thr Leu Gly Lys Tyr
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Leu Gln Lys Ile Asn Thr Arg Ile Gly Gln Gly Val Pro Val Val
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Ala Leu Ile Val Glu Gly Gly Pro Asn Val Ile Ser Ile Val Leu
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Glu Tyr Leu Arg Asp Thr Pro Pro Val Pro Val Val Val Cys Asp
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Gly Ser Gly Arg Ala Ser Asp Ile Leu Ala Phe Gly His Lys Tyr
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Ser Glu Glu Gly Gly Leu Ile Asn Glu Ser Leu Arg Asp Gln Leu
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Leu Val Thr Ile Gln Lys Thr Phe Thr Tyr Thr Arg Thr Gln Ala
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Gln His Leu Phe Ile Ile Leu Met Glu Cys Met Lys Lys Glu Leu Ile Thr Val Phe Arg Met Gly Ser Glu Gly His Gln Asp Ile Asp Leu Ala Ile Leu Thr Ala Leu Leu Lys Gly Ala Asn Ala Ser Ala Pro Asp Gln Leu Ser Leu Ala Leu Ala Trp Asn Arg Val Asp Ile Ala Arg Ser Gln Ile Phe Ile Tyr Gly Gln Gln Trp Pro Val Gly Ser Leu Glu Gln Ala Met Leu Asp Ala Leu Val Leu Asp Arg Val Asp Phe Val Lys Leu Leu Ile Glu Asn Gly Val Ser Met His Arg Phe Leu Thr Ile Ser Arg Leu Glu Glu Leu Tyr Asn Thr Arg His Gly Pro Ser Asn Thr Leu Tyr His Leu Val Arg Asp Val Lys Lys Gly Asn Leu Pro Pro Asp Tyr Arg Ile Ser Leu Ile Asp Ile Gly Leu Val Ile Glu Tyr Leu Met Gly Gly Ala Tyr Arg Cys Asn Tyr Thr Arg Lys Arg Phe Arg Thr Leu Tyr His Asn Leu Phe Gly Pro Lys Arg Asp Asp Ile Pro Leu Arg Arg Gly Arg Lys Thr Thr Lys Lys Arg Glu Glu Glu Val Asp Ile Asp Leu Asp Asp Pro Glu Ile Asn His Phe Pro Phe Pro Phe His Glu Leu Met Val Trp Ala Val Leu Met Lys Arg Gln Lys Met Ala Leu Phe Phe Trp Gln His Gly Glu Glu Ala Met Ala Lys Ala Leu Val Ala Cys Lys Leu Cys Lys Ala Met Ala His Glu Ala Ser Glu Asn Asp Met Val Asp Asp Ile Ser Gln Glu Leu Asn His Asn Ser Arg Asp Phe Gly Gln Leu Ala Val Glu Leu Leu Asp Gln Ser Tyr Lys Gln Asp Glu Gln Leu Ala Met Lys Leu Leu Thr Tyr Glu Leu Lys Asn Trp Ser Asn Ala Thr Cys Leu Gln Leu Ala Val Ala Ala Lys His Arg Asp Phe Ile Ala His Thr Cys Ser Gln Met Leu Leu Thr Asp Met Trp Met Gly Arg Leu Arg Met Arg Lys Asn Ser Gly Leu Lys Val Ile Leu Gly Ile Leu Leu Pro Pro Thr Ile Leu Phe Leu Glu Phe Arg Thr Tyr Asp Asp Phe Ser Tyr Gln Thr Ser Lys Glu Asn Glu Asp Gly Lys Glu Lys Glu Glu Glu Asn Thr Asp Ala Asn Ala Asp Ala Gly Ser Arg Lys Gly Asp Glu Glu Asn Glu His Lys Lys Gln Arg Ser Ile Pro Ile Gly Thr Lys Ile Cys Glu Phe Tyr Asn Ala Pro Ile Val Lys Phe Trp Phe Phe Gln Ile Ser Tyr Leu Gly Tyr Leu Leu Phe Asn Tyr Val Ile Leu Val Arg Met Asp Gly Trp Pro Ser Leu 

Gln Glu Trp Ile Val Ile Ser Tyr Ile Val Ser Leu Ala Leu Glu Lys Ile Arg Glu Val Ala Thr Pro Lys Ala Ser Pro Ser Pro Leu Ala Arg Lys Ser Lys Phe Gly Phe Gln Glu Tyr Trp Asn Ile Thr Asp Leu Val Ala Ile Ser Thr Phe Met Ile Gly Ala Ile Leu Arg Leu Gln Asn Gln Pro Tyr Met Gly Tyr Gly Arg Val Ile Tyr Cys Val Asp Ile Ile Phe Trp Tyr Ile Arg Val Leu Asp Ile Phe Gly Val Asn Lys Tyr Leu Gly Pro Tyr Val Met Met Ile Gly Lys Met Val Ser Ser Gly Ile Leu Trp Val Val Ile Met Leu Val Val Leu Met Ser Phe Gly Val Ala Arg Gln Ala Ile Leu His Pro Glu Glu Lys Pro Ser Trp Lys Leu Ala Arg Asn Ile Phe Tyr Met Pro Tyr Trp Met Ile Tyr Gly Glu Val Phe Ala Asp Gln Ile Asp Arg Lys Ser Phe Phe Leu Ser Ala Pro Cys Gly Glu Asn Leu Tyr Asp Glu Glu Gly Lys Arg Leu Pro Pro Cys Ile Pro Gly Ala Trp Leu Thr Pro Ala Leu Met Ala Cys Tyr Leu Leu Val Ala Asn Ile Leu Leu Val Asn Leu Leu Ile Ala Val Phe Ser Asn Thr Phe Phe Glu Val Lys Ser Ile Ser Asn Gln Val Trp Lys Phe Gln Arg Tyr Gln Leu Ile Met Thr Phe His Asp Arg Pro Val Leu Pro Pro Pro Met Ile Ile Leu Ser His Ile Tyr Ile Ile Ile Met Arg Leu Ser Gly Arg Cys Arg Lys Lys Arg Glu Gly Asp Gln Glu Glu Arg Asp Arg Gly Leu Ser Met Phe Leu Ser Asp Glu Glu Leu Lys Arg Leu His Glu Phe Glu Glu Gln Cys Val Gln Glu His Phe Arg Glu Lys Glu Asp Glu Gln Gln Ser Ser Ser Asp Glu Arg Ile Arg Val Thr Ser Glu Arg Val Glu Asn Met Ser Met Arg Leu Glu Glu Ile Asn Glu Arg Glu Thr Phe Met Lys Thr Ser Leu Gln Thr Val Asp Leu Arg Leu Ala Gln Leu Glu Glu Leu Ser Asn Arg Met Val Asn Ala Leu Glu Asn Leu Ala Gly Ile Asp Arg Ser Asp Leu Ile Gln Ala Arg Ser Arg Ala Ser Ser Glu Cys Glu Ala Thr Tyr Leu Leu Arg Gln Ser Ser Ile Asn Ser Ala Asp Gly Tyr Ser Leu Tyr Arg Tyr His Phe Asn Gly Glu Glu Leu Leu Phe Glu Asp Thr Ser Leu Ser Thr Ser Pro Gly Thr Gly Val Arg Lys Lys Thr Cys Ser Phe Arg Ile Lys Glu Glu Lys Asp Val Lys Thr His Leu Val Pro Glu Cys Gln Asn Ser Leu His Leu Ser Leu Gly Thr Ser Thr Ser Ala Thr Pro Asp

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Gly Ser His Leu Ala Val Asp Asp Leu Lys Asn Ala Glu Glu Ser
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Lys Leu Gly Pro Asp Ile Gly Ile Ser Lys Glu Asp Asp Glu Arg
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                                    1225
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Gln Thr Asp Ser Lys Lys Glu Glu Thr Ile Ser Pro Ser Leu Asn
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Lys Thr Asp Val Ile His Gly Gln Asp Lys Ser Asp Val Gln Asn
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Thr Gln Leu Thr Val Glu Thr Thr Asn Ile Glu Gly Thr Ile Ser
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Tyr Pro Leu Glu Glu Thr Lys Ile Thr Arg Tyr Phe Pro Asp Glu
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Thr Ile Asn Ala Cys Lys Thr Met Lys Ser Arg Ser Phe Val Tyr
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                                                        1305
Ser Arg Gly Arg Lys Leu Val Gly Gly Val Asn Gln Asp Val Glu
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Tyr Ser Ser Ile Thr Asp Gln Gln Leu Thr Thr Glu Trp Gln Cys
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Gln Val Gln Lys Ile Thr Arg Ser His Ser Thr Asp Ile Pro Tyr
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Ile Val Ser Glu Ala Ala Val Gln Ala Glu His Lys Glu Gln Phe
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Ala Asp Met Gln Asp Glu His His Val Ala Glu Ala Ile Pro Arg
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Ile Pro Arg Leu Ser Leu Thr Ile Thr Asp Arg Asn Gly Met Glu
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Asn Leu Leu Ser Val Lys Pro Asp Gln Thr Leu Gly Phe Pro Ser
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Leu Arg Ser Lys Ser Leu His Gly His Pro Arg Asn Val Lys Ser
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Ile Gln Gly Lys Leu Asp Arg Ser Gly His Ala Ser Ser Val Ser
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Lys Lys Glu Lys Ala Ser Thr Glu Thr Glu Cys
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Arg Val Gln Leu Leu Gln Met Gln His Ala Ser Met Pro Met
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Ala Ala Ala Lys Gln Cys Lys Gly Ile Val Asp Cys Ile Val Arg
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                                     55
Ile Pro Lys Asp Gln Gly Val Leu Ser Phe Trp Arg Gly Asn Leu
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                                     70
Ala Asn Val Ile Arg Tyr Ser Pro Thr Gln Ala Leu Asn Phe Ala
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                                     85
Phe Lys Asp Lys Tyr Lys Gln Ile Phe Leu Ala Gly Val Asp Lys
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                                    100
                                                         105
His Thr Gln Phe Cys Arg Tyr Phe Ala Gly Asn Leu Ala Ser Gly
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Gly Thr Ala Val Val Tyr Pro Leu Asp Phe Thr Arg Thr Arg Leu
                125
                                     130
Ala Ala Asp Val Gly Lys Ser Gly Thr Glu Arg Glu Phe Arg Gly
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Leu Gly Asp Cys Leu Val Lys Ile Ser Lys Ser Asp Gly Ile Arg
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                                     160
                                                          165
Gly Leu Tyr Gln Gly Phe Ser Val Ser Val Gln Ala Ile Ile Ile
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                                     175
Tyr Gln Ala Ala Tyr Phe Arg Val Tyr Asp Thr Ala Asn Gly Met
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                                     190
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Phe Pro Asp Pro Lys Asn Thr His Ile Leu Val Ser Trp Met Thr
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Ala Gln Thr Val Thr Ala Val Ala Gly Val Leu Ser
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Leu Ile Met Gly Val Leu Gly Leu Gly Phe Ile Ala Thr Tyr Leu
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Pro Glu Ser Ala Met Ser Ala Tyr Leu Ala Ala Val Ala Leu His
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                                      55
Ile Met Leu Ser Gln Leu Thr Phe Ile Phe Gly Ile Met Ile Ser
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Phe His Ala Gly Pro Ile Ser Phe Phe Tyr Asp Ile Ile Asn Tyr
Cys Val Ala Leu Pro Lys Ala Asn Ser Thr Ser Ile Leu Val Phe
                 95
                                     100
                                                         105
Leu Thr Val Val Val Ala Leu Arg Ile Asn Lys Cys Ile Arg Ile
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                                     115
Ser Phe Asn Gln Tyr Pro Ile Glu Phe Pro Met Glu Leu Phe Leu
                125
                                     130
Ile Ile Leu Gln Ala Phe Ser Leu Ser Leu Val Ser Ser Phe Leu
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                                     145
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Leu Ile Phe Leu Gly Lys Lys Ile Ala Ser Leu His Asn Tyr Ser
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Val Asn Ser Asn Gln Asp Leu Ile Ala Ile Gly Leu Cys Asn Val
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Val Ser Ser Phe Phe Arg Ser Cys Val Phe Thr Gly Ala Ile Ala
                185
                                     190
                                                         195
Arg Thr Ile Ile Gln Asp Lys Ser Gly Gly Ser Thr Thr Val Cys
                200
                                     205
                                                          210
Ile Ser Gly Arg Arg Ala Lys Ile Leu Leu Gly Gln Ile
                                     220
                215
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Pro Asn Thr Asn Ile Tyr Arg Ser Ile Asn Asp Tyr Arg Glu Ile
                230
                                     235
Ile Thr Ile Pro Gly Val Lys Ile Phe Gln Cys Cys Ser Ser Ile
                245
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Thr Phe Val Asn Val Tyr Tyr Leu Lys His Lys Leu Leu Lys Glu
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                260
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Val Asp Met Val Lys Val Pro Leu Lys Glu Glu Glu Ile Phe Ser
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275
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Leu Phe Asn Ser Ser Asp Thr Asn Leu Gln Gly Gly Lys Ile Cys
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Arg Cys Phe Cys Asn Cys Asp Asp Leu Glu Pro Leu Pro Arg Ile
                 305
                                     310
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Leu Tyr Thr Glu Arg Phe Glu Asn Lys Leu Asp Pro Glu Ala Ser
                 320
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                                                          330
Ser Val Asn Leu Ile His Cys Ser His Phe Glu Ser Met Asn Thr
                 335
                                     340
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Ser Gln Thr Ala Ser Glu Asp Gln Val Pro Tyr Thr Val Ser Ser
                350
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Val Ser Gln Lys Asn Gln Gly Gln Gln Tyr Glu Glu Val Glu Glu
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Val Trp Leu Pro Asn Asn Ser Ser Arg Asn Ser Ser Pro Gly Leu
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Pro Asp Val Ala Glu Ser Gln Gly Arg Arg Ser Leu Ile Pro Tyr
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                                     400
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Ser Asp Ala Ser Leu Leu Pro Ser Val His Thr Ile Ile Leu Asp
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                                     415
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Phe Ser Met Val His Tyr Val Asp Ser Arg Gly Leu Val Val Leu
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Ile Arg Leu Val Leu Asp Pro His Leu Leu Lys Tyr Tyr Asn Lys
                 35
                                      40
Gln Lys Thr Tyr Phe Ala His Asn Ala Leu Gln Gln Cys Thr Ile
                 50
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Gly Asp Ile Val Leu Leu Lys Ala Leu Pro Val Pro Arg Thr Lys
                 65
                                      70
His Val Lys His Glu Leu Ala Glu Ile Val Phe Lys Val Gly Lys
                 80
                                      85
                                                           90
Leu Val Asp Pro Val Thr Gly Lys Pro Arg Ala Gly Thr Thr Tyr
                 95
                                     100
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Leu Glu Ser Pro Leu Ser Ser Glu Thr Thr Gln Gly Val Asp Gly
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                                     115
                                                          120
Ala Ser Arg Pro Ser Arg Gly Pro Ala Pro Cys Arg Ala Gly Pro
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                                     130
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Gly Ala Arg Arg Leu Arg Pro Trp Pro Glu Ser Pro Arg Pro Glu
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                                     145
                                                          150
Pro Arg Gly Leu Pro Gly Pro Gly Arg Gly Ser Met Ala Thr Trp
                155
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                                                          165
Arg Arg Asp Gly Arg Leu Thr Gly Gly Gln Arg Leu Leu Cys Ala
                170
                                     175
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Gly Leu Ala Gly Thr Leu Ser Leu Ser Leu Thr Ala Pro Leu Glu
                185
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Leu Ala Thr Val Leu Ala Gln Val Gly Val Val Arg Gly His Ala
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205
Arg Gly Pro Trp Ala Thr Gly His Arg Val Trp Arg Ala Glu Gly
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Leu Arg Ala Leu Trp Lys Gly Asn Ala Val Ala Cys Leu Arg Leu
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Phe Pro Cys Ser Ala Val Gln Leu Ala Ala Tyr Arg Lys Phe Val
                 245
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Val Leu Phe Thr Asp Asp Leu Gly His Ile Ser Gln Trp Ser Ser
                 260
                                     265
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Ile Met Ala Gly Ser Leu Ala Gly Met Val Ser Thr Ile Val Thr
                 275
                                     280
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Tyr Pro Thr Asp Leu Ile Lys Thr Arg Leu Ile Met Gln Asn Ile
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Leu Glu Pro Ser Tyr Arg Gly Leu Leu His Ala Phe Ser Thr Ile
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Tyr Gln Gln Glu Gly Phe Leu Ala Leu Tyr Arg Gly Val Ser Leu
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Thr Val Val Gly Ala Leu Pro Phe Ser Ala Gly Ser Leu Leu Val
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Tyr Met Asn Leu Glu Lys Ile Trp Asn Gly Pro Arg Asp Gln Phe
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Ser Leu Pro Gln Asn Phe Ala Asn Val Cys Leu Ala Ala Val
                 365
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Thr Gln Thr Leu Ser Phe Pro Phe Glu Thr Val Lys Arg Lys Met
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                                     385
Gln Ala Gln Ser Pro Tyr Leu Pro His Ser Gly Gly Val Asp Val
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                                     400
His Phe Ser Gly Ala Val Asp Cys Phe Arg Gln Ile Val Lys Ala
                 410
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                                                          420
Gln Gly Val Leu Gly Leu Trp Asn Gly Leu Thr Ala Asn Leu Leu
                 425
                                     430
Lys Ile Val Pro Tyr Phe Gly Ile Met Phe Ser Thr Phe Glu Phe
                 440
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Cys Lys Arg Ile Cys Leu Tyr Gln Asn Gly Tyr Ile Leu Ser Pro
                 455
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Leu Ser Tyr Lys Leu Thr Pro Gly Val Asp Gln Ser Leu Gln Pro
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Gln Glu Leu Arg Glu Leu Lys Lys Phe Phe Lys Thr Arg Lys Leu
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Ile Val Ala Arg Gly Thr Leu Lys Val Ala Val Pro Gln Asp Phe
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Pro Pro Phe Gly Ser Val Gly Pro Asp Met Gln Pro Arg Gly Leu
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Asp Ile Asp Thr Ala Lys Leu Leu Ala Asp Gln Leu Lys Val Lys
                 65
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Leu Glu Leu Thr Pro Val Asn Ser Thr Asn Arg Ile Pro Phe Leu
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Thr Thr Gly Lys Val Asp Leu Val Ile Ser Ser Leu Gly Lys Asn
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Pro Glu Arg Ala Lys Val Ile Asp Phe Ser Asn Ala Tyr Ala Pro
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Phe Tyr Leu Ala Val Phe Gly Pro Pro Asp Ala Ala Ile Ala Ser
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                                     130
Leu Asp Asp Leu Lys Gly Lys Thr Ile Ser Val Thr Arg Gly Ala
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Ile Glu Asp Ile Glu Leu Thr Ala Val Ala Pro Lys Glu Ala Thr
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Ile Lys Arg Phe Glu Asp Asn Asn Ser Thr Ile Ala Ala Tyr Leu
                170
                                     175
Ala Gly Gln Val Asp Leu Ile Ala Ser Gly Asn Val Val Met Val
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                                                         195
Ala Ile Ser Glu Arg Asn Pro Lys Arg Val Pro Ala Leu Lys Val
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                                     205
                                                         210
Lys Leu Lys Asp Ser Pro Val Tyr Val Gly Val Asn Lys Asn Glu
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Pro Ala Leu Leu Glu Lys Val Asn Gln Ile Leu Val Ala Ala Lys
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Pro Leu Pro Ala Asp Leu
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Ala Arg Arg His Thr Gly Leu Ala Pro Val Leu Gly Val Arg Val
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Arg Val Cys Val Gly Cys Gln Gly Ala Arg Val Pro Ala Trp Pro
Lys Arg Glu Gly Arg Gly Ser Ala Glu Glu Pro Val Glu Arg Lys
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                                      85
Asp Gly Gln Gln Gly Glu Arg Gly Glu Ala Ser Leu Thr Lys
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                                     100
                                                         105
Asp Ala Val Thr Phe Gln Ser Ser Leu Pro Val Gln Ile Thr Arg
                110
                                     115
                                                         120
Ile Pro Ala Cys Trp Arg Met Thr Gln Met Ser Gln Val Gln Glu
                125
                                     130
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Leu Phe His Glu Ala Ala Gln Gln Asp Ala Leu Ala Gln Pro Gln
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Pro Trp Trp Lys Thr Gln Leu Phe Met Trp Glu Pro Val Leu Phe
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Gly Thr Trp Asp Gly Val Phe Thr Ser Cys Met Ile Asn Ile Phe
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Gly Val Val Leu Phe Leu Arg Thr Gly Trp Leu Val Gly Asn Thr
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Gly Val Leu Leu Gly Met Phe Leu Val Ser Phe Val Ile Leu Val
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Ser Ile Gly Ser Gly Gly Val Tyr Ser Met Ile Ser Ser Val Leu
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Gly Gly Gln Thr Gly Gly Thr Ile Gly Leu Leu Tyr Val Phe Gly
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Gln Met Tyr Ile Thr Gly Phe Ala Glu Ser Ile Ser Asp Leu Leu
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Gly Leu Gly Asn Ile Trp Ala Val Arg Gly Ile Ser Val Ala Val
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                                      280
Leu Leu Ala Leu Leu Gly Ile Asn Leu Ala Gly Val Lys Trp Ile
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                                      295
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Ile Arg Leu Gln Leu Leu Leu Phe Leu Leu Ala Val Ser Thr
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Leu Asp Phe Val Val Gly Ser Phe Thr His Leu Asp Pro Glu His
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Gly Phe Ile Gly Tyr Ser Pro Glu Leu Leu Gln Asn Asn Thr Leu
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Pro Asp Tyr Ser Pro Gly Glu Ser Phe Phe Thr Val Phe Gly Val
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Phe Phe Pro Ala Ala Thr Gly Val Met Ala Gly Phe Asn Met Gly
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Gly Asp Leu Arg Glu Pro Ala Ala Ser Ile Pro Leu Gly Ser Leu
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Ala Ala Val Gly Ile Ser Trp Phe Leu Tyr Ile Gly Tyr Arg Ser
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His Gly Arg Leu Gln His Gly Gly Arg Pro Gln Gly Ala Cys Arg
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Gln His Ser Pro Gly Leu Pro Gly Ser Cys Trp His Leu Val Ser
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Ser Pro His Gly Asn Gln Ser Glu Leu Glu Ser Gly Arg Glu Asn
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Pro Arg Gln Lys Arg Asp Cys Pro Phe Cys Ser Glu Ser Ala Pro
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Thr Val Thr Asn Leu Phe His Ile Pro Ile Leu Leu Thr Leu
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Gly Gly Leu Glu Asn Ser Arg Leu Val Gly Ser Asp Thr Ala Leu
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Ser Cys Ala Leu Pro Gly Asn Ala Ala Glu Glu Glu Cys Arg Asp
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Pro Asp His Arg Ser Asn Pro Phe Phe Pro Phe Leu Gly Leu Pro
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Pro Pro Ser Leu Pro His Pro His Phe Glu Leu Gly Asp Phe Phe
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Asn His Arg Val Ser Arg Ile Thr Gly Gly Ser Ile Lys Met Lys
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Ser Phe Ala Ile Arg Arg Ile Gly Ala Ala Ser Gln Leu Cys Arg
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Leu Asp Leu Trp Gly Thr Val Pro Thr Ala Asp Asn Cys Phe Gln Glu Ala His Ser Phe Ala Ser Gly Gln Ala Val Pro Trp Ser Pro Leu Cys Ala Ala Met Ala Ser Cys His Ser Leu Ile Leu Leu Asn Gly Thr Ile Gln Gly Asp Pro Leu Asp Leu Lys Met Phe Glu Gly Thr Ala Trp Lys Met Glu Asp Cys Ile Val Asp Ser Cys Lys Phe Gly Thr Ser Val Ser Asn Ile Ile Lys Pro Gly Pro Lys Ala Ser Lys Ser Pro Val Glu Ala Ile Ile Thr Leu Cys Gln Phe Pro Phe Ser Ser Ser Leu Gln Arg Met Ser Val Ile Ala Gln Leu Ala Gly Glu Asn His Phe His Val Tyr Met Lys Gly Ala Pro Glu Met Val Ala Arg Phe Cys Arg Ser Glu Thr Val Pro Lys Asn Phe Pro Gln Glu Leu Arg Ser Tyr Thr Val Gln Gly Phe Arg Val Ile Ala Leu Ala His Lys Thr Leu Lys Met Gly Asn Leu Ser Glu Val Glu His Leu Ala Arg Glu Lys Val Glu Ser Glu Leu Thr Phe Leu Gly Leu Leu Ile Met Glu Asn Arg Leu Lys Lys Glu Thr Lys Leu Val Leu Lys Glu Leu Ser Glu Ala Arg Ile Arg Thr Val Met Ile Thr Gly Asp Asn Leu Gln Thr Ala Ile Thr Val Ala Lys Asn Ser Glu Met Ile Pro Pro Gly Ser Gln Val Ile Ile Val Glu Ala Asp Glu Pro Glu Glu Phe Val Pro Ala Ser Val Thr Trp Gln Leu Val Glu Asn Gln Glu Thr Gly Pro Gly Lys Lys Glu Ile Tyr Met His Thr Gly Asn Ser Ser Thr Pro Arg Gly Glu Gly Gly Ser Cys Tyr His Phe Ala Met Ser Gly Lys Ser Tyr Gln Val Ile Phe Gln His Phe Asn Ser Leu Leu Pro Lys Ile Leu Val Asn Gly Thr Val Phe Ala Arg Met Ser Pro Gly Gln Lys Ser Ser Leu Ile Glu Glu Phe Gln Lys Leu Asn Ala Cys Thr Val Gln Asn Glu Ser Ile Ser Glu Leu Thr Met Ser Pro Thr Ala Pro Glu Lys Met Glu Ser Asn Ser Thr Phe Thr Ser Phe Glu Asn Thr Thr Val Trp Phe Leu Gly Thr Ile Asn Cys Ile Thr Val Ala Leu Val Phe Ser Lys Gly Lys Pro Phe Arg Gln Pro Thr Tyr Thr Asn Tyr Ile Phe Val Leu Val Leu Ile Ile Gln Leu Gly Val Cys Leu Phe Ile Leu Phe Ala Asp Ile Pro Glu Leu Tyr Arg Arg Leu Asp Leu Cys Thr Pro Val Leu Trp Arg Ala Ser Ile Val Ile Met Leu Ser Leu Asn Phe Ile Val Ser Leu 

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Val Ala Glu Glu Ala Val Ile Glu Asn Arg Ala Leu Trp Met Met
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Ile Lys Arg Cys Phe Gly Tyr Gln Ser Lys Ser Gln Tyr Arg Ile
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Trp Gln Arg Asp Leu Ala Asn Asp Pro Ser Trp Pro Pro Leu Asn
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Gln Thr Ser His Ser Asp Met Pro Glu Cys Gly Arg Gly Val Ser
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Ser Glu Pro Phe Leu Ile Pro Tyr Leu Ser Gly Pro Asp Lys Asn
                 35
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Leu Thr Ser Ala Glu Ile Thr Asn Glu Ile Phe Pro Val Trp Thr
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                                      55
Tyr Ser Tyr Leu Val Leu Leu Pro Val Phe Val Leu Thr Asp
                                                          75
                                      70
                 65
Tyr Val Arg Tyr Lys Pro Val Ile Ile Leu Gln Gly Ile Ser Phe
                                                          90
                 80
                                      85
Ile Ile Thr Trp Leu Leu Leu Phe Gly Gln Gly Val Lys Thr
                 95
                                     100
Met Gln Val Val Glu Phe Phe Tyr Gly Met Val Thr Ala Ala Glu
                110
                                     115
Val Ala Tyr Tyr Ala Tyr Ile Tyr Ser Val Val Ser Pro Glu His
                125
                                     130
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Tyr Gln Arg Val Ser Gly Tyr Cys Arg Ser Val Thr Leu Ala Ala
                140
                                     145
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Tyr Thr Ala Gly Ser Val Leu Ala Gln Leu Leu Val Ser Leu Ala
                155
                                     160
Asn Met Ser Tyr Phe Tyr Leu Asn Val Ile Ser Leu Ala Ser Val
                170
                                                         180
                                     175
Ser Val Ala Phe Leu Phe Ser Leu Phe Leu Pro Met Pro Lys Lys
                185
                                     190
                                                         195
Ser Met Phe Phe His Ala Lys Pro Ser Arg Glu Ile Lys Lys Ser
                                     205
                200
Ser Ser Val Asn Pro Val Leu Glu Glu Thr His Glu Gly Glu Ala
                215
                                     220
                                                         225
Pro Gly Cys Glu Glu Gln Lys Pro Thr Ser Glu Ile Leu Ser Thr
                230
                                     235
                                                         240
Ser Gly Lys Leu Asn Lys Gly Gln Leu Asn Ser Leu Lys Pro Ser
                                     250
                                                         255
                245
Asn Val Thr Val Asp Val Phe Val Gln Trp Phe Gln Asp Leu Lys
                260
                                     265
                                                         270
Glu Cys Tyr Ser Ser Lys Arg Leu Phe Tyr Trp Ser Leu Trp Trp
                275
                                     280
Ala Phe Ala Thr Ala Gly Phe Asn Gln Val Leu Asn Tyr Val Gln
                                     295
                290
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Ile Leu Trp Asp Tyr Lys Ala Pro Ser Gln Asp Ser Ser Ile Tyr
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320
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Ala Phe Ala Val Gly Tyr Val Lys Val Asn Trp Asp Leu Leu Gly
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                                                          345
Glu Leu Ala Leu Val Val Phe Ser Val Val Asn Ala Gly Ser Leu
                 350
                                      355
                                                          360
Phe Leu Met His Tyr Thr Ala Asn Ile Trp Ala Cys Tyr Ala Gly
                 365
                                      370
Tyr Leu Ile Phe Lys Ser Ser Tyr Met Leu Leu Ile Thr Ile Ala
                 380
                                      385
Val Phe Gln Ile Ala Val Asn Leu Asn Val Glu Arg Tyr Ala Leu
                 395
                                      400
Val Phe Gly Ile Asn Thr Phe Ile Ala Leu Val Ile Gln Thr Ile
                 410
                                      415
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Met Thr Val Ile Val Val Asp Gln Arg Gly Leu Asn Leu Pro Val
                 425
                                      430
                                                          435
Ser Ile Gln Phe Leu Val Tyr Gly Ser Tyr Phe Ala Val Ile Ala
                 440
                                      445
                                                          450
Gly Ile Phe Leu Met Arg Ser Met Tyr Ile Thr Tyr Ser Thr Lys
                 455
                                     460
                                                          465
Ser Gln Lys Asp Val Gln Ser Pro Ala Pro Ser Glu Asn Pro Asp
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Val Ser His Pro Glu Glu Glu Ser Asn Ile Ile Met Ser Thr Lys
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Leu
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Gly Pro Ser Ser Gly Gly Gly Phe Val Asp Trp Thr Leu Arg Leu
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Asn Thr Ile Gln Ser Asp Lys Phe Leu Asn Leu Leu Ser Met
Val Pro Val Ile Tyr Gln Lys Asn Gln Glu Asp Arg His Lys Lys
                  50
                                      55
Pro Asn Gly Ile Trp Gln Asp Gly Leu Ser Thr Ala Val Gln Thr
                  65
                                      70
Phe Ser Asn Arg Ser Glu Gln His Met Glu Tyr His Ser Phe Ser
                 80
                                      85
Glu Gln Ser Phe His Ala Asn Asn Gly His Ala Ser Ser Ser Cys
                  95
                                     100
                                                          105
Ser Gln Lys Tyr Asp Asp Tyr Ala Asn Tyr Asn Tyr Cys Asp Gly
                 110
                                     115
                                                          120
Arg Glu Thr Ser Glu Thr Thr Ala Met Leu Gln Asp Glu Asp Ile
                 125
                                     130
                                                          135
Ser Ser Asp Gly Asp Glu Asp Ala Ile Val Glu Val Thr Pro Lys
                 140
                                     145
Leu Pro Lys Glu Ser Ser Gly Ile Met Ala Leu Gln Ile Leu Val
                155
                                     160
Pro Phe Leu Leu Ala Gly Phe Gly Thr Val Ser Ala Gly Met Val
                170
                                     175
                                                          180
Leu Asp Ile Val Gln His Trp Glu Val Phe Arg Lys Val Thr Glu
                 185
                                     190
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Asn Gly Ala Val Glu Ala Ile Ala Thr Phe Gly Gly Ala Val Ala

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Val Phe Ile Leu Val Pro Ala Leu Leu Gly Leu Lys Gly Asn Leu
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Glu Met Thr Leu Ala Ser Arg Leu Ser Thr Ala Val Asn Ile Gly
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                                     220
Lys Met Asp Ser Pro Ile Glu Lys Trp Asn Leu Ile Ile Gly Asn
Leu Ala Leu Lys Gln Val Gln Ala Thr Val Val Gly Phe Leu Ala
                245
                                                         255
                                     250
Ala Val Ala Ala Ile Ile Leu Gly Trp Ile Pro Glu Gly Lys Tyr
                260
                                     265
                                                          270
Tyr Leu Asp His Ser Ile Leu Leu Cys Ser Ser Ser Val Ala Thr
                                     280
                275
                                                         285
Ala Phe Ile Ala Ser Leu Leu Gln Gly Ile Ile Met Val Gly Val
                290
                                     295
                                                         300
Ile Val Gly Ser Lys Lys Thr Gly Ile Asn Pro Asp Asn Val Ala
                305
                                     310
Thr Pro Ile Ala Ala Ser Phe Gly Asp Leu Ile Thr Leu Ala Ile
                320
                                                         330
                                     325
Leu Ala Trp Ile Ser Gln Gly Leu Tyr Ser Cys Leu Glu Thr Tyr
                335
                                     340
                                                         345
Tyr Tyr Ile Ser Pro Leu Val Gly Val Phe Phe Leu Ala Leu Thr
                350
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Pro Ile Trp Ile Ile Ile Ala Ala Lys His Pro Ala Thr Arg Thr
                365
                                     370
                                                          375
Val Leu His Ser Gly Trp Glu Pro Val Ile Thr Ala Met Val Ile
                380
                                     385
                                                         390
Ser Ser Ile Gly Gly Leu Ile Leu Asp Thr Thr Val Ser Asp Pro
                395
                                     400
Asn Leu Val Gly Ile Val Val Tyr Thr Pro Val Ile Asn Gly Ile
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                410
                                     415
Gly Gly Asn Leu Val Ala Ile Gln Ala Ser Arg Ile Ser Thr Tyr
                425
                                                          435
                                     430
Leu His Leu His Ser Ile Pro Gly Glu Leu Pro Asp Glu Pro Lys
                440
                                     445
                                                          450
Gly Cys Tyr Tyr Pro Phe Arg Thr Phe Phe Gly Pro Gly Val Asn
                455
                                     460
                                                          465
Asn Lys Ser Ala Gln Val Leu Leu Leu Val Ile Pro Gly His
                470
                                     475
                                                          480
Leu Ile Phe Leu Tyr Thr Ile His Leu Met Lys Ser Gly His Thr
                485
                                     490
                                                          495
Ser Leu Thr Ile Ile Phe Ile Val Val Tyr Leu Phe Gly Ala Val
                500
                                     505
                                                          510
Leu Gln Val Phe Thr Leu Leu Trp Ile Ala Asp Trp Met Val His
                                                          525
                515
                                     520
His Phe Trp Arg Lys Gly Lys Asp Pro Asp Ser Phe Ser Ile Pro
                530
                                     535
                                                          540
Tyr Leu Thr Ala Leu Gly Asp Leu Leu Gly Thr Ala Leu Leu Ala
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                545
                                     550
Leu Ser Phe His Phe Leu Trp Leu Ile Gly Asp Arg Asp Gly Asp
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Val Gly Asp
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<400> 19

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470
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Glu Thr Leu Leu Pro Lys Ile Gln Asn Lys Asn Leu Thr Gln Met
                485
                                     490
Ile Ile Val Thr Leu Ala Glu Lys Thr Pro Tyr Leu Glu Ser Lys
                500
                                     505
                                                          510
Arg Leu Val Glu Asp Leu Asn Arg Ala Asn Ile Gly His Asn Trp
                515
                                                          525
                                     520
Trp Val Val Asn Gln Ser Leu Val Thr Leu Asn Gln Arg Asp Asp
                530
                                     535
                                                          540
Leu Phe Ser Asn Lys Lys Glu Asp Glu Ser Phe Trp Ile Asn Lys
                545
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Ile Lys Asn Glu Ser Phe Asp Asn Tyr Phe Val Ile Pro Tyr Gly.
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Gly Leu Ser
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Val Met Asn Ala Ile Ser Ser Leu Pro Leu Leu Ile Leu Ala Pro
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Phe Leu Glu Tyr Phe Ser Thr Cys Leu Phe Pro Ser Lys Arg Val
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                 35
                                      40
Gly Ser Phe Leu Ser Thr Cys Ile Ile Ala Gly Asn Leu Phe Ala
Ala Leu Ser Val Met Ile Ala Gly Phe Phe Glu Ile His Arg Lys
                 65
                                      70
His Phe Pro Ala Val Glu Gln Pro Leu Ser Gly Lys Val Leu Thr
                                      85
                 80
Val Ser Ser Met Pro Cys Phe Tyr Leu Ile Leu Gln Tyr Val Leu
                 95
                                     100
Leu Gly Val Ala Glu Thr Leu Val Asn Pro Ala Leu Ser Val Ile
                110
                                     115
                                                          120
Ser Tyr Arg Phe Val Pro Ser Asn Val Arg Gly Thr Ser Met Asn
                125
                                     130
                                                         135
Phe Leu Thr Leu Phe Asn Gly Phe Gly Cys Phe Thr Gly Ala Leu
Leu Val Lys Leu Val Tyr Leu Ile Ser Glu Gly Lys Asn Arg Gln
                155
                                     160
Trp Phe Pro Asn Thr Leu Asn Lys Gly Asn Leu Glu Gly Phe Phe
                170
                                     175
                                                         180
Phe Phe Leu Ala Ser Leu Thr Leu Leu Asn Val Leu Gly Phe Cys
                185
                                     190
                                                          195
Ser Val Ser Gln Arg Tyr Cys Asn Leu Asn His Phe Asn Ala Gln
                200
                                     205
                                                          210
Asn Ile Arg Gly Ser Asn Leu Glu Glu Thr Leu Leu Leu His Glu
                215
                                     220
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Lys Ser Leu Lys Phe Tyr Gly Ser Ile Gln Glu Phe Ser Ser Ser
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Ile Asp Leu Trp Glu Thr Ala Leu
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Glu Ser Pro Gln Cys Ser Ser Ser Ser Ala Cys Cys Thr Glu Thr
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Glu Lys Gln His Gly Asp Ala Gly Leu Leu Asn Gly Lys Ala Glu
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                                      40
Ser Leu Pro Gly Gln Pro Leu Ala Cys Asn Leu Cys Tyr Glu Ala
                                      55
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Glu Ser Pro Asp Glu Ala Ala Leu Val Tyr Ala Ala Arg Ala Tyr
                  65
                                                           75
Gln Cys Thr Leu Arg Ser Arg Thr Pro Glu Gln Val Met Val Asp
                 80
                                      85
Phe Ala Ala Leu Gly Pro Leu Thr Phe Gln Leu Leu His Ile Leu
                 95
                                     100
Pro Phe Asp Ser Val Arg Lys Arg Met Ser Val Val Arg His
                110
                                     115
Pro Leu Ser Asn Gln Val Val Val Tyr Thr Lys Gly Ala Asp Ser
                125
                                     130
                                                         135
Val Ile Met Glu Leu Leu Ser Val Ala Ser Pro Asp Gly Ala Ser
                140
                                     145
                                                         150
Leu Glu Lys Gln Gln Met Ile Val Arg Glu Lys Thr Gln Lys His
                155
                                     160
Leu Asp Asp Tyr Ala Lys Gln Gly Leu Arg Thr Leu Cys Ile Ala
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Lys Lys Val Met Ser Asp Thr Glu Tyr Ala Glu Trp Leu Arg Asn
                185
                                     190
                                                         195
His Phe Leu Ala Glu Thr Ser Ile Asp Asn Arg Glu Glu Leu Leu
                                     205
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Leu Glu Ser Ala Met Arg Leu Glu Asn Lys Leu Thr Leu Leu Gly
                215
                                                         225
Ala Thr Gly Ile Glu Asp Arg Leu Gln Glu Gly Val Pro Glu Ser
                230
                                     235
Ile Glu Ala Leu His Lys Ala Gly Ile Lys Ile Trp Met Leu Thr
                245
                                     250
Gly Asp Lys Gln Glu Thr Ala Val Asn Ile Ala Tyr Ala Cys Lys
                260
                                     265
Leu Leu Glu Pro Asp Asp Lys Leu Phe Ile Leu Asn Thr Gln Ser
                275
                                     280
                                                         285
Lys Asp Ala Cys Gly Met Leu Met Ser Thr Ile Leu Lys Glu Leu
                290
                                     295
                                                         300
Gln Lys Lys Thr Gln Ala Leu Pro Glu Gln Val Ser Leu Ser Glu
                305
                                     310
Asp Leu Leu Gln Pro Pro Val Pro Arg Asp Ser Gly Leu Arg Ala
                320
                                     325
                                                         330
Gly Leu Ile Ile Thr Gly Lys Thr Leu Glu Phe Ala Leu Gln Glu
                335
                                     340
Ser Leu Gln Lys Gln Phe Leu Glu Leu Thr Ser Trp Gly Gln Ala
                                     355
Val Val Cys Cys Arg Ala Thr Pro Leu Gln Lys Ser Glu Val Val
                365
                                     370
                                                         375
Lys Leu Val Arg Ser His Leu Gln Val Met Thr Leu Ala Ile Gly
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Asp Gly Ala Asn Asp Val Ser Met Ile Gln Val Ala Asp Ile Gly
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Ile Gly Val Ser Gly Gln Glu Gly Met Gln Ala Val Met Ala Ser
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<212> PRT

<213> Homo sapiens

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Val His Gly His Trp Cys Tyr Thr Arg Leu Ser Asn Met Ile Leu
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                                     445
                                                          450
Tyr Phe Phe Tyr Lys Asn Val Ala Tyr Val Asn Leu Leu Phe Trp
                 455
                                     460
Tyr Gln Phe Phe Cys Gly Phe Ser Gly Thr Ser Met Thr Asp Tyr
                 470
                                     475
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Trp Val Leu Ile Phe Phe Asn Leu Leu Phe Thr Ser Ala Pro Pro
                 485
                                     490
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Val Ile Tyr Gly Val Leu Glu Lys Asp Val Ser Ala Glu Thr Leu
                 500
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Met Gln Leu Pro Glu Leu Tyr Lys Ser Gly Gln Lys Ser Glu Ala
                 515
                                     520
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Tyr Leu Pro His Thr Phe Trp Ile Thr Leu Leu Asp Ala Phe Tyr
                530
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Gln Ser Leu Val Cys Phe Phe Val Pro Tyr Phe Thr Tyr Gln Gly
                545
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Ser Asp Thr Asp Ile Phe Ala Phe Gly Asn Pro Leu Asn Thr Ala
                 560
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Ala Leu Phe Ile Val Leu Leu His Leu Val Ile Glu Ser Lys Ser
                575
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Leu Thr Trp Ile His Leu Leu Val Ile Ile Gly Ser Ile Leu Ser
                590
                                     595
Tyr Phe Leu Phe Ala Ile Val Phe Gly Ala Met Cys Val Thr Cys
                605
                                     610
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Asn Pro Pro Ser Asn Pro Tyr Trp Ile Met Gln Glu His Met Leu
                620
                                     625
Asp Pro Val Phe Tyr Leu Val Cys Ile Leu Thr Thr Ser Ile Ala
                635
                                     640
                                                          645
Leu Leu Pro Arg Phe Val Tyr Arg Val Leu Gln Gly Ser Leu Phe
                                     655
                                                          660
Pro Ser Pro Ile Leu Arg Ala Lys His Phe Asp Arg Leu Thr Pro
                665
                                     670
Glu Glu Arg Thr Lys Ala Leu Lys Lys Trp Arg Gly Ala Gly Lys
                680
                                     685
                                                          690
Met Asn Gln Val Thr Ser Lys Tyr Ala Asn Gln Ser Ala Gly Lys
                695
                                     700
                                                          705
Ser Gly Arg Arg Pro Met Pro Gly Pro Ser Ala Val Phe Ala Met
                710
                                     715
Lys Ser Ala Ser Ser Cys Ala Ile Glu Gln Gly Asn Leu Ser Leu
                725
                                     730
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Cys Glu Thr Ala Leu Asp Gln Gly Tyr Ser Glu Thr Lys Ala Phe
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Glu Met Ala Gly Pro Ser Lys Gly Lys Glu Ser
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Met Ala Ser Ser Gly Leu Glu Leu Leu Trp Val Ser Leu Pro Gln
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Leu Gly Lys Gly Ala Ala Gln Thr Leu Ser Ile Ser Phe Leu Ser
Ile Gly Phe Ser Thr Val Gly Gly Val Leu Tyr Gly Val Leu Arg
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35
Thr Leu Asn Asn Lys Ala Ile Asn Gly Val Leu Arg Val Tyr Leu
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Glu Leu Phe Arg Ala Ile Pro Val Leu Val Trp Leu Tyr Leu Leu
                  65
                                       70
Phe Phe Gly Val Pro Ile Phe Phe Gly Leu Ser Ile Pro Ser Phe
                  80
                                       85
Trp Cys Ala Val Leu Val Leu Ser Leu Trp Gly Ala Ser Glu Val
                  95
                                      100
Gly Glu Val Val Arg Gly Ala Leu His Ser Leu Pro Arg Gly Gln
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                                     115
                                                          120
Arg Glu Ala Gly Leu Ser Ile Gly Leu Ser Asp Leu Gln Leu Tyr
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Gly Tyr Val Leu Leu Pro Gln Ala Leu Arg Arg Met Thr Pro Pro
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                                     145
Thr Ile Asn Val Tyr Thr Arg Ile Ile Lys Thr Ser Ser Leu Ala
                155
                                      160
Val Leu Ile Gly Val Val Asp Val Ile Lys Val Gly Gln Gln Ile
                170
                                     175
Ile Glu Arg Thr Tyr Glu Ser Val Leu Ile Tyr Gly Ala Leu Phe
                185
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Leu Phe Phe Phe Ile Cys Tyr Pro Leu Ser Ala Ala Ser Lys
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Leu Leu Glu Arg Arg Trp Ala Gln Ala
                215
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Met Glu Gly Gln Thr Pro Gly Ser Arg Gly Leu Pro Glu Lys Pro
His Pro Ala Thr Ala Ala Ala Thr Leu Ser Ser Met Gly Ala Val
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Phe Ile Leu Met Lys Ser Ala Leu Gly Ala Gly Leu Leu Asn Phe
                 35
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Pro Trp Ala Phe Ser Lys Ala Gly Gly Val Val Pro Ala Phe Leu
                 50
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Val Glu Leu Val Ser Leu Val Phe Leu Ile Ser Gly Leu Val Ile
                 65
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Leu Gly Tyr Ala Ala Ala Val Ser Gly Gln Ala Thr Tyr Gln Gly
                 80
                                      85
Val Val Arg Gly Leu Cys Gly Pro Ala Ile Gly Lys Leu Cys Glu
                 95
                                     100
Ala Cys Phe Leu Leu Asn Leu Leu Met Ile Ser Val Ala Phe Leu
                110
                                     115
Arg Val Ile Gly Asp Gln Leu Glu Lys Leu Cys Asp Ser Leu Leu
                125
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Ser Gly Thr Pro Pro Ala Pro Gln Pro Trp Tyr Ala Asp Gln Arg
                140
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Phe Thr Leu Pro Leu Leu Ser Val Leu Val Ile Leu Pro Leu Ser
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                                     160
                                                          165
Ala Pro Arg Glu Ile Ala Phe Gln Lys Tyr Thr Ser Pro Ser His
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Gly His Cys Val Ser Ile Leu Gly Thr Leu Ala Ala Cys Tyr Leu
                185
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Ala Leu Val Ile Thr Val Gln Tyr Tyr Leu Trp Pro Gln Gly Leu
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Val Arg Glu Ser His Pro Ser Leu Ser Pro Ala Ser Trp Thr Ser
                 215
                                      220
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 Val Phe Ser Val Phe Pro Thr Ile Cys Phe Gly Phe Gln Cys His
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                                                          240
 Glu Ala Ala Val Ser Ile Tyr Cys Ser Met Arg Lys Arg Ser Leu
                 245
                                     250
                                                          255
 Ser His Trp Ala Leu Val Ser Val Leu Ser Leu Leu Ala Cys Cys
                 260
                                      265
                                                          270
 Leu Ile Tyr Ser Leu Thr Gly Val Tyr Gly Phe Leu Thr Phe Gly
                 275
                                     280
                                                          285
 Thr Glu Val Ser Ala Asp Val Leu Met Ser Tyr Pro Gly Asn Asp
                 290
                                     295
                                                          300
Met Val Ile Ile Val Ala Arg Val Leu Phe Ala Val Ser Ile Val
                 305
                                     310
                                                          315
 Thr Val Tyr Pro Ile Val Leu Phe Leu Gly Arg Ser Val Met Gln
                 320
                                     325
                                                          330
Asp Phe Trp Arg Arg Ser Cys Leu Gly Gly Trp Gly Pro Ser Ala
                 335
                                     340
Leu Ala Asp Pro Ser Gly Leu Trp Val Arg Met Pro Leu Thr Ile
                 350
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                                                          360
Leu Trp Val Thr Val Thr Leu Ala Met Ala Leu Phe Met Pro Asp
                 365
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                                                          375
Leu Ser Glu Ile Val Ser Ile Ile Gly Gly Ile Ser Ser Phe Phe
                 380
                                     385
                                                          390
· Ile Phe Ile Phe Pro Gly Pro Gly Gly Lys His Arg Phe Cys Glu
                 395
                                     400
                                                          405
Gly Trp Phe Leu Arg Lys Pro Ser Gln Thr Ala Gln Ala Leu Ser
                 410
                                     415
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Glu Lys Gly Lys Pro Ala Arg Phe Val Pro His Leu Cys Asn Gly
                 425
                                     430
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Cys Arg Ala Tyr Arg Thr Lys Ser Gln Val Arg Asn Leu Arg Ala
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Thr Gly Arg Glu Ser Leu Ser Thr Gln Tyr Glu Gly Ile
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Met Glu Phe Val Arg Ala Leu Trp Leu Gly Leu Ala Leu Ala Leu
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Gly Pro Gly Ser Ala Gly Gly His Pro Gln Pro Cys Gly Val Leu
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Ala Arg Leu Gly Gly Ser Val Arg Leu Gly Ala Leu Leu Pro Arg
                  35
                                      40
                                                           45
Ala Pro Leu Ala Arg Ala Arg Ala Arg Ala Ala Leu Ala Arg Ala
                  50
                                      55
                                                           60
Ala Leu Ala Pro Arg Leu Pro His Asn Leu Ser Leu Glu Leu Val
                  65
Val Ala Ala Pro Pro Ala Arg Asp Pro Ala Ser Leu Thr Arg Gly
                  80
                                      85
Leu Cys Gln Ala Leu Val Pro Pro Gly Val Ala Ala Leu Leu Ala
                  95
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                                                          105
Phe Pro Glu Ala Arg Pro Glu Leu Leu Gln Leu His Phe Leu Ala
                 110
                                     115
Ala Ala Thr Glu Thr Pro Val Leu Ser Leu Leu Arg Arg Glu Ala
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Arg Ala Pro Leu Gly Ala Pro Asn Pro Phe His Leu Gln Leu His Trp Ala Ser Pro Leu Glu Thr Leu Leu Asp Val Leu Val Ala Val Leu Gln Ala His Ala Trp Glu Asp Val Gly Leu Ala Leu Cys Arg Thr Gln Asp Pro Gly Gly Leu Val Ala Leu Trp Thr Ser Arg Ala Gly Arg Pro Pro Gln Leu Val Leu Asp Leu Ser Arg Arg Asp Thr Gly Asp Ala Gly Leu Arg Ala Arg Leu Ala Pro Met Ala Ala Pro Val Gly Glu Ala Pro Val Pro Ala Ala Val Leu Leu Gly Cys Asp Ile Ala Arg Ala Arg Val Leu Glu Ala Val Pro Pro Gly Pro His Trp Leu Leu Gly Thr Pro Leu Pro Pro Lys Ala Leu Pro Thr Ala Gly Leu Pro Pro Gly Leu Leu Ala Leu Gly Glu Val Ala Arg Pro Pro Leu Glu Ala Ala Ile His Asp Ile Val Gln Leu Val Ala Arg Ala Leu Gly Ser Ala Ala Gln Val Gln Pro Lys Arg Ala Leu Leu Pro Ala Pro Val Asn Cys Gly Asp Leu Gln Pro Ala Gly Pro Glu Ser Pro Gly Arg Phe Leu Ala Arg Phe Leu Ala Asn Thr Ser Phe Gln Gly Arg Thr Gly Pro Val Trp Val Thr Gly Ser Ser Gln Val His Met Ser Arg His Phe Lys Val Trp Ser Leu Arg Arg Asp Pro Arg Gly Ala Pro Ala Trp Ala Thr Val Gly Ser Trp Arg Asp Gly Gln Leu Asp Leu Glu Pro Gly Gly Ala Ser Ala Arg Pro Pro Pro Pro Gln Gly Ala Gln Val Trp Pro Lys Leu Arg Val Val Thr Leu Leu Glu His Pro Phe Val Phe Ala Arg Asp Pro Asp Glu Asp Gly Gln Cys Pro Ala Gly Gln Leu Cys Leu Asp Pro Gly Thr Asn Asp Ser Ala Thr Leu Asp Ala Leu Phe Ala Ala Leu Ala Asn Gly Ser Ala Pro Arg Ala Leu Arg Lys Cys Cys Tyr Gly Tyr Cys Ile Asp Leu Leu Glu Arg Leu Ala Glu Asp Thr Pro Phe Asp Phe Glu Leu Tyr Leu Val Gly Asp Gly Lys Tyr Gly Ala Leu Arg Asp Gly Arg Trp Thr Gly Leu Val Gly Asp Leu Leu Ala Gly Arg Ala His Met Ala Val Thr Ser Phe Ser Ile Asn Ser Ala Arg Ser Gln Val Val Asp Phe Thr Ser Pro Phe Phe Ser Thr Ser Leu Gly Ile Met Val Arg Ala Arg Asp Thr Ala Ser Pro Ile Gly Ala Phe Met Trp Pro Leu His Trp Ser Thr Trp Leu Gly Val Phe Ala Ala Leu His Leu Thr Ala Leu Phe Leu Thr Val Tyr Glu Trp Arg Ser Pro 

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Tyr Gly Leu Thr Pro Arg Gly Arg Asn Arg Ser Thr Val Phe Ser.
                605
                                     610
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Tyr Ser Ser Ala Leu Asn Leu Cys Tyr Ala Ile Leu Phe Arg Arg
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                                     625
Thr Val Ser Ser Lys Thr Pro Lys Cys Pro Thr Gly Arg Leu Leu
                635
                                     640
                                                          645
Met Asn Leu Trp Ala Ile Phe Cys Leu Leu Val Leu Ser Ser Tyr
                650
                                     655
                                                          660
Thr Ala Asn Leu Ala Ala Val Met Val Gly Asp Lys Thr Phe Glu
                665
                                     670
                                                          675
Glu Leu Ser Gly Ile His Asp Pro Lys Leu His His Pro Ala Gln
                680
                                     685
                                                          690
Gly Phe Arg Phe Gly Thr Val Trp Glu Ser Ser Ala Glu Ala Tyr
                695
                                     700
                                                          705
Ile Lys Lys Ser Phe Pro Asp Met His Ala His Met Arg Arg His
                710
                                     715
                                                          720
Ser Ala Pro Thr Thr Pro Arg Gly Val Ala Met Leu Thr Ser Asp
                725
                                     730
Pro Pro Lys Leu Asn Ala Phe Ile Met Asp Lys Ser Leu Leu Asp
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                                     745
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Tyr Glu Val Ser Ile Asp Ala Asp Cys Lys Leu Leu Thr Val Gly
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                                     760
                                                          765
Lys Pro Phe Ala Ile Glu Gly Tyr Gly Ile Gly Leu Pro Gln Asn
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                                     775
                                                          780
Ser Pro Leu Thr Ser Asn Leu Ser Glu Phe Ile Ser Arg Tyr Lys
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                                     790
                                                          795
Ser Ser Gly Phe Ile Asp Leu Leu His Asp Lys Trp Tyr Lys Met
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                                     805
                                                          810
Val Pro Cys Gly Lys Arg Val Phe Ala Val Thr Glu Thr Leu Gln
                815
                                     820
                                                          825
Met Ser Ile Tyr His Phe Ala Gly Leu Phe Val Leu Leu Cys Leu
                830
                                    835
                                                          840
Gly Leu Gly Ser Ala Leu Leu Ser Ser Leu Gly Glu His Ala Phe
                845
                                     850
Phe Arg Leu Ala Leu Pro Arg Ile Arg Lys Gly Ser Arg Leu Gln
                860
                                     865
                                                          870
Tyr Trp Leu His Thr Ser Gln Lys Ile His Arg Ala Leu Asn Thr
                875
                                    880
                                                          885
Glu Pro Pro Glu Gly Ser Lys Glu Glu Thr Ala Glu Ala Glu Pro
                890
                                    895
                                                          900
Ser Gly Pro Glu Val Glu Gln Gln Gln Gln Gln Asp Gln Pro
                905
                                    910
Thr Ala Pro Glu Gly Trp Lys Arg Ala Arg Arg Ala Val Asp Lys
                920
                                    925
                                                         930
Glu Arg Arg Val Arg Phe Leu Leu Glu Pro Ala Val Val Ala
                935
                                    940
                                                         945
Pro Glu Ala Asp Ala Glu Ala Glu Ala Ala Pro Arg Glu Gly Pro
                950
                                    955
                                                         960
Val Trp Leu Cys Ser Tyr Gly Arg Pro Pro Ala Ala Arg Pro Thr
                965
                                    970
                                                         975
Gly Ala Pro Gln Pro Gly Glu Leu Gln Glu Leu Glu Arg Arg Ile
                980
                                    985
                                                         990
Glu Val Ala Arg Glu Arg Leu Arg Gln Ala Leu Val Arg Arg Gly
                995
                                   1000
                                                        1005
Gln Leu Leu Ala Gln Leu Gly Asp Ser Ala Arg His Arg Pro Arg
               1010
                                   1015
                                                        1020
Arg Leu Leu Gln Ala Arg Ala Ala Pro Ala Glu Ala Pro Pro His
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Ser Gly Arg Pro Gly Ser Gln Glu
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Ser Ala Ser Arg Val Ala Gly Ile Thr Gly Phe His His Val Ala
                440
                                     445
                                                         450
Gln Ala His Leu Gly Leu Val Gly Ser Arg Asn Ser Ala Ala Phe
                                     460
Ser Leu Pro Thr Cys Trp Asp Tyr Arg Lys Pro Val Val Met Pro
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                                     475
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Asp Thr Pro Gly Ser Arg Glu Trp Asp Ile Leu Glu Thr Glu Glu
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His Tyr Lys Ser Arg Trp Arg Ser Ile Arg Ile Leu Tyr Leu Thr
                                                           45
                                      40
Met Phe Leu Ser Ser Val Gly Phe Ser Val Val Met Met Ser Ile
                                                           60
                 50
                                      55
Trp Pro Tyr Leu Gln Lys Ile Asp Pro Thr Ala Asp Thr Ser Phe
                                                           75
                 65
                                      70
Leu Gly Trp Val Ile Ala Ser Tyr Ser Leu Gly Gln Met Val Ala
                 80
                                      85
                                                           90
Ser Pro Ile Phe Gly Leu Trp Ser Asn Tyr Arg Pro Arg Lys Glu
                 95
                                     100
                                                         105
Pro Leu Ile Val Ser Ile Leu Ile Ser Val Ala Ala Asn Cys Leu
                110
                                     115
                                                         120
Tyr Ala Tyr Leu His Ile Pro Ala Ser His Asn Lys Tyr Tyr Met
                125
                                     130
                                                         135
Leu Val Ala Arg Gly Leu Leu Gly Ile Gly Ala Gly Asn Val Ala
                140
                                     145
                                                          150
Val Val Arg Ser Tyr Thr Ala Gly Ala Thr Ser Leu Gln Glu Arg
                155
                                     160
                                                          165
Thr Ser Ser Met Ala Asn Ile Ser Met Cys Gln Ala Leu Gly Phe
                170
                                     175
Ile Leu Gly Pro Val Phe Gln Thr Cys Phe Thr Phe Leu Gly Glu
                185
                                     190
                                                         195
Lys Gly Val Thr Trp Asp Val Ile Lys Leu Gln Ile Asn Met Tyr
                200
                                     205
                                                          210
Thr Thr Pro Val Leu Leu Ser Ala Phe Leu Gly Ile Leu Asn Ile
                215
                                     220
Ile Leu Ile Leu Ala Ile Leu Arg Glu His Arg Val Asp Asp Ser
                                                          240
                230
                                     235
Gly Arg Gln Cys Lys Ser Ile Asn Phe Glu Glu Ala Ser Thr Asp
                245
                                     250
                                                          255
Glu Ala Gln Val Pro Gln Gly Asn Ile Asp Gln Val Ala Val Val
                                     265
                260
Ala Ile Asn Val Leu Phe Phe Val Thr Leu Phe Ile Phe Ala Leu
                275
                                     280
                                                          285
Phe Glu Thr Ile Ile Thr Pro Leu Thr Met Asp Met Tyr Ala Trp
                                     295
                290
                                                          300
Thr Gln Glu Gln Ala Val Leu Tyr Asn Gly Ile Ile Leu Ala Ala
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Leu Gly Val Glu Ala Val Val Ile Phe Leu Gly Val Lys Leu Leu
Ser Lys Lys Ile Gly Glu Arg Ala Ile Leu Leu Gly Gly Leu Ile
                 335
                                      340
Val Val Trp Val Gly Phe Phe Ile Leu Leu Pro Trp Gly Asn Gln
                 350
                                     355
Phe Pro Lys Ile Gln Trp Glu Asp Leu His Asn Asn Ser Ile Pro
                 365
                                     370
Asn Thr Thr Phe Gly Glu Ile Ile Ile Gly Leu Trp Lys Ser Pro
                 380
                                     385
Met Glu Asp Asp Asn Glu Arg Pro Thr Gly Cys Ser Ile Glu Gln
                 395
                                     400
Ala Trp Cys Leu Tyr Thr Pro Val Ile His Leu Ala Gln Phe Leu
                 410
                                     415
Thr Ser Ala Val Leu Ile Gly Leu Gly Tyr Pro Val Cys Asn Leu
                 425
                                     430
                                                          435
Met Ser Tyr Thr Leu Tyr Ser Lys Ile Leu Gly Pro Lys Pro Gln
                 440
                                     445
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Gly Val Tyr Met Gly Trp Leu Thr Ala Ser Gly Ser Gly Ala Arg
                 455
                                     460
Ile Leu Gly Pro Met Phe Ile Ser Gln Val Tyr Ala His Trp Gly
                                     475
                                                          480
Pro Arg Trp Ala Phe Ser Leu Val Cys Gly Ile Ile Val Leu Thr
                 485
                                     490
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Ile Thr Leu Leu Gly Val Val Tyr Lys Arg Leu Ile Ala Leu Ser
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Val Arg Tyr Gly Arg Ile Gln Glu
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Leu Ile Tyr Ala Cys Tyr His Met Ser Arg Lys Pro Ile Ser Ile
                  35
                                      40
Val Lys Ser Arg Leu His Gln Asn Cys Ser Glu Gln Ile Lys Pro
                 50
                                      55
Ile Asn Asp Thr His Ser Leu Asn Asp Thr Met Trp Cys Ser Trp
                 65
                                      70
Ala Pro Phe Asp Lys Asp Asn Tyr Lys Glu Leu Leu Gly Gly Val
                 80
                                      85
Asp Asn Ala Phe Leu Ile Ala Tyr Ala Ile Gly Met Phe Ile Ser
                 95
                                     100
                                                          105
Gly Val Phe Gly Glu Arg Leu Pro Leu Arg Tyr Tyr Leu Ser Ala
                110
                                     115
                                                          120
Gly Met Leu Leu Ser Gly Leu Phe Thr Ser Leu Phe Gly Leu Gly
                125
                                     130
                                                          135
Tyr Phe Trp Asn Ile His Glu Leu Trp Tyr Phe Val Val Ile Gln
                140
                                     145
Val Cys Asn Gly Leu Val Gln Thr Thr Gly Trp Pro Ser Val Val
                155
                                     160
                                                          165
Thr Cys Val Gly Asn Trp Phe Gly Lys Gly Lys Arg Gly Phe Ile
                170
                                                          180
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Pro Gln His His Gly Glu Pro Ala Glu Asn Gln Asp Asn Pro Glu
                245
                                     250
Asp Pro Gly Asn Ser Pro Cys Ser Ile Arg Glu Ser Gly Leu Glu
                260
                                     265
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Thr Val Ala Lys Cys Ser Lys Gly Pro Cys Glu Glu Pro Ala Ala
                275
                                     280
                                                          285
Ile Ser Phe Phe Gly Ala Leu Arg Ile Pro Gly Val Val Glu Phe
                290
                                     295
Ser Leu Cys Leu Leu Phe Ala Lys Leu Val Ser Tyr Thr Phe Leu
                305
                                    310
                                                          315
Tyr Trp Leu Pro Leu Tyr Ile Ala Asn Val Ala His Phe Ser Ala
                320
                                    325
                                                          330
Lys Glu Ala Gly Asp Leu Ser Thr Leu Phe Asp Val Gly Gly Ile
                                     340
                                                          345
Ile Gly Gly Ile Val Ala Gly Leu Val Ser Asp Tyr Thr Asn Gly
                350
                                    355
Arg Ala Thr Thr Cys Cys Val Met Leu Ile Leu Ala Ala Pro Met
                365
                                    370
                                                          375
Met Phe Leu Tyr Asn Tyr Ile Gly Gln Asp Gly Ile Ala Ser Ser
                380
                                    385
                                                          390
Ile Val Met Leu Ile Ile Cys Gly Gly Leu Val Asn Gly Pro Tyr
                395
                                    400
                                                          405
Ala Leu Ile Thr Thr Ala Val Ser Ala Asp Leu Gly Thr His Lys
                410
                                    415
                                                          420
Ser Leu Lys Gly Asn Ala Lys Ala Leu Ser Thr Val Thr Ala Ile
                                     430
                                                          435
Ile Asp Gly Thr Gly Ser Ile Gly Ala Ala Leu Gly Pro Leu Leu
                440
                                    445
                                                          450
Ala Gly Leu Ile Ser Pro Thr Gly Trp Asn Asn Val Phe Tyr Met
                455
                                    460
Leu Ile Ser Ala Asp Val Leu Ala Cys Leu Leu Cys Arg Leu
                470
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Val Tyr Lys Glu Ile Leu Ala Trp Lys Val Ser Leu Ser Arg Gly
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Ser Gly Tyr Lys Glu Ile
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Val Leu Leu His Thr Ser Arg Lys Tyr Leu Lys Leu Lys Asn Phe
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Lys Glu Glu Ile Arg Ala His Arg Asp Leu Asp Gly Phe Leu Ala
                 35
                                     40
Gln Ala Ser Ile Val Leu Asn Glu Thr Ala Thr Ser Leu Asp Asn
                 50
                                     55
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Met Gly Ile Trp Asn Ser His Thr Ser Val Gly Asn Ile Leu Gly

Ser Leu Ile Ala Gly Ile Trp Val Asn Gly Gln Trp Gly Leu Ser

Phe Ile Val Pro Gly Ile Ile Thr Ala Val Met Gly Val Ile Thr

Phe Leu Phe Leu Ile Glu His Pro Glu Asp Val Asp Cys Ala Pro

Val Leu Arg Thr Met Leu Arg Arg Phe Ala Arg Asp Pro Asp Asn Asn Glu Pro Asn Cys Asn Leu Asp Leu Leu Met Ala Met Leu Phe Thr Asp Ala Gly Ala Pro Met Arg Gly Lys Val His Leu Leu Ser Asp Thr Ile Gln Gly Val Thr Ala Thr Val Thr Gly Val Arg Tyr Gln Gln Ser Trp Leu Cys Ile Ile Cys Thr Met Lys Ala Leu Gln Lys Arg His Val Cys Ile Ser Arg Leu Val Arg Pro Gln Asn Trp Gly Glu Asn Ser Cys Glu Val Arg Phe Val Ile Leu Val Leu Ala Pro Pro Lys Met Lys Ser Thr Lys Thr Ala Met Glu Val Ala Arg Thr Phe Ala Thr Met Phe Ser Asp Ile Ala Phe Arg Gln Lys Leu Leu Glu Thr Arg Thr Glu Glu Glu Phe Lys Glu Ala Leu Val His Gln Arg Gln Leu Leu Thr Met Val Ser His Gly Pro Val Ala Pro Arg Thr Lys Glu Arg Ser Thr Val Ser Leu Pro Ala His Arg His Pro Glu Pro Pro Lys Cys Lys Asp Phe Val Pro Phe Gly Lys Gly Ile Arg Glu Asp Ile Ala Arg Arg Phe Pro Leu Tyr Pro Leu Asp Phe Thr Asp Gly Ile Ile Gly Lys Asn Lys Ala Val Gly Lys Tyr Ile Thr Thr Leu Phe Leu Tyr Phe Ala Cys Leu Leu Pro Thr Ile Ala Phe Gly Ser Leu Asn Asp Glu Asn Thr Asp Gly Ala Ile Asp Val Gln Lys Thr Ile Ala Gly Gln Ser Ile Gly Gly Leu Leu Tyr Ala Leu Phe Ser Gly Gln Pro Leu Val Ile Leu Leu Thr Thr Ala Pro Leu Ala Leu Tyr Ile Gln Val Ile Arg Val Ile Cys Asp Asp Tyr Asp Leu Asp Phe Asn Ser Phe Tyr Ala Trp Thr Gly Leu Trp Asn Ser Phe Phe Leu Ala Leu Tyr Ala Phe Phe Asn Leu Ser Leu Val Met Ser Leu Phe Lys Arg Ser Thr Glu Glu Ile Ile Ala Leu Phe Ile Ser Ile Thr Phe Val Leu Asp Ala Val Lys Gly Thr Val Lys Ile Phe Trp Lys Tyr Tyr Tyr Gly His Tyr Leu Asp Asp Tyr His Thr Lys Arg Thr Ser Ser Leu Val Ser Leu Ser Gly Leu Gly Ala Ser Leu Asn Ala Ser Leu His Thr Ala Leu Asn Ala Ser Phe Leu Ala Ser Pro Thr Glu Leu Pro Ser Ala Thr His Ser Gly Gln Ala Thr Ala Val Leu Ser Leu Leu Ile Met Leu Gly Thr Leu Trp Leu Gly Tyr Thr Leu Tyr Gln Phe Lys Lys Ser Pro Tyr Leu His Pro Cys Val Arg Glu Ile Leu Ser Asp Cys Ala Leu Pro Ile Ala Val Leu Ala Phe Ser Leu Ile Ser Ser His Gly Phe Arg Glu

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Ile Glu Met Ser Lys Phe Arg Tyr Asn Pro Ser Glu Ser Pro Phe
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                                    550
Ala Met Ala Gln Ile Gln Ser Leu Ser Leu Arg Ala Val Ser Gly
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                                    565
                                                         570
Ala Met Gly Leu Gly Phe Leu Leu Ser Met Leu Phe Phe Ile Glu
                575
                                    580
                                                         585
Gln Asn Leu Val Ala Ala Leu Val Asn Ala Pro Glu Asn Arg Leu
                590
                                    595
                                                         600
Val Lys Gly Thr Ala Tyr His Trp Asp Leu Leu Leu Leu Ala Ile
                605
                                    610
                                                         615
Ile Asn Thr Gly Leu Ser Leu Phe Gly Leu Pro Trp Ile His Ala
                620
                                    625
Ala Tyr Pro His Ser Pro Leu His Val Arg Ala Leu Ala Leu Val
                                    640
                635
                                                         645
Glu Glu Arg Val Glu Asn Gly His Ile Tyr Asp Thr Ile Val Asn
                650
                                    655
                                                         660
Val Lys Glu Thr Arg Leu Thr Ser Leu Gly Ala Ser Val Leu Val
                665
                                    670
                                                         675
Gly Leu Ser Leu Leu Leu Pro Val Pro Leu Gln Trp Ile Pro
                                                         690
                680
                                    685
Lys Pro Val Leu Tyr Gly Leu Phe Leu Tyr Ile Ala Leu Thr Ser
                695
                                    700
                                                         705
Leu Asp Gly Asn Gln Leu Val Gln Arg Val Ala Leu Leu Lys
                                                         720
                710
                                    715
Glu Gln Thr Ala Tyr Pro Pro Thr His Tyr Ile Arg Arg Val Pro
                725
                                    730
                                                         735
Gln Arg Lys Ile His Tyr Phe Thr Gly Leu Gln Val Leu Gln Leu
                740
                                    745
                                                         750
Leu Leu Cys Ala Phe Gly Met Ser Ser Leu Pro Tyr Met Lys
                755
                                    760
                                                         765
Met Ile Phe Pro Leu Ile Met Ile Ala Met Ile Pro Ile Arg Tyr
                                                         780
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Ile Leu Leu Pro Arg Ile Ile Glu Ala Lys Tyr Leu Asp Val Met
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Asp Ala Glu His Arg Pro
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Leu Ser Thr Ala Tyr Leu Gly Leu Val Leu Leu Gly Glu Gln His
                 35
                                      40
Leu Thr Ala Ala Ala Thr Phe Ile Tyr Phe Tyr Leu Thr Thr
                 50
                                     55
                                                          60
Leu Thr Val Gly Tyr Gly Asp Leu Ala Pro Gln Thr Ser Ala Gly
                 65
                                     70
                                                          75
Arg Ile Phe Val Ala Ala Trp Val Met Leu Gly Gly Ile Ala Leu
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                                      85
Leu Thr Ala Ala Ile Gly Lys Thr Thr Ser Ser Val Ile Asp Ala
                                    100
Trp Arg Lys Gly Met Lys Gly Lys Gly Asp Phe Thr Gly Lys Val
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110
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Gly His Thr Val Leu Ile Gly Trp. Glu Gly Ala Ser Ser Glu Arg
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                                     130
                                                          135
Val Ile Glu Leu Leu Gln Asp Glu Thr Ser Asn Asp Asn Leu
                 140
                                      145
                                                          150
Ile Val Ile Cys Asp Cys Ser Leu Glu Glu Asn Pro Met Pro Gly
                 155
                                     160
Lys Ala Ala Phe Ile Arg Gly Glu Ser Leu Ser Ser Thr Ala Leu
                 170
                                     175
Leu Leu Arg Ala Gly Val Pro Gly Ala Glu Arg Val Leu Val Arg
                 185
                                     190
Thr Pro Ser Asp Asp Leu Thr Leu Ala Thr Val Leu Ala Val Asn
                 200
                                     205
                                                          210
Gln Leu Ser Pro Val Gly His Val Val Ala His Phe Asn Glu Ser
                 215
                                     220
                                                          225
Glu Ile Ala Ala Leu Ala Ser Ser Tyr Ala Pro Arg Leu Glu Cys
                 230
                                     235
Thr Ser Ser Met Ala Ile Glu Met Leu Val Arg Ala Ser Gln Asp
                 245
                                     250
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Pro Gly Ser Ser Val Val Ile Asn Glu Leu Leu Cys Val Gly Gln
                 260
                                     265
                                                          270
Gly Ala Thr Gln Tyr Leu Met Lys Leu Pro Glu Ala Phe Glu Ala
                 275
                                     280
                                                          285
Thr Phe Gly Glu Leu Tyr Thr Gln Met Lys Glu Arg His Asn Ala
                290
                                                          300
Thr Leu Ile Gly Tyr Arg Ala Lys Gly Val Gln Gln Pro Ser Ile
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Asn Pro Pro Ser Ala Thr Glu Val Lys Gly Gly Glu Leu Phe Tyr
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                                     325
Ile Ala Ser Thr Arg Leu Lys Glu Ile Ser His Gly Met Ala
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Asp Tyr Val His Val Phe Tyr Thr Ile Ser Gln Ile Glu Thr Asp
                 35
                                      40
Gly Ile Asn Tyr Leu Val Asp Asp Phe Ala Asn Ala Ser Gly Thr
                 50
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Ile Thr Phe Leu Pro Trp Gln Arg Ser Glu Val Leu Asn Ile Tyr
                 65
                                      70
                                                          75
Val Leu Asp Asp Asp Ile Pro Glu Leu Asn Glu Tyr Phe Arg Val
                 80
                                      85
Thr Leu Val Ser Ala Ile Pro Gly Asp Gly Lys Leu Gly Ser Thr
                 95
                                     100
Pro Thr Ser Gly Ala Ser Ile Asp Pro Glu Lys Glu Thr Thr Asp
                110
                                     115
                                                         120
Ile Thr Ile Lys Ala Ser Asp His Pro Tyr Gly Leu Leu Gln Phe
                125
                                     130
                                                         135
Ser Thr Gly Leu Pro Pro Gln Pro Lys Asp Ala Met Thr Leu Pro
                140
                                     145
Ala Ser Ser Val Pro His Ile Thr Val Glu Glu Glu Asp Gly Glu
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Asp Asp Ile Pro Glu Glu Lys Ser Phe Tyr Glu Phe Gln Leu Thr
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